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(54) Title: **P-SELECTIN LIGAND PROTEINS**

		Selectin Binding		
		P	E	L
YYY	↑ <sup>sLe<sup>x</sup></sup>			
YYD 18.Fo		+++	+	-
FYY	↑ <sup>sLe<sup>x</sup></sup>			
FYYD 19.Fo		++	+	-
FFY	↑ <sup>sLe<sup>x</sup></sup>			
FFYD 19.Fo		+	+	-
FFF	↑ <sup>sLe<sup>x</sup></sup>			
FFFD 19.Fo		+	+	-
YYY N18				
FFYDN18.Fo		-	-	-

(57) Abstract

A novel P-selectin ligand glycoprotein is disclosed, comprising the amino acid sequence set forth in SEQ ID NO 2 or by the amino acid sequence set forth in SEQ ID NO 4. DNA sequences encoding the P-selectin ligand protein are also disclosed, along with vectors, host cells, and methods of making the P-selectin ligand protein. Pharmaceutical compositions containing the P-selectin ligand protein and methods of treating inflammatory disease states characterized by P-selectin- and E-selectin-mediated intercellular adhesion are also disclosed.

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**P-SELECTIN LIGAND PROTEINS**

This application is a continuation-in-part of copending application U.S. Serial No. 08/428,734, filed April 25, 1995, which was a continuation-in-part of copending application U.S. Serial No. 08/316,305, filed September 30, 1994, which was a continuation-in-part of copending application U.S. Serial No. 08/235,398, filed April 28, 1994, which was a continuation-in-part of copending application U.S. Serial No. 08/112,608, filed August 26, 1993, which was a continuation-in-part of U.S. Serial No. 07/965,662, filed October 23, 1992, now abandoned. This application also claims priority from International Application No. PCT/US93/10168, filed October 22, 1993.

15

**BACKGROUND OF THE INVENTION**

The present invention relates to the field of anti-inflammatory substances which act by inhibiting leukocyte adhesion to endothelial cells. More particularly, the present invention is directed to novel ligands for the mammalian adhesion proteins known as selectins.

20

During inflammation leukocytes adhere to the vascular endothelium and enter subendothelial tissue, an interaction which is mediated by specific binding of the selectin or LEC-CAM class of proteins to ligands on target cells. Such selectin-mediated cellular adhesion also occurs in thrombotic disorders and parasitic diseases and may be implicated in metastatic spread of tumor cells.

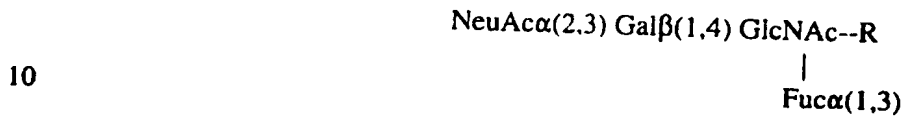
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The selectin proteins are characterized by a N-terminal lectin-like domain, an epidermal growth factor-like domain, and regions of homology to complement binding proteins. Thus far three human selectin proteins have been identified, E-selectin (formerly ELAM-1), L-selectin (formerly LAM-1) and P-selectin (formerly PADGEM or GMP-140). E-selectin is induced on endothelial cells several hours after activation by cytokines, mediating the calcium-dependent interaction between neutrophils and the endothelium. L-selectin is the lymphocyte homing receptor, and P-selectin rapidly appears on the cell surface of platelets when they are activated, mediating calcium-dependent adhesion of neutrophils or monocytes to platelets. P-selectin is also found in the Weibel-Palade bodies of endothelial cells; upon its release from these vesicles P-selectin mediates early binding of neutrophils to histamine- or thrombin-stimulated endothelium.

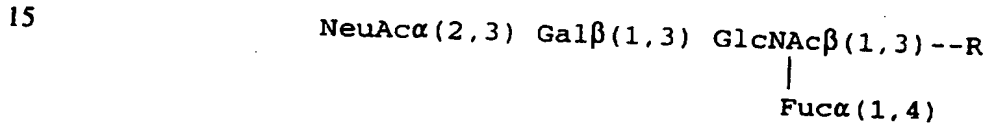
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Selectins are believed to mediate adhesion through specific interactions with ligands present on the surface of target cells. Generally the ligands of selectins are comprised at least in

5 part of a carbohydrate moiety. For example, E-selectin binds to carbohydrates having the terminal structure

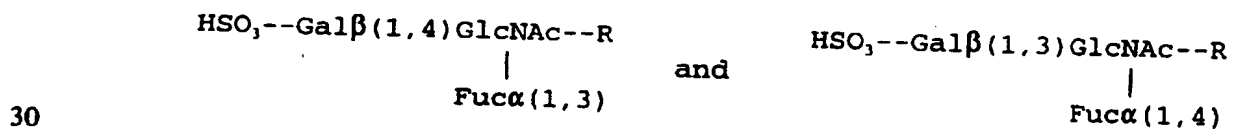


and also to carbohydrates having the terminal structure



20 where R =the remainder of the carbohydrate chain. These carbohydrates are known blood group antigens and are commonly referred to as sialyl Lewis<sup>x</sup> and sialyl Lewis<sup>y</sup>, respectively. The presence of the sialyl Lewis<sup>x</sup> antigen alone on the surface of an endothelial cell may be sufficient to promote binding to an E-selectin expressing cell. E-selectin also binds to carbohydrates having

25 the terminal structures



35 As with E-selectin, each selectin appears to bind to a range of carbohydrates with varying affinities. The strength of the selectin mediated adhesive event (binding affinity) may also depend on the density of the carbohydrate and on the density of the selectin on the cell surface.

P-selectin binds to carbohydrates containing the non-sialated form of the Lewis<sup>x</sup> blood group antigen and with higher affinity to sialyl Lewis<sup>x</sup>. P-selectin may also recognize sulfatides, which are heterogeneous 3-sulfated galactosyl ceramides, isolated from myeloid and tumor cells

40 by lipid extraction. However, the binding of cells bearing P-selectin to cells bearing P-selectin ligands is abolished when the ligand-bearing cells are treated with proteases, indicating that the P-selectin ligand may be a glycoprotein.

5 Two putative glycoprotein ligands for P-selectin have recently been identified, one of which has been partially purified, (Moore *et al.*, J. Cell Biol. 118, 445-456 (1992)). However, neither amino acid composition nor the amino acid sequence of these glycoproteins are disclosed.

### SUMMARY OF THE INVENTION

10 In one embodiment, the present invention provides a composition comprising an isolated DNA encoding a P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402. Also provided is a composition comprising an isolated DNA encoding a soluble P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 310. The  
15 invention further provides a composition comprising an isolated DNA encoding a mature P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402. In another embodiment, the invention provides a composition comprising an isolated DNA encoding a soluble mature P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42  
20 to amino acid 310. In another embodiment, the invention provides a composition comprising an isolated DNA encoding a P-selectin ligand protein, said protein comprising the amino acid sequence set forth in SEQ ID NO:4. The invention further provides a composition comprising an expression vector comprising any one of the isolated DNAs of the invention, said DNA being operably linked to an expression control sequence; a host cell transformed with the expression  
25 vector containing any one of the DNAs described above; and a process for producing the P-selectin ligand protein, which comprises:

(a) culturing a host cell transformed with an expression vector containing any one of the DNAs of the invention in a suitable culture medium; and

(b) purifying the P-selectin ligand protein from the culture medium.

30 In another embodiment, the invention provides a composition comprising a protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 21 to amino acid 402, said protein being substantially free from other mammalian proteins. The invention further comprises a soluble P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 21 to amino acid 310, said protein being substantially free from other  
35 mammalian proteins. In another embodiment, the invention comprises a P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402, said protein being substantially free from other mammalian proteins. The invention also provides a composition comprising a mature P-selectin ligand protein comprising the amino acid

5 sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402, said protein being substantially free from other mammalian proteins. Further provided is a composition comprising a soluble mature P-selectin ligand protein comprising the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310, said protein being substantially free from other mammalian proteins. In another embodiment, the invention provides a composition comprising  
10 a protein comprising the amino acid sequence set forth in SEQ ID NO:4.

In yet another embodiment, the invention provides compositions comprising antibodies specific for P-selectin ligand proteins.

In another embodiment, the invention provides a method of identifying an inhibitor of P-selectin-mediated intercellular adhesion which comprises:

15 (a) combining a P-selectin protein with a P-selectin ligand protein comprising an amino acid sequence selected from the group consisting of the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402, the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402, the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310, and the amino acid sequence set forth in SEQ ID  
20 NO:4, said combination forming a first binding mixture;

(b) measuring the amount of binding between the P-selectin protein and the P-selectin ligand protein in the first binding mixture;

(c) combining a compound with the P-selectin protein and the P-selectin ligand protein to form a second binding mixture;

25 (d) measuring the amount of binding in the second binding mixture; and

(e) comparing the amount of binding in the first binding mixture with the amount of binding in the second binding mixture;

wherein the compound is capable of inhibiting P-selectin-mediated intercellular adhesion when a decrease in the amount of binding of the second binding mixture occurs.

30 In another embodiment, the invention provides a method of identifying an inhibitor of E-selectin-mediated intercellular adhesion which comprises:

(a) combining a E-selectin protein with a P-selectin ligand protein comprising an amino acid sequence selected from the group consisting of the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402, the amino acid sequence set forth in SEQ  
35 ID NO:2 from amino acid 42 to amino acid 402, the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310, and the amino acid sequence set forth in SEQ ID NO:4, said combination forming a first binding mixture;

- 5 (b) measuring the amount of binding between the E-selectin protein and the P-selectin ligand protein in the first binding mixture;
- (c) combining a compound with the E-selectin protein and the P-selectin ligand protein to form a second binding mixture;
- (d) measuring the amount of binding in the second binding mixture; and
- 10 (e) comparing the amount of binding in the first binding mixture with the amount of binding in the second binding mixture;

wherein the compound is capable of inhibiting E-selectin-mediated intercellular adhesion when a decrease in the amount of binding of the second binding mixture occurs. These methods may also be used to look for L-selectin inhibitors by substituting L-selectin for E-selectin.

15 The invention also encompasses processes for producing P-selectin ligand proteins which comprise (a) co-transforming a host cell with a DNA encoding a P-selectin ligand protein and a DNA encoding a fucosyltransferase capable of synthesizing sialyl Lewis X (sLe<sup>x</sup>) or sialyl Lewis A (sLe<sup>a</sup>) (such as an ( $\alpha$ 1,3/ $\alpha$ 1,4) fucosyltransferase or an ( $\alpha$ 1, 3) fucosyltransferase), each of said DNAs being operably linked to an expression control sequence; (b) culturing the host cell in

20 suitable culture medium; and (c) purifying the P-selectin ligand protein from the culture medium. In certain other embodiments, the host cell is also co-transformed with a DNA encoding a paired basic amino acid converting enzyme and/or a DNA encoding a GlcNAc transferase (preferably a "core2 transferase"). In preferred embodiments, the P-selectin ligand protein is a full-length or soluble form.

25 In other embodiments, the present invention includes a P-selectin ligand protein having P-selectin ligand protein activity. In preferred embodiments, the ligand protein is a protein comprising the sequence from amino acid 42 to amino acid 60 of SEQ ID NO: 2, consisting essentially of the sequence from amino acid 42 to amino acid 60 of SEQ ID NO: 2, comprising the sequence from amino acid 42 to amino acid 88 of SEQ ID NO: 2, consisting essentially of the

30 sequence from amino acid 42 to amino acid 88 of SEQ ID NO: 2, consisting essentially of the sequence from amino acid 42 to amino acid 118 of SEQ ID NO: 2, or consisting essentially of the sequence from amino acid 42 to amino acid 189 of SEQ ID NO: 2. In other preferred embodiments, at least one of the asparagine residues at positions 65, 111 and 292 of SEQ ID NO: 2 have been deleted or replaced. Certain preferred embodiments of the ligand protein comprises

35 at least one of the tyrosine residues at positions 46, 48 and 51 of SEQ ID NO: 2. DNAs encoding these P-selectin ligand proteins, host cells transformed with such DNAs, process for producing protein by culturing such host cells, pharmaceutical compositions comprising the proteins, methods of identifying selectin binding inhibitors using the proteins, antibodies to the proteins

5 and methods of inhibiting selectin mediated binding using the proteins are also encompassed by the invention.

In yet other embodiments the invention provides an isolated DNA encoding a fusion protein comprising (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second amino acid sequence derived from the  
10 sequence of a protein other than P-selectin ligand. Preferably, an expression control sequence is operably linked to the nucleotide sequence. Host cells transformed with such DNAs are also provided. The invention also provides a process for producing a fusion protein, which comprises: (a) culturing the host cell under condition suitable for expression of the fusion protein; and (b) purifying the fusion protein from the culture  
15 medium. Fusion proteins produced according to such process are also provide.

In certain preferred embodiments, the first amino acid sequence of such fusion protein comprises amino acid 42 to amino acid 402 of SEQ ID NO:2, amino acid 42 to amino acid 310 of SEQ ID NO:2, amino acid 42 to amino acid 88 of SEQ ID NO:2, amino acid 42 to amino acid 118 of SEQ ID NO:2, or amino acid 42 to amino acid 189  
20 of SEQ ID NO:2.

In other preferred embodiments, the DNA comprises the nucleotide sequence of SEQ ID NO:35 from nucleotide 123 to nucleotide 939, the nucleotide sequence of SEQ ID NO:35, the nucleotide sequence of SEQ ID NO:37 from nucleotide 123 to nucleotide 807, the nucleotide sequence of SEQ ID NO:37, the nucleotide sequence of SEQ ID  
25 NO:39 from nucleotide 123 to nucleotide 1311, the nucleotide sequence of SEQ ID NO:39, the nucleotide sequence of SEQ ID NO:41 from nucleotide 123 to nucleotide 792, or the nucleotide sequence of SEQ ID NO:41.

The present invention also provides a fusion protein comprising (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a  
30 second amino acid sequence derived from the sequence of a protein other than P-selectin ligand. Preferably, the first amino acid sequence comprises amino acid 42 to amino acid 402 of SEQ ID NO:2, amino acid sequence comprises amino acid 42 to amino acid 310 of SEQ ID NO:2, amino acid 42 to amino acid 88 of SEQ ID NO:2, amino acid 42 to amino acid 118 of SEQ ID NO:2, or amino acid 42 to amino acid 189 of SEQ ID NO:2.

35 In certain particularly preferred embodiments, the fusion protein comprises the amino acid sequence of SEQ ID NO:36 from amino acid 42 to amino acid 313, the



5 amino acid sequence of SEQ ID NO:36, the amino acid sequence of SEQ ID NO:38 from amino acid 42 to amino acid 269, the amino acid sequence of SEQ ID NO:38. the amino acid sequence of SEQ ID NO:40 from amino acid 42 to amino acid 437. the amino acid sequence of SEQ ID NO:40, the amino acid sequence of SEQ ID NO:42 from amino acid 42 to amino acid 264, or the amino acid sequence of SEQ ID NO:42.

10 In other preferred embodiments, the second amino acid sequence is linked to the C-terminus or the N-terminus of the first amino acid sequence, with or without being linked by a linking sequence.

In yet other embodiments, the second amino acid sequence is derived from a protein selected from the group consisting of an antibody, a cytokine, a growth factor, a  
15 differentiation factor, a hormone, an enzyme, a receptor or fragment thereof and a ligand. Preferably, the second amino acid sequence is derived from the sequence of an antibody, from the Fc portion of an antibody, or is a mutation of a sequence derived from an antibody.

In yet further embodiments, the present invention provides for a composition  
20 comprising (a) a first peptide comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second peptide derived from the sequence of a protein other than P-selectin ligand, wherein the first peptide and the second peptide are chemically linked by a moiety other than a peptide bond. Any P-selectin ligand protein of the invention can be used in such a composition.

### 25 BRIEF DESCRIPTION OF THE FIGURES

Fig. 1 is a graph comparing the binding of P-selectin ligand proteins expressed with and without core2.

Fig. 2 is an autoradiograph of immunoprecipitations of P-selectin ligand protein expressed with and without core2.

30 Fig. 3 depicts the results of flow cytometry analysis of the binding of P-selectin ligand protein (expressed with and without core2) to P-selectin/IgG chimera (LEC- $\gamma$ 1) and anti-P-selectin ligand protein monoclonal antibody (MAb 275).

Fig. 4 is an autoradiograph of proteins, including P-selectin ligand protein, which bound to P- and E-selectin/IgG chimeras.

35 Fig. 5 is a schematic representation of structural features of the full length P-selectin ligand protein of SEQ ID NO: 2.

5 Fig. 6 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the role of N-linked glycosylation sites in binding of the P-selectin ligand proteins to selectins.

Fig. 7 depicts the results of experiments to determine the role of N-linked glycosylation sites in binding of the P-selectin ligand proteins to selectins.

10 Fig. 8 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the role of sulfated tyrosine residues in binding of the P-selectin ligand proteins to selectins.

Figs. 9-11 depicts the results of experiments to determine the role of sulfated tyrosine residues in binding of the P-selectin ligand proteins to selectins.

15 Fig. 12 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the effects of various deletions on the binding of the P-selectin ligand proteins to selectins.

Fig. 13 is a schematic depiction of the quantitative plate binding assay of Example 4(c).

20 Figs. 14-17 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

Fig. 18 is a schematic representation of several P-selectin ligand protein fragments constructed for the purpose of examining the effects of alteration of tyrosine residues in the anionic region of the P-selectin ligand proteins on selectin binding.

25 Figs. 19-21 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

Fig. 22 depicts a proposed model for binding of P-selectin ligand proteins to P- and E-selectin.

Figs. 23 and 24 depict the results of experiments comparing the binding of various deleted and altered P-selectin ligand proteins to selectins.

30 Some of the foregoing figures employ a convention for numbering amino acids within the depicted constructs which is different that the residue numbering employed in SEQ ID NO:2. In the figures, residues are numbered using the first amino acid of soluble mature P-selectin ligand as a starting point. Hence, the residue numbers used in the figures are 41 less than those of SEQ ID NO:2. For example, residue 19 in the figures corresponds to residue 60 in SEQ ID NO:2.

35 Fig. 25 is an analysis of the expression products of CHO cells, already expressing 3/4 fucosyltransferase and Core2 transferase, which were transfected with psPSL.T7, ΔTM, I316 or psPSL.QC and amplified using methotrexate. Conditioned media was either analyzed directly or

5 first precipitated with LEC-γ1 and then analyzed by SDS-PAGE under non-reducing and reducing conditions.

Fig. 26. SDS-PAGE separation of myeloid cell membrane proteins affinity captured by P-and E-selectin. Membrane lysates were prepared from U937 cells metabolically labeled with <sup>3</sup>H-glucosamine and subjected to affinity precipitation with immobilized P-and E-selectin and control human IgG<sub>1</sub>. Eluted proteins were treated with ("reduced") or without ("non-reduced")  
10 DTT prior to gel electrophoresis. Lanes: 1, affinity capture by human IgG<sub>1</sub>; 2, affinity capture by P-selectin; 3, affinity capture by E-selectin.

Fig. 27. Sequential affinity capture experiments. <sup>3</sup>H-labeled U937 lysate species were affinity captured by P-or E-selectin, eluted, and then subjected to immunoprecipitation with anti-PSGL-1 antiserum Rb3443. Lanes: 1 and 2, control immunoprecipitations of fresh myeloid  
15 cell membrane lysates using pre-immune rabbit serum (lane 1) and Rb3443 (lane 2); 3-5, immunoprecipitation with Rb3443 of myeloid cell membrane lysates previously affinity captured and eluted from P-selectin (lane 3), E-selectin (lane 4), and human IgG<sub>1</sub> (lane 5).

Fig. 28. Comparison of CD43 and PSGL-1 content of myeloid cell membrane extracts. Labeled U937 cell extracts were immunoprecipitated with anti-PSGL-1 rabbit polyclonal antibody Rb3443 or an anti-CD43 mouse MAb and then subjected to SDS-PAGE/autoradiography. Lanes:  
20 1, immunoprecipitation with control pre-immune rabbit serum; 2, immunoprecipitation with Rb3443; 3, immunoprecipitation with control isotype-matched mouse antibody; 4, immunoprecipitation with anti-CD43 antibody.

Fig. 29. COS transfection experiments. COS M6 cells transfected with plasmids encoding PSGL-1 or CD43 as well as Fuc-TIII or Fuc-TVII were metabolically labeled with <sup>35</sup>S-methionine, and membranes were prepared for affinity capture experiments as described in Materials and Methods. The cDNAs employed in the transfections are indicated above the lanes. Precipitations were performed using (A) E-selectin, (B) P-selectin, and (C) anti-PSGL-1 antiserum  
25 Rb3443 and anti-CD43 MAb.  
30

Fig. 30 summarizes the results of screening of various P-selectin ligand proteins for inhibition of P- and E-selectin binding (see Example 13).

#### DETAILED DESCRIPTION OF THE INVENTION

The present inventors have for the first time identified and isolated a novel DNA which  
35 encodes a protein which acts as a ligand for P-selectin on human endothelial cells and platelets. The sequence of the DNA is set forth in SEQ ID NO:1. The complete amino acid sequence of the P-selectin ligand protein (i.e., the mature peptide plus the leader sequence) is characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402.



5 ID NO:2) is a homodimeric protein having an apparent molecular of 220 kD as shown by non-reducing SDS-polyacrylamide gel electrophoresis.

The structure of the full-length P-selectin ligand protein is schematically represented in Fig. 5. Three regions of the P-selectin ligand protein of SEQ ID NO:2 are: an extracellular domain (from about amino acid 21 to 310 of SEQ ID NO:2), a transmembrane domain (from  
10 about amino acid 311 to 332 of SEQ ID NO:2), and an intracellular, cytoplasmic domain (from about amino acid 333 to 402 of SEQ ID NO:2). The extracellular domain contains three consensus tripeptide sites (Asn-X-Ser/Thr) of potential N-linked glycosylation beginning at Asn residues 65, 111, and 292. The extracellular domain further contains three potential sites of tyrosine sulfation at residues 46, 48, and 51. The region comprised of residues 55-267 contains  
15 a high percentage of proline, serine, and threonine including a subdomain of fifteen decameric repeats of the ten amino acid consensus sequence Ala-Thr/Met-Glu-Ala-Gln-Thr-Thr-X-Pro/Leu-Ala/Thr, wherein X can be either Pro, Ala, Gln, Glu, or Arg. Regions such as these are characteristic of highly O-glycosylated proteins.

COS or CHO cells co-transfected with a gene encoding the P-selectin ligand protein and  
20 a gene encoding fucosyltransferase (hereinafter FT), preferably an ( $\alpha$ 1,3/ $\alpha$ 1,4) fucosyltransferase ("3/4FT"), are capable of binding to CHO cells expressing P-selectin on their surface, but are not capable of binding to CHO cells which do not express P-selectin on their surface. In order to bind to P-selectin, either in purified form or expressed on the surface of CHO cells, the gene encoding the P-selectin ligand protein must be co-transfected with the gene encoding an FT, since  
25 transfection of either gene in the absence of the other either abolishes or substantially reduces the P-selectin binding activity. The binding of the P-selectin ligand protein of the invention to P-selectin can be inhibited by EDTA or by a neutralizing monoclonal antibody specific for P-selectin. The binding of the P-selectin ligand protein of the invention to P-selectin is not inhibited by a non-neutralizing monoclonal antibody specific for P-selectin or by an isotype control. These  
30 results characterize the binding specificity of the P-selectin ligand protein of the invention.

For the purposes of the present invention, a protein is defined as having "P-selectin ligand protein activity", i.e., variably referred to herein as a "P-selectin ligand protein", or as a "P-selectin ligand glycoprotein" or simply as a "P-selectin ligand", when it binds in a calcium-dependent  
35 manner to P-selectin which is present on the surface of cells as in the CHO-P-selectin binding assay of Example 4, or to P-selectin which is affixed to another surface, for example, as the chimeric P-selectin-IgGy1 protein of Example 4 is affixed to Petri dishes.

The glycosylation state of the P-selectin ligand protein of the invention was studied using a chimeric, soluble form of the P-selectin ligand protein, described in detail in Example 5(C) and



5 by the present invention. Such fragments comprise amino acids 21 to 54 of SEQ ID NO:2, a region of the P-selectin ligand protein having a low frequency of serine and threonine residues; amino acids 55 to 127 of SEQ ID NO:2, having a high frequency of proline, serine, and threonine in addition to two consensus sequences for asparagine-linked glycosylation (Asn-X-Ser/Thr);  
10 another larger fragment, amino acids 128 to 267 of SEQ ID NO:2, having both a high frequency of proline, serine, and threonine and containing fifteen repeats of the following ten amino acid consensus sequence: Ala-(Thr/Met)-Glu-Ala-Gln-Thr-Thr-(Pro/Arg/Gln/Ala/Glu)-(Leu/Pro)-(Ala/Thr) (smaller fragments within this large fragment may also retain the capacity to interact with P-selectin or act as inhibitors of P-selectin-mediated intercellular adhesion); the region containing a consensus sequence for asparagine-linked glycosylation and comprising amino acids  
15 268 to 308 of SEQ ID NO:2; the hydrophobic region of the protein represented by amino acids 309 to 333 of SEQ ID NO:2; and the amphophilic region of the P-selectin ligand protein from amino acids 334 to 402 of SEQ ID NO:2. Additional fragments may comprise amino acid 43 to amino acid 56 of SEQ ID NO:2 or amino acid 42 to amino acid 60 of SEQ ID NO:2, with one or more sulfated or phosphorylated (Domcheck *et al.*, *Biochemistry* 31:9865-9870 (1992)) tyrosines  
20 at amino acid 46, amino acid 48, and/or amino acid 51. Fragments of the P-selectin ligand protein may be in linear form or they may be cyclized using known methods, for example, as described in H.U. Saragovi, *et al.*, *Bio/Technology* 10, 773-778 (1992) and in R.S. McDowell, *et al.*, *J. Amer. Chem. Soc.* 114, 9245-9253 (1992), both of which are incorporated herein by reference. For the purposes of the present invention, all references to "P-selectin ligand protein" herein  
25 include fragments capable of binding to P-selectin.

Such fragments may be fused to carrier molecules such as immunoglobulins, to increase the valency of P-selectin ligand binding sites. For example, soluble forms of the P-selectin ligand protein such as the fragments from amino acid 42 to amino acid 295 or from amino acid 42 to amino acid 88 of SEQ ID NO:2 may be fused through "linker" sequences to the Fc portion of an  
30 immunoglobulin (native sequence or mutated sequences for conferring desirable qualities (such as longer half-life or reduced immunogenicity) to the resulting chimera). For a bivalent form of the P-selectin ligand protein, such a fusion could be to the Fc portion of an IgG molecule as in Example 5(D) and in SEQ ID NO:6. Other immunoglobulin isotypes may also be used to generate such fusions. For example, a P-selectin ligand protein - IgM fusion would generate a  
35 decavalent form of the P-selectin ligand protein of the invention.

Fusions of any of the P-selectin ligand proteins of the present invention to amino acid sequences derived from other proteins may also be constructed. Preferred P-selectin ligand proteins for such purpose include the fragments from amino acid 42 to amino acid 295 or from

5 amino acid 42 to amino acid 88 of SEQ ID NO:2. Desirable fusion proteins may incorporate amino acid sequence from proteins having a biological activity different from that of P-selectin ligand, such as, for example, cytokines, growth and differentiation factors (such as bone morphogenetic proteins (e.g., BMPs), hormones, enzymes, receptor components or fragments and other ligands. Also, P-selectin ligand protein can be chemically coupled to other proteins or  
10 pharmaceutical agents. In such usage, the P-selectin ligand protein, by virtue of the ability to interact with selectin molecules, alters the pharmacokinetics and/or biodistribution of the fused or coupled agent thereby enhancing its therapeutic efficacy. For example, fusion of a P-selectin ligand protein sequence to a cytokine sequence can direct the cytokine's activity to an area of inflammation. In such instance, the P-selectin ligand protein portion of the fusion protein will  
15 bind to selectins expressed at the site of inflammation. This binding will cause the cytokine portion of the fusion protein to become localized and available to bind its cognate receptor or any proximal cell surface. Other ligands could similarly be used in such fusion proteins to attract cells expressing their corresponding receptors to a site of P-selectin expression. Preferred examples of such fusions are described in Example 15.

20 In any fusion protein incorporating a P-selectin ligand protein, the amino acid sequence derived from a protein or proteins other than P-selectin ligand can be linked to either the C-terminus or N-terminus of the P-selectin ligand-derived sequence. The linkage may be direct (i.e., without an intervening linking sequence not derived from either protein) or through a linking sequence.

25 Methods of treating a mammalian subject using such fusion proteins are also contemplated by the present invention. In such instances, the fusion protein is used to treat a condition which is affected by the protein to which the P-selectin ligand protein is fused. For example, a fusion of a P-selectin ligand protein to IL-11 could be used to localize the activity of IL-11 to bone marrow endothelial cells which express selectins on their surface. Once localized,  
30 the IL-11 portion of the fusion protein will stimulate megakaryocyte progenitors. Similarly, a fusion of a P-selectin ligand protein to a BMP could be used to stimulate bone or cartilage formation in an area of injury. Injured tissues express P-selectin, which will bind the fusion protein. Once localized, the BMP portion of the fusion protein will stimulate bone or cartilage production in the area of injury.

35 As detailed in the Examples below, the P-selectin ligand protein of the invention was initially obtained using an expression cloning approach (Clark *et al.*, U.S. 4,675,285). A cDNA library was constructed from the human promyelocytic cell line HL-60 (S.J. Collins, *et al.*, Nature 270, 347-349 (1977), ATCC No. CCL 240). This library was cotransfected into COS cells with



5 a DNA encoding a 3/4FT, and the cotransfectants were screened for binding to a chimeric molecule consisting of the extracellular portion of P-selectin and the Fc portion of a human IgG1 monoclonal antibody. Cotransfectants which bound to the chimeric P-selectin were enriched for cDNAs encoding the P-selectin ligand protein. This screening process was repeated several times to enrich the plasmid population further for cDNAs encoding the P-selectin ligand protein. In a  
10 second cloning stage, the enriched plasmid population was again cotransfected into COS cells with the 3/4FT gene and screened for binding to a fluorescently labeled CHO cell line which expressed P-selectin on the cell surface. A single cDNA clone was obtained from this approach and was designated pMT21:PL85. The pMT21:PL85 plasmid was deposited with the American Type Culture Collection on October 16, 1992 and given the accession number ATCC 69096.

15 One novel DNA of the present invention is set forth in SEQ ID NO:1. The DNA of the present invention may encode a variety of forms of the P-selectin ligand protein. For example, in one embodiment, the DNA of the invention encodes the entire P-selectin ligand protein having the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 402. In another embodiment, the DNA of the invention encodes a form of the P-selectin ligand protein  
20 which lacks the signal sequence and which is characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 21 to amino acid 402. In yet another embodiment, the DNA of the invention encodes the mature P-selectin ligand protein characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 402. Another embodiment of the DNA of the invention encodes a soluble form of the P-selectin ligand protein characterized  
25 by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 1 to amino acid 310. The DNA of the invention is also embodied in a DNA encoding a soluble form of the mature P-selectin ligand protein, said protein being characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 42 to amino acid 310. The DNA of the invention is further embodied in a DNA sequence encoding a soluble form of the P-selectin ligand protein which  
30 lacks the signal sequence, said protein being characterized by the amino acid sequence set forth in SEQ ID NO:2 from amino acid 21 to amino acid 310. The DNA of the present invention is free from association with other human DNAs and is thus characterized as an isolated DNA. As detailed above, DNAs which encode P-selectin ligand fragments which interact with P-selectin are also included in the present invention.

35 The expression of P-selectin ligand protein mRNA transcripts has been observed in a variety of human cell lines (HL-60, THP-1, U937) and in human monocytes and polymorphonuclear leukocytes by Northern analysis using a P-selectin ligand protein cDNA probe. In all of these cell lines, a major transcript of 2.5 kb was observed. A minor species of

5 approximately 4 kb was observed in the HL60 and U937 cell lines and in polymorphonuclear leukocytes. In contrast, no P-selectin ligand mRNA expression was detected in the human hepatoblastoma cell line HepG2.

The P-selectin ligand protein of the invention is encoded by a single copy gene and is not part of a multi-gene family, as determined by Southern blot analysis. The genomic form of the P-selectin ligand protein of the invention contains a large intron of approximately 9 kb located at nucleotide 54 in the 5' untranslated region. In polymorphonuclear leukocytes and monocytes, the P-selectin ligand protein of the invention is encoded by the DNA sequence set forth in SEQ ID NO:3. In this embodiment, the P-selectin ligand protein contains sixteen repeat regions. The isolated DNA of the invention is correspondingly also embodied in the DNA sequence set forth in SEQ ID NO:3 and is contained on plasmid pPL85R16 which was deposited with the American Type Culture Collection on October 22, 1993 and given the Accession Number ATCC 75577.

The invention also encompasses allelic variations of the isolated DNA as set forth in SEQ ID NO:1 or of the isolated DNA as set forth in SEQ ID NO:3, that is, naturally-occurring alternative forms of the isolated DNA of SEQ ID NO: 1 or SEQ ID NO:3 which also encode proteins having P-selectin ligand activity. Also included in the invention are isolated DNAs which hybridize to the DNA set forth in SEQ ID NO:1 or to the DNA set forth in SEQ ID NO:3 under stringent (e.g. 4xSSC at 65°C or 50% formamide and 4xSSC at 42°C), or relaxed (4xSSC at 50°C or 30-40% formamide at 42°C) conditions, and which have P-selectin ligand protein activity. Isolated DNA sequences which encode the P-selectin ligand protein but which differ from the DNA set forth in SEQ ID NO:1 or from the DNA set forth in SEQ ID NO:3 by virtue of the degeneracy of the genetic code and which have P-selectin ligand protein activity are also encompassed by the present invention. Variations in the DNA as set forth in SEQ ID NO:1 or in the DNA as set forth in SEQ ID NO:3 which are caused by point mutations or by induced modifications which enhance the P-selectin ligand activity, half-life or production level are also included in the invention. For the purposes of the present invention all references herein to the "DNA of SEQ ID NO:1" include, in addition to DNAs comprising the specific DNA sequence set forth in SEQ ID NO:1, DNAs encoding the mature P-selectin ligand protein of SEQ ID NO:2; DNAs encoding fragments of the P-selectin ligand protein of SEQ ID NO:2 which are capable of binding to P-selectin; DNAs encoding soluble forms of the P-selectin ligand protein of SEQ ID NO:2; allelic variations of the DNA sequence of SEQ ID NO:1; DNAs which hybridize to the DNA sequence of SEQ ID NO:1 and which encode proteins having P-selectin ligand protein activity; DNAs which differ from the DNA of SEQ ID NO:1 by virtue of degeneracy of the genetic code; and the variations of the DNA sequence of SEQ ID NO:1 set forth above. Similarly,

5 all references to the "DNA of SEQ ID NO:3" include in addition to the specific sequence set forth  
in SEQ ID NO:3, DNAs encoding the mature P-selectin ligand protein of SEQ ID NO:4; DNAs  
encoding fragments of the P-selectin ligand protein of SEQ ID NO:4 which are capable of binding  
to P-selectin; DNAs encoding soluble forms of the P-selectin ligand protein of SEQ ID NO:4;  
10 allelic variations of the DNA of SEQ ID NO:3; DNAs which hybridize to the DNA sequence of  
SEQ ID NO:3 and which encode proteins having P-selectin ligand protein activity; DNAs which  
differ from the DNA of SEQ ID NO:3 by virtue of degeneracy of the genetic code; and the  
variations of the DNA of SEQ ID NO:3 set forth above.

A DNA encoding a soluble form of the P-selectin ligand protein may be prepared by  
expression of a modified DNA in which the regions encoding the transmembrane and cytoplasmic  
15 domains of the P-selectin ligand protein are deleted and/or a stop codon is introduced 3' to the  
codon for the amino acid at the carboxy terminus of the extracellular domain. For example,  
hydrophobicity analysis predicts that the P-selectin ligand protein set forth in SEQ ID NO:2 has  
a transmembrane domain comprised of amino acids 311 to 332 of SEQ ID NO:2 and a  
cytoplasmic domain comprised of amino acids 333 to 402 of SEQ ID NO:2. A modified DNA  
20 as described above may be made by standard molecular biology techniques, including site-  
directed mutagenesis methods which are known in the art or by the polymerase chain reaction  
using appropriate oligonucleotide primers. Methods for producing several DNAs encoding  
various soluble P-selectin ligand proteins are set forth in Example 5.

A DNA encoding other fragments and altered forms of P-selectin ligand protein may be  
25 prepared by expression of modified DNAs in which portions of the full-length sequence have  
been deleted or altered. Substantial deletions of the P-selectin ligand protein sequence can be  
made while retaining P-selectin ligand protein activity. For example, P-selectin ligand proteins  
comprising the sequence from amino acid 42 to amino acid 189 of SEQ ID NO: 2, the sequence  
from amino acid 42 to amino acid 118 of SEQ ID NO: 2, or the sequence from amino acid 42 to  
30 amino acid 89 of SEQ ID NO: 2 each retain the P-selectin protein binding activity and the ability  
to bind to E-selectin. P-selectin ligand proteins in which one or more N-linked glycosylation sites  
(such as those at amino acids 65, 111 and 292 of SEQ ID NO: 2) have been changed to other  
amino acids or deleted also retain P-selectin protein binding activity and the ability to bind E-  
selectin. P-selectin ligand proteins comprising from amino acid 42 to amino acid 60 of SEQ ID  
35 NO:2 (which includes a highly anionic region of the protein from amino acid 45 to amino acid  
58 of SEQ ID NO:2) also retain P-selectin ligand protein activity; however, P-selectin ligand  
proteins limited to such sequence do not bind to E-selectin. Preferably, a P-selectin ligand protein  
retains at least one (more preferably at least two and most preferably all three) of the tyrosine

5 residues found at amino acids 46, 48 and 51 of SEQ ID NO: 2, sulfation of which may contribute to P-selectin ligand protein activity. Construction of DNAs encoding these and other active fragments or altered forms of P-selectin ligand protein may be accomplished in accordance with methods known to those skilled in the art.

10 The isolated DNA of the invention may be operably linked to an expression control sequence such as the pMT2 or pED expression vectors disclosed in Kaufman *et al.*, Nucleic Acids Res. 19, 4485-4490 (1991), in order to produce the P-selectin ligand recombinantly. Many suitable expression control sequences are known in the art. General methods of expressing recombinant proteins are also known and are exemplified in R. Kaufman, Methods in Enzymology 185, 537-566 (1990). As defined herein "operably linked" means enzymatically or  
15 chemically ligated to form a covalent bond between the isolated DNA of the invention and the expression control sequence, in such a way that the P-selectin ligand protein is expressed by a host cell which has been transformed (transfected) with the ligated DNA/expression control sequence.

Several endoproteolytic enzymes are known which cleave precursor peptides at the carboxyl side of paired amino acid sequences (e.g., -Lys-Arg- and -Arg-Arg-) to yield mature  
20 proteins. Such enzymes are generally known as paired basic amino acid converting enzymes or PACE, and their use in recombinant production of mature peptides is extensively disclosed in WO 92/09698 and U.S. Application Serial No. 07/885,972, both of which are incorporated herein by reference. The PACE family of enzymes are known to increase the efficiency of proteolytic processing of precursor polypeptides in recombinant host cells. As mentioned above, the P-  
25 selectin ligand protein of the invention contains such a PACE cleavage site.

The soluble mature P-selectin ligand protein of the present invention may be made by a host cell which contains a DNA sequence encoding any soluble P-selectin ligand protein as described herein and a DNA sequence encoding PACE as described in WO 92/09698 and U.S. Application Serial No. 07/885,972, incorporated herein by reference, or using the DNA sequence  
30 of SEQ ID NO:5. Such a host cell may contain the DNAs as the result of co-transformation or sequential transformation of separate expression vectors containing the soluble P-selectin ligand protein DNA and the PACE DNA, respectively. A third DNA which encodes a 3/4FT may also be co-transformed with the DNAs encoding the P-selectin ligand protein and PACE. Alternatively, the host cell may contain the DNAs as the result of transformation of a single  
35 expression vector containing both soluble P-selectin ligand protein DNA and PACE DNA. Construction of such expression vectors is within the level of ordinary skill in molecular biology. Methods for co-transformation and transformation are also known.

5 Many DNA sequences encoding PACE are known. For example, a DNA encoding one form of PACE, known as furin, is disclosed in A.M.W. van den Ouweland *et al.*, Nucl. Acids Res. 18, 664 (1990), incorporated herein by reference. A cDNA encoding a soluble form of PACE, known as PACESOL, is set forth in SEQ ID NO:5. DNAs encoding other forms of PACE also exist, and any such PACE-encoding DNA may be used to produce the soluble mature P-selectin ligand protein of the invention, so long as the PACE is capable of cleaving the P-selectin ligand protein at amino acids 38-41. Preferably, a DNA encoding a soluble form of PACE is used to produce the soluble mature P-selectin ligand protein of the present invention.

10 The DNAs encoding a soluble form of the P-selectin ligand protein and PACE, separately or together, may be operably linked to an expression control sequence such as those contained in the pMT2 or pED expression vectors discussed above, in order to produce the PACE-cleaved soluble P-selectin ligand recombinantly. Additional suitable expression control sequences are known in the art. Examples 3(C) and 3(D) below set forth methods for producing the soluble mature P-selectin ligand protein of the invention.

15 A number of types of cells may act as suitable host cells for expression of the P-selectin ligand protein. Suitable host cells are capable of attaching carbohydrate side chains characteristic of functional P-selectin ligand protein. Such capability may arise by virtue of the presence of a suitable glycosylating enzyme within the host cell, whether naturally occurring, induced by chemical mutagenesis, or through transfection of the host cell with a suitable expression plasmid containing a DNA sequence encoding the glycosylating enzyme. Host cells include, for example, monkey COS cells, Chinese Hamster Ovary (CHO) cells, human kidney 293 cells, human epidermal A431 cells, human Colo205 cells, 3T3 cells, CV-1 cells, other transformed primate cell lines, normal diploid cells, cell strains derived from in vitro culture of primary tissue, primary explants, HeLa cells, mouse L cells, BHK, HL-60, U937, or HaK cells.

20 The P-selectin ligand protein may also be produced by operably linking the isolated DNA of the invention and one or more DNAs encoding suitable glycosylating enzymes to suitable control sequences in one or more insect expression vectors, and employing an insect expression system. Materials and methods for baculovirus/insect cell expression systems are commercially available in kit form from, *e.g.*, Invitrogen, San Diego, California, U.S.A. (the MaxBac® kit), and such methods are well known in the art, as described in Summers and Smith, Texas Agricultural Experiment Station Bulletin No. 1555 (1987), incorporated herein by reference. Soluble forms of the P-selectin ligand protein may also be produced in insect cells using appropriate isolated DNAs as described above. A DNA encoding a form of PACE may further be co-expressed in an insect host cell to produce a PACE-cleaved form of the P-selectin ligand protein.

5           Alternatively, it may be possible to produce the P-selectin ligand protein in lower  
eukaryotes such as yeast or in prokaryotes such as bacteria. Potentially suitable yeast strains  
include *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Kluyveromyces* strains, *Candida*,  
or any yeast strain capable of expressing heterologous proteins. Potentially suitable bacterial  
strains include *Escherichia coli*, *Bacillus subtilis*, *Salmonella typhimurium*, or any bacterial strain  
10           capable of expressing heterologous proteins. If the P-selectin ligand protein is made in yeast or  
bacteria, it is necessary to attach the appropriate carbohydrates to the appropriate sites on the  
protein moiety covalently, in order to obtain the glycosylated P-selectin ligand protein. Such  
covalent attachments may be accomplished using known chemical or enzymatic methods.

          The P-selectin ligand protein of the invention may also be expressed as a product of  
15           transgenic animals, e.g., as a component of the milk of transgenic cows, goats, pigs, or sheep  
which are characterized by somatic or germ cells containing a DNA sequence encoding the P-  
selectin ligand protein.

          The P-selectin binding activity of a P-selectin protein may be enhanced by co-  
transformation of a host cell with a GlcNAc transferase, preferably UDP-GlcNAc:Gal  $\beta$ 1-  
20           3GalNAc-R(GlcNAc to GalNAc) $\beta$ 1-6 GlcNAc transferase (EC 2.4.1.102), also known as "core2  
transferase."

          O-linked glycans present on P-selectin ligand protein have been shown to be important  
for binding to P-selectin (D. Sako *et al.*, *Cell* 75, 1179-1186 (1993)). It has been reported that  
sialyl Le<sup>x</sup> on O-linked glycans of myeloid cells are presented on complex, branched structures  
25           (Maemura, K. and Fukuda, M., *J. Biol. Chem.* 267, 24379-24386 (1992)). The enzyme  
responsible for generating such oligosaccharide structures is "core2". The core2 enzyme activity  
is found at very low levels in COS cells and at trace levels in CHO cells. Host cells co-  
transformed with DNAs encoding a P-selectin ligand protein, an ( $\alpha$ 1,3/ $\alpha$ 1,4) fucosyltransferase  
and core2 can produce P-selectin ligand protein exhibiting 20-30 fold enhanced binding to P-  
30           selectin.

          In certain preferred embodiments, P-selectin ligand protein is produced by co-transfecting  
a host cell with DNAs encoding soluble P-selectin ligand protein, 3/4FT, core2 and PACE.

          The P-selectin ligand protein of the invention may be prepared by culturing transformed  
host cells under culture conditions necessary to express a P-selectin binding glycoprotein. The  
35           resulting expressed glycoprotein may then be purified from culture medium or cell extracts.  
Soluble forms of the P-selectin ligand protein of the invention can be purified by affinity  
chromatography over Lentil lectin-Sepharose® and subsequent elution with 0.5M  $\alpha$ -methyl-  
mannoside. The eluted soluble P-selectin ligand protein can then be further purified and

5 concentrated by a 0-70% ammonium sulfate precipitation step. The protein is then recovered, resuspended, and further purified by size exclusion chromatography over a TSK G4000SW<sub>XL</sub>. Alternatively, full length forms of the P-selectin ligand protein of the invention can be purified by preparing a total membrane fraction from the expressing cell and extracting the membranes with a non-ionic detergent such as Triton X-100. The detergent extract can then be passed over  
10 an affinity column comprised of immobilized P-selectin, and the P-selectin ligand protein can be eluted from the column with 10mM EDTA in a buffer containing 0.1% detergent. The material eluted from the affinity column can then be dialyzed to remove EDTA and further purified over a Lentil lectin-Sepharose® affinity column, again eluting with 0.5M  $\alpha$ -methyl-mannoside.

Alternatively, the P-selectin ligand protein of the invention is concentrated using a  
15 commercially available protein concentration filter, for example, an Amicon or Millipore Pellicon ultrafiltration unit. Following the concentration step, the concentrate can be applied to a purification matrix such as a gel filtration medium. Alternatively, an anion exchange resin can be employed, for example, a matrix or substrate having pendant diethylaminoethyl (DEAE) groups. The matrices can be acrylamide, agarose, dextran, cellulose or other types commonly  
20 employed in protein purification. Alternatively, a cation exchange step can be employed. Suitable cation exchangers include various insoluble matrices comprising sulfopropyl or carboxymethyl groups. Sulfopropyl groups are preferred (e.g., S-Sepharose® columns). The purification of the P-selectin ligand protein from culture supernatant may also include one or more  
25 column steps over such affinity resins as concanavalin A-agarose, heparin-toyopearl® or Cibacrom blue 3GA Sepharose®; or by hydrophobic interaction chromatography using such resins as phenyl ether, butyl ether, or propyl ether; or by immunoaffinity chromatography.

Finally, one or more reverse-phase high performance liquid chromatography (RP-HPLC) steps employing hydrophobic RP-HPLC media, e.g., silica gel having pendant methyl or other  
30 aliphatic groups, can be employed to further purify the P-selectin ligand protein. Some or all of the foregoing purification steps, in various combinations, can also be employed to provide a substantially homogeneous isolated recombinant protein. The P-selectin ligand protein thus purified is substantially free of other mammalian proteins and is defined in accordance with the present invention as "isolated P-selectin ligand protein".

Isolated P-selectin ligand protein may be useful in treating conditions characterized by  
35 P-, E- or L-selectin mediated intercellular adhesion. Such conditions include, without limitation, myocardial infarction, bacterial or viral infection, metastatic conditions, inflammatory disorders such as arthritis, gout, uveitis, acute respiratory distress syndrome, asthma, emphysema, delayed type hypersensitivity reaction, systemic lupus erythematosus, thermal injury such as burns or

5 frostbite, autoimmune thyroiditis, experimental allergic encephalomyelitis, multiple sclerosis,  
multiple organ injury syndrome secondary to trauma, diabetes, Reynaud's syndrome, neutrophilic  
dermatosis (Sweet's syndrome), inflammatory bowel disease, Grave's disease, glomerulonephritis,  
gingivitis, periodontitis, hemolytic uremic syndrome, ulcerative colitis, Crohn's disease,  
necrotizing enterocolitis, granulocyte transfusion associated syndrome, cytokine-induced toxicity,  
10 and the like. Isolated P-selectin ligand protein may also be useful in organ transplantation, both  
to prepare organs for transplantation and to quell organ transplant rejection. Accordingly, P-  
selectin ligand protein may be administered to a living or non-living organ donor, prior to organ  
removal. In addition, P-selectin ligand protein may be administered "*ex-vivo*" to the donor organ  
concomitantly with organ preservation solution, prior to, and/or subsequent to surgical  
15 anastomosis with the recipient. Isolated P-selectin ligand protein may be used to treat  
hemodialysis and leukapheresis patients. Additionally, isolated P-selectin ligand protein may be used as an  
antimetastatic agent. Isolated P-selectin ligand protein may be used itself as an  
inhibitor of P-, E- or L-selectin-mediated intercellular adhesion or to design inhibitors of P-, E-  
or L-selectin-mediated intercellular adhesion. The present invention encompasses both  
20 pharmaceutical compositions containing isolated P-selectin ligand protein and therapeutic  
methods of treatment or use which employ isolated P-selectin ligand protein.

Isolated P-selectin ligand protein, purified from cells or recombinantly produced, may be  
used as a pharmaceutical composition when combined with a pharmaceutically acceptable carrier.  
Such a composition may contain, in addition to P-selectin ligand protein and carrier, diluents,  
25 fillers, salts, buffers, stabilizers, solubilizers, and other materials well known in the art. The term  
"pharmaceutically acceptable" means a non-toxic material that does not interfere with the  
effectiveness of the biological activity of the active ingredient(s). The characteristics of the carrier  
will depend on the route of administration. The pharmaceutical composition of the invention may  
also contain cytokines, lymphokines, or other hematopoietic factors such as M-CSF, GM-CSF,  
30 IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-11, IL-12, G-CSF, Meg-CSF, stem  
cell factor, and erythropoietin. The pharmaceutical composition may contain thrombolytic or anti-  
thrombotic factors such as plasminogen activator and Factor VIII. The pharmaceutical  
composition may further contain other anti-inflammatory agents. Such additional factors and/or  
agents may be included in the pharmaceutical composition to produce a synergistic effect with  
35 isolated P-selectin ligand protein, or to minimize side effects caused by the isolated P-selectin  
ligand protein. Conversely, isolated P-selectin ligand protein may be included in formulations  
of the particular cytokine, lymphokine, other hematopoietic factor, thrombolytic or anti-  
thrombotic factor, or anti-inflammatory agent to minimize side effects of the cytokine.



5 lymphokine, other hematopoietic factor, thrombolytic or anti-thrombotic factor, or anti-inflammatory agent.

The pharmaceutical composition of the invention may be in the form of a liposome in which isolated P-selectin ligand protein is combined, in addition to other pharmaceutically acceptable carriers, with amphipathic agents such as lipids which exist in aggregated form as micelles, insoluble monolayers, liquid crystals, or lamellar layers which in aqueous solution. Suitable lipids for liposomal formulation include, without limitation, monoglycerides, diglycerides, sulfatides, lysolecithin, phospholipids, saponin, bile acids, and the like. Preparation of such liposomal formulations is within the level of skill in the art, as disclosed, for example, in U.S. Patent No. 4,235,871; U.S. Patent No. 4,501,728; U.S. Patent No. 4,837,028; and U.S. Patent No. 4,737,323, all of which are incorporated herein by reference.

As used herein, the term "therapeutically effective amount" means the total amount of each active component of the pharmaceutical composition or method that is sufficient to show a meaningful patient benefit, i.e., healing of chronic conditions characterized by P-selectin- or E-selectin-mediated cellular adhesion or increase in rate of healing of such conditions. When applied to an individual active ingredient, administered alone, the term refers to that ingredient alone. When applied to a combination, the term refers to combined amounts of the active ingredients that result in the therapeutic effect, whether administered in combination, serially or simultaneously.

In practicing the method of treatment or use of the present invention, a therapeutically effective amount of isolated P-selectin ligand protein is administered to a mammal having a P-selectin-mediated disease state. Isolated P-selectin ligand protein may be administered in accordance with the method of the invention either alone or in combination with other therapies such as treatments employing receptor antagonists, ligand antagonists, cytokines, lymphokines or other hematopoietic factors. When co-administered with one or more cytokines, lymphokines or other hematopoietic factors, isolated P-selectin ligand protein may be administered either simultaneously with the cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors, or sequentially. If administered sequentially, the attending physician will decide on the appropriate sequence of administering isolated P-selectin ligand protein in combination with cytokine(s), lymphokine(s), other hematopoietic factor(s), thrombolytic or anti-thrombotic factors.

Administration of isolated P-selectin ligand protein used in the pharmaceutical composition or to practice the method of the present invention can be carried out in a variety of

5 conventional ways, such as oral ingestion, inhalation, or cutaneous, subcutaneous, or intravenous injection. Intravenous administration to the patient is preferred.

When a therapeutically effective amount of isolated P-selectin ligand protein is administered orally, isolated P-selectin ligand protein will be in the form of a tablet, capsule, powder, solution or elixir. When administered in tablet form, the pharmaceutical composition of  
10 the invention may additionally contain a solid carrier such as a gelatin or an adjuvant. The tablet, capsule, and powder contain from about 5 to 95% isolated P-selectin ligand protein, and preferably from about 25 to 90% isolated P-selectin ligand protein. When administered in liquid form, a liquid carrier such as water, petroleum, oils of animal or plant origin such as peanut oil, mineral oil, soybean oil, or sesame oil, or synthetic oils may be added. The liquid form of the  
15 pharmaceutical composition may further contain physiological saline solution, dextrose or other saccharide solution, or glycols such as ethylene glycol, propylene glycol or polyethylene glycol. When administered in liquid form, the pharmaceutical composition contains from about 0.5 to 90% by weight of isolated P-selectin ligand protein and preferably from about 1 to 50% isolated P-selectin ligand protein.

20 When a therapeutically effective amount of isolated P-selectin ligand protein is administered by intravenous, cutaneous or subcutaneous injection, isolated P-selectin ligand protein will be in the form of a pyrogen-free, parenterally acceptable aqueous solution. The preparation of such parenterally acceptable protein solutions, having due regard to pH, isotonicity, stability, and the like, is within the skill in the art. A preferred pharmaceutical composition for  
25 intravenous, cutaneous, or subcutaneous injection should contain, in addition to isolated P-selectin ligand protein an isotonic vehicle such as Sodium Chloride Injection, Ringer's Injection, Dextrose Injection, Dextrose and Sodium Chloride Injection, Lactated Ringer's Injection, or other vehicle as known in the art. The pharmaceutical composition of the present invention may also contain stabilizers, preservatives, buffers, antioxidants, or other additive known to those of skill in the art.

30 The amount of isolated P-selectin ligand protein in the pharmaceutical composition of the present invention will depend upon the nature and severity of the condition being treated, and on the nature of prior treatments which the patient has undergone. Ultimately, the attending physician will decide the amount of isolated P-selectin ligand protein with which to treat each individual patient. Initially, the attending physician will administer low doses of isolated P-  
35 selectin ligand protein and observe the patient's response. Larger doses of isolated P-selectin ligand protein may be administered until the optimal therapeutic effect is obtained for the patient, and at that point the dosage is not increased further. It is contemplated that the various

5 pharmaceutical compositions used to practice the method of the present invention should contain about 0.1 µg to about 100 mg of isolated P-selectin ligand protein per kg body weight.

The duration of intravenous therapy using the pharmaceutical composition of the present invention will vary, depending on the severity of the disease being treated and the condition and potential idiosyncratic response of each individual patient. It is contemplated that the duration  
10 of each application of the isolated P-selectin ligand protein will be in the range of 12 to 24 hours of continuous intravenous administration. Ultimately the attending physician will decide on the appropriate duration of intravenous therapy using the pharmaceutical composition of the present invention.

Isolated P-selectin ligand protein of the invention may also be used to immunize animals  
15 to obtain polyclonal and monoclonal antibodies which specifically react with the P-selectin ligand protein and which may inhibit P-selectin-mediated cellular adhesion. Such antibodies may be obtained using the entire P-selectin ligand protein as an immunogen, or by using fragments of P-selectin ligand protein such as the soluble mature P-selectin ligand protein. Smaller fragments of the P-selectin ligand protein may also be used to immunize animals, such as the fragments set  
20 forth below: amino acid 42 to amino acid 56 of SEQ ID NO:2 and amino acid 127 to amino acid 138 of SEQ ID NO:2. An additional peptide immunogen comprises amino acid 238 to amino acid 248 of SEQ ID NO:2, with an alanine residue added to the amino terminus of the peptide. Another peptide immunogen comprises amino acid 43 to amino acid 56 of SEQ ID NO:2 having a sulfated tyrosine in any or all of positions 46, 48 or 51. The peptide immunogens additionally  
25 may contain a cysteine residue at the carboxyl terminus, and are conjugated to a hapten such as keyhole limpet hemocyanin (KLH). Additional peptide immunogens may be generated by replacing tyrosine residues with sulfated tyrosine residues. Methods for synthesizing such peptides are known in the art, for example, as in R.P. Merrifield, J.Amer.Chem.Soc. 85, 2149-2154 (1963); J.L. Krstenansky, *et al.*, FEBS Lett. 211, 10 (1987).

30 Monoclonal antibodies binding to P-selectin ligand glycoprotein or to complex carbohydrate moieties characteristic of the P-selectin ligand glycoprotein may be useful diagnostic agents for the immunodetection of inflammatory diseases and some forms of cancer. Some cancerous cells, such as small cell lung carcinomas, may express detectable levels of the P-selectin ligand protein. This abnormal expression of the P-selectin ligand protein by cancer cells may play  
35 a role in the metastasis of these cells.

Neutralizing monoclonal antibodies binding to P-selectin ligand glycoprotein or to complex carbohydrates characteristic of P-selectin ligand glycoprotein may also be useful therapeutics for both inflammatory diseases and also in the treatment of some forms of cancer



5 In such a screening assay, a first binding mixture is formed by combining P-selectin or E-selectin and P-selectin ligand protein, and the amount of binding in the first binding mixture ( $B_0$ ) is measured. A second binding mixture is also formed by combining P-, E- or L-selectin, P-selectin ligand protein, and the compound or agent to be screened, and the amount of binding in the second binding mixture (B) is measured. The amounts of binding in the first and second  
10 binding mixtures are compared, for example, by performing a  $B/B_0$  calculation. A compound or agent is considered to be capable of inhibiting P-, E- or L-selectin mediated intercellular adhesion if a decrease in binding in the second binding mixture as compared to the first binding mixture is observed. The formulation and optimization of binding mixtures is within the level of skill in the art, such binding mixtures may also contain buffers and salts necessary to enhance or to  
15 optimize binding, and additional control assays may be included in the screening assay of the invention.

Compounds found to reduce by at least about 10%, preferably greater than about 50% or more of the binding activity of P-selectin ligand protein to P-, E- or L-selectin may thus be identified and then secondarily screened in other selectin binding assays, including assays binding  
20 to L-selectin and *in vivo* assays. By these means compounds having inhibitory activity for selectin-mediated intercellular adhesion which may be suitable as anti-inflammatory agents may be identified.

### EXAMPLE 1

#### 25 CLONING OF THE P-SELECTIN LIGAND PROTEIN GENE

##### A. Construction of the HL60 cDNA library

An HL60 cDNA library was constructed for expression cloning the P-selectin ligand. PolyA<sup>+</sup> RNA was isolated from total RNA from the human promyelocytic cell line HL60 (S.J. Collins, *et al.*, *supra*) using a Fast Track mRNA Isolation Kit (Invitrogen; San Diego, CA).  
30 Double stranded cDNA was synthesized from the polyA<sup>+</sup> RNA fraction and blunt-end ligated with EcoRI adaptors (5'-AATTCCGTCGACTCTAGAG-3', SEQ ID NO:7; 5'-CTCTAGAGTCGACGG-3', SEQ ID NO:8). The cDNA was ligated into the expression vector pMT21 (R. Kaufman *et al.*, J. Mol. Cell. Biol. 2, 946-958 (1989) that had been incubated sequentially with EcoRI  
35 endonuclease and calf intestinal alkaline phosphatase and gel purified. The ligation product was electroporated in 2  $\mu$ l aliquots into competent *E. coli* DH5 $\alpha$  cells and grown in 1 ml of SOB medium (J. Sambrook *et al.*, Molecular Cloning: A Laboratory Manual, New York, Cold Spring Harbor Laboratory Press, p1.90 (1989)) which has been supplemented with 10 mM MgCl<sub>2</sub>, 10

5 mM MgSO<sub>4</sub>, and 2% glycerol for one hour at 37°C. In order to divide the library into smaller subsets, an aliquot from each ml of bacterial suspension was plated onto agar plates in the presence of ampicillin, and the number of colonies per ml was calculated. Assuming that each colony represented one cDNA clone, 600,000 clones were generated and divided into subsets of approximately 16,000 clones per pool. Each of the 38 pools were grown overnight in L-broth in  
10 the presence of ampicillin and the plasmids were purified over a CsCl gradient.

**B. Screening for the P-selectin ligand protein gene**

In the first stage, the LEC-γ1 binding assay of Example 4(A) was utilized to pan the HL60 cDNA library and thereby to enrich for the plasmid of interest. Six μg of each HL60 cDNA  
15 library pool was co-transfected with 2 μg of α 3/4FT gene (Example 2) into COS cells. Approximately 45 hours post-transfection, the COS cells were lifted from the plates by incubating the cells in 1 mM EGTA for 15 min. at 37°C, followed by scraping with cell lifters. The cells were washed twice in Hanks buffered saline solution containing 1 mM calcium (HBSS). The cells were resuspended in 4 ml of HBSS. The resuspended transfected COS cells were screened using  
20 the LEC-γ1 binding assay described in Example 4(A).

The plasmids from adherent COS cells were recovered from a Hirts extract [B. Hirts, J. Mol. Biol., 26, 365-369 (1967)] and then electroporated into *E. coli* DH5α cells for amplification. The enriched population of plasmids was purified over a CsCl gradient and re-transfected along with the 3/4FT gene (Example 2) into COS cells. The transfection, screening, and plasmid  
25 amplification process was repeated for a total of three times before a pool that bound to the LEC-γ1-coated plates was visually detected. The positive plasmid pool was subsequently broken down into subsets. This involved electroporating the Hirts extract from the positive pool into *E. coli* DH5α cells and quantitating colonies per ml as described above. Various pool sizes were produced by plating out a predetermined number of colonies on agar plates in the presence of  
30 ampicillin. Duplicate plates were prepared by performing nitrocellulose lifts and storing the filters on new agar plates. The duplicate plates served as reference plates for selecting individual or groups of colonies from any pool identified as being positive.

In the second stage of cloning, COS cells were co-transfected with the sublibrary pools and the 3/4FT gene by the same procedure used in the initial steps of screening. Forty-eight hours  
35 post-transfection, the transfected cells were screened using the fluorescent CHO:P-selectin assay of Example 4(B). Positive pools were further subdivided, as described above, until finally individual colonies were screened and positive clones identified. Using this method, a single positive clone, pMT21:PL85, was found to encode the P-selectin ligand protein. The DNA

5 sequence of the P-selectin ligand contained in pMT21:PL85 is set forth in SEQ ID NO:1. and the binding characteristics of the P-selectin ligand protein encoded by pMT21:PL85 are set forth in Example 4(C) below.

## EXAMPLE 2

10

### CLONING THE $\alpha$ 1,3/1,4 FUCOSYLTRANSFERASE GENE

The  $\alpha$  1,3/1,4 fucosyltransferase gene (3/4FT) was cloned from total human genomic DNA (Clontech Laboratories) by means of PCR. The sense oligonucleotide primer contained an XbaI site and the 5' terminus of the gene (5'-TAGCATACGCTCTAGAGCATGGATCCCCTGGGTGCAGCCAAGC-3', SEQ ID NO:9), and the antisense oligonucleotide primer contained an EcoRI site and the 3' terminus of the gene (5'-CCGGAAATTCTCAGGTGAACCAAGCCGC-3', SEQ ID NO:10). The PCR product was sequentially digested with XbaI and EcoRI and purified by standard gel purification methods. This gene was then ligated with vector pMT3Sv2ADA (R. Kaufman, Methods in Enzymology, *supra*) that had also been sequentially digested with XbaI and EcoRI and purified by standard gel purification methods. Competent HB101 cells (Biorad) were transformed with this ligation product and then plated on agar plates in the presence of ampicillin. Nitrocellulose filter lifts of ampicillin-resistant transformants were probed with a radiolabelled oligonucleotide (5'-AAGTATCTGTCCAGGGCTTCCAGGT-3', SEQ ID NO:11) complementary to the nucleotide region 506-530 in the middle of the gene (J. Sambrook *et al.*, *supra*).

Plasmid DNA minipreps were prepared from twelve positive clones. The purified DNA was then digested with EcoRI and XbaI to identify the correct clone with the proper size insert. This clone (pEA.3/4FT) was then grown up large scale and the DNA isolated by CsCl density gradient banding (J. Sambrook *et al.*, *supra*). DNA sequencing confirmed the identity of the 3/4FT gene. The functionality of the gene was assessed in a cell-cell binding assay as follows. COS-1 monkey cells [(clone M6; M. Horwitz *et al.*, Mol. Appl. Genet., 2:147-149, (1983))] were transfected with 3/4FT using DEAE dextran followed by DMSO shock treatment and chloroquine incubation [L. Sompeyrac and K. Dana, Proc. Natl. Acad. Sci., 78:7575-7578 (1981); M. Lopata *et al.*, Nucleic Acids Res., 12:5707-5717, (1984); H. Luthman and G. Magnuson, Nucleic Acids Res., 11:1295-1308, (1983)]. The transfected COS cells were suspended and quantitated for binding to a CHO line expressing E-selectin [G. Larsen *et al.*, J. Biol. Chem. 267:11104-11110, (1992)]. This assay confirmed that the COS cells transfected with 3/4FT can express the sialylated Lewis<sup>x</sup> epitope on the cell surface.

5

**EXAMPLE 3****EXPRESSION OF THE P-SELECTIN LIGAND PROTEIN****A. Expression of the P-selectin Ligand in LEC11 cells**

10 Functional P-selectin ligand was expressed in the SLe<sup>x</sup>-positive Chinese hamster ovary (CHO) cell line LEC11 (Campbell, C. and Stanley, P. Cell 35:303-309 (1983) as follows: approximately 8 µg of plasmid containing the P-selectin ligand gene (pMT21:PL85, Example 1) was transfected into LEC11 cells. At 68 hours post-transfection, the cells were treated with 2.5 mM sodium butyrate for 4 hours. The cells were observed to induce P-selectin adhesion, as  
15 determined using the 6-CFD labeled CHO:P-selectin cell binding assay (described in Example 4, section B). In contrast, neither LEC11 cells alone nor LEC11 cells transfected with a control plasmid induced P-selectin adhesion.

**B. Expression of Soluble P-Selectin Ligand in COS cells**

20 COS cells were transfected with 8 µg pED.sPSL.T7 (see Example 5C) and 4 µg pEA.3/4 FT plasmid of Example 2, 8 µg pED.sPSL.T7 alone, or 8 µg plasmid vector (pMT21) and 4 µg pEA.3/4 FT gene. Forty-five hr post-transfection, the cells were rinsed twice in PBS and incubated overnight at 37°C in serum-free DMEM minus phenol red (JRH Biosciences) supplemented with 2 mM L-glutamine, 100 U/ml penicillin and 100 µg/ml streptomycin.  
25 Phenylmethylsulfonyl fluoride, aprotinin and NaN<sub>3</sub> were added to final concentrations of 1mM, 2 µg/ml and 0.02%, respectively, and the conditioned medium was centrifuged to remove all debris.

For immunoprecipitation experiments, the labeled soluble P-selectin ligand protein was produced by co-transfecting COS cells with pED.sPSL.T7 and pEA.3/4 FT. At forty-five hr post-  
30 transfection, the COS cells were labeled with 250 µCi/ml <sup>35</sup>S methionine (NEN) for 5 hours and the medium was collected. Expression of sPSL.T7 protein was confirmed by immunoprecipitation with anti-T7 antibodies.

**C. Expression of PACE-cleaved P-selectin ligand in COS Cells**

35 COS cells were co-transfected with the pED.sPSL.T7 plasmid of Example 5(C), the pEA.3/4FT cDNA of Example 2, and a plasmid containing the PACE cDNA as set forth in SEQ ID NO:5. A parallel control co-transfection was done using only the pED.sPSL.T7 plasmid and the pEA.3/4FT plasmid. After 45 hours, conditioned medium from these transfected COS cells



5 was coated onto plastic dishes and binding to CHO:P-selectin cells (Example 4) was determined. An approximately two-fold increase in bound CHO:P-selectin cells was observed for dishes coated with medium containing the P-selectin ligand co-expressed with PACE, as compared with medium containing P-selectin ligand which had not been co-expressed with PACE. Amino acid sequencing of the N-terminus of purified sPSL.T7 protein from the PACE co-transfection showed  
10 that all of the ligand had been cleaved at the PACE consensus site (amino acids 38-41 of SEQ ID NO:1). Radiolabelling of co-transfected COS cells with <sup>35</sup>S-methionine and subsequent SDS-polyacrylamide gel electrophoresis and autoradiography showed that comparable quantities of the P-selectin ligand had been secreted in both co-transfections.

15 **D. Expression of the P-selectin Ligand Protein in CHO Cells**

A full-length form (amino acids 1-402) of the P-selectin ligand protein was expressed in the CHO(DUKX) cell line (Urlaub & Chasin, Proc. Natl. Acad. Sci. USA 77, 4216-4220 (1980)) as follows: approximately 25 µg of the pMT21:PL85 plasmid and approximately 8 µg of the pED.3/4FT (produced by restriction of pEA.3/4FT with EcoRI and XbaI and insertion of the  
20 resulting fragment into the pED plasmid) were co-transfected into CHO(DUKX) cells using the calcium phosphate method. Transfectants were selected for resistance to methotrexate. After two weeks, individual colonies were screened for SLe<sup>x</sup> expression by using a conjugate of an anti SLe<sup>x</sup> antibody (CSLEX-1, U.S. 4,752,569) and sheep red blood cells (sRBC) prepared by the chromic chloride method (Goding, J. W., J. Immunol. Methods 10:61-66 (1976) as follows: sRBC were  
25 washed with 0.15M NaCl until the wash became clear and then a 50% suspension of sRBC was prepared in 0.15M NaCl. One ml of 0.01% chromic chloride solution was added dropwise while vortexing to 0.2 ml of a sRBC suspension containing 50 µg of CSLEX-1. After incubating at 37°C for 30 minutes, 10 ml of phosphate buffered saline (PBS) solution was added to the reaction. The conjugate was washed once before resuspending into 10 ml of PBS. The plates  
30 containing transfectants were washed with PBS and then 3 ml of PBS and one ml of the sRBC/CSLEX-1 conjugate was added to each plate. Positive colonies were red on a transilluminator and were picked into alpha medium with 10% fetal bovine serum. After two weeks, colonies were subjected to stepwise amplification using methotrexate at concentrations of 2, 10, 25, 100, 250 nM. The stable cell line obtained was designated CD-PSGL-1 (R3.4).  
35 Expression of the P-selectin ligand protein was confirmed by immunoprecipitation studies using the polyclonal anti-P-selectin ligand protein antibody of Example 7(A). The functionality of the P-selectin ligand protein produced by the CD-PSGL-1 (R3.4) cell line was tested by assaying the transfectants for binding to LEC-γ1 as in Example 4(A).

5 The sPSL.T7 protein was expressed in a stable CHO-PACE line which was already  
expressing the cDNA encoding PACE as set forth in SEQ ID NO:5 under adenosine deaminase  
selection (Kaufman, *et al.*, PNAS (USA) 83:3136-3140 (1986)). The psPSL.T7 (25  $\mu$ g) and  
pED.3/4FT (8  $\mu$ g) plasmids were cotransfected into CHO-PACE cells using the calcium  
10 phosphate method. Transfectants were selected for resistance to methotrexate, and individual  
colonies which bound to the sRBC/CSLEX-1 conjugate were picked. After two weeks in culture,  
the colonies were subjected to stepwise amplification as described above. The stable cell line  
obtained was designated CP/PSL-T7 (R4.1). Expression of sPSL.T7 protein was confirmed by  
standard immunoprecipitation methods using either a T7 specific monoclonal antibody or the  
15 LEC- $\gamma$ 1 chimera of Example 4(A). In a similar fashion, a stable cell line expressing the mature  
full length form (amino acids 42-402) of the P-selectin ligand protein was obtained by co-  
transfection of pMT21:PL85 and pED.3/4FT into the CHO-PACE line.

Stable cell lines expressing the sPSL.Q protein of Example 5(B) and the sPSL.Fc protein  
of Example 5(D) were constructed as follows: plasmids pED.sPSL.Q (25  $\mu$ g) or pED.sPSL.Fc  
20 (25  $\mu$ g) were cotransfected with approximately 25  $\mu$ g of the pED.3/4FT plasmid described above  
and approximately 20  $\mu$ g of a plasmid containing the PACE cDNA as set forth in SEQ ID NO:5)  
as well as the neomycin resistance gene into CHO(DUKX) cells using the calcium phosphate  
method. Transfectants were selected for resistance to methotrexate and the G418 antibiotic.  
Approximately two weeks later, individual colonies were screened for SLe<sup>x</sup> expression using  
sRBC/CSLEX-1 conjugate binding. The positive colonies were picked in G418 medium at 1  
25 mg/ml concentration. After 2-3 weeks in culture, cells were amplified with methotrexate in a  
stepwise selection. The stable cell lines obtained were designated CD-sPSL.Q (R8.2) and CD-  
sPSL.Fc (R8.1), respectively. The expression of sPSL.Q and sPSL.Fc protein was confirmed by  
standard immunoprecipitation method using the anti P-selectin ligand protein polyclonal antibody  
of Example 7(A).

30

#### **EXAMPLE 4**

##### **ASSAYS OF P-SELECTIN-MEDIATED INTERCELLULAR ADHESION**

###### 35 A. **LEC- $\gamma$ 1 Binding Assay**

A DNA encoding a chimeric form of P-selectin conjugated to the Fc portion of a human  
IgG $\gamma$ 1 (LEC- $\gamma$ 1) was constructed using known methods (Aruffo *et al.* Cell 67, 35-44 (1991)), and

5 stably transfected into dhfr<sup>r</sup> CHO cells (CHO DUKX) for high level production of the chimeric LEC- $\gamma$ 1 protein, which was purified for use in the binding assay set forth below.

Petri dishes were coated first with a polyclonal anti-human IgG $\gamma$ 1 Fc antibody and then with LEC- $\gamma$ 1. This method orients the LEC- $\gamma$ 1 construct such that the P-selectin portion of the chimeric molecule is presented on the surface of the plates. Adhesion of HL60 cells to the oriented LEC- $\gamma$ 1 was quantitated in the presence and absence of calcium. HL60 adhesion was shown to be calcium dependent, confirming that the chimeric molecule had retained functional binding of P-selectin to its ligand on HL60 cells. The binding of HL60 cells to oriented LEC- $\gamma$ 1 was also shown to be blocked by a neutralizing monoclonal antibody to P-selectin, demonstrating the specificity of P-selectin binding.

15

#### **B. Fluorescent CHO-P-selectin Binding Assay**

The assay employed a fluorescently labeled CHO:P-selectin cell line (Larsen *et al.*, J. Biol. Chem. 267, 11104-11110 (1992)) that can bind to and form clusters on the surface of COS cells that are co-transfected with the P-selectin ligand gene and the 3/4 FT gene. The CHO:P-selectin cells were suspended at  $1.5 \times 10^6$  cells/ml in 1% fetal bovine serum in DME medium and labeled by adding 6-carboxyfluorescein diacetate (6-CFD) to a final concentration of 100  $\mu$ g/ml. After incubation at 37°C for 15 minutes, the cells were washed in medium and resuspended at  $1 \times 10^5$  cells/ml. Five ml of the labeled cells were added to each washed COS transfectant-containing plate to be assayed and incubated at room temperature for 10 minutes. Nonadherent cells were removed by four washes with medium. The plates were then scanned by fluorescence microscopy for rosettes of adherent CHO:P-selectin cells.

25

#### **C. Quantitative adhesion assay using radioactively labeled CHO:P-selectin cells**

COS cells were co-transfected with the pMT21:PL85 plasmid of Example 1 and the pEA.3/4FT plasmid of Example 2 by the same procedure used in the initial stages of screening. As controls, COS cells were transfected with pMT21:PL85 alone, or with pEA.3/4FT alone, or with a similar plasmid containing no insert ("mock"). 24 hours post-transfection, the transfected cells were trypsinized and distributed into Costar 6-well tissue culture plates. CHO:P-selectin cells were labeled for 16 hours with  $^3$ H-thymidine using known methods and preincubated at  $0.5 \times 10^6$  cells/ml for 30 minutes at 4°C in  $\alpha$  medium containing 1% BSA (control);  $\alpha$  medium containing 1% BSA, 5 mM EDTA and 5 mM EGTA;  $\alpha$  medium containing 1% BSA and 10  $\mu$ g/ml of a neutralizing anti P-selectin monoclonal antibody; and  $\alpha$  medium containing 1% BSA and a non-neutralizing anti-P-selectin monoclonal antibody. The preincubated cells were then

35



- 5 The resulting plasmid was designated pED.sPSL.QC, and the protein expressed from the plasmid was designated sPSL.QC.

**B. Construction of psPSL.Q**

The fragment was purified and ligated into the pED plasmid (Kaufman *et al.*, 1991) between the XbaI and EcoRI sites, along with the double stranded synthetic oligonucleotide DNA that recreated the codons from Asn 296 to Gln 309 and introduced a novel stop codon immediately following Gln 309. The sequence of the oligos is as follows:

5'-AACTACCCAGTGGGAGCACCAGACCACATCTCTGTGAAGCAGTAG (SEQ ID NO:14)

5'-AATTCTACTGCTTCACAGAGATGTGGTCTGGTGCTCCCACTGGGTAGTT (SEQ ID NO:15)

- 15 The resulting plasmid was designated pED.sPSL.Q, and the protein expressed from the plasmid was designated sPSL.Q.

**C. Construction of psPSL.T7**

Oligonucleotides encoding 14 amino acids including an epitope derived from the phage T7 major capsid protein were synthesized, creating a C-terminal fusion of the epitope "tag" with an additional 32 amino acids derived from the vector sequence. Two oligonucleotides having the sequences

5'-CTAGACCCGGGATGGCATCCATGACAGGAGGACAACAAATGGTAGGCCGTAG (SEQ ID NO: 16) and

- 25 5'-AATTCTACGGCCTACCCATTTGTTGTCTCCTGTTCATGGATGCCATCCCGGGT (SEQ ID NO:17)

were duplexed and ligated with the large XbaI-EcoRI fragment of mammalian expression plasmid pED. The resulting plasmid, pED.T7 was restricted with XbaI and SmaI and ligated to the 950 bp XbaI-HincII fragment described above, resulting in plasmid pED.sPSL.T7.

- 30 The protein resulting from expression of pED.sPSL.T7 was designated sPSL.T7.

**D. Construction of Soluble P-selectin Ligand--IgGFc Chimera**

The plasmid DNA encoding a soluble, extracellular form of the P-selectin ligand protein fused to the Fc portion of human immunoglobulin IgG1 was constructed as follows: the mammalian expression vector pED.Fc contains sequences encoding the Fc region of a human IgG1 with a novel linker sequence enabling the fusion of coding sequences amino terminal to the hinge region via a unique XbaI restriction site. A three fragment ligation was performed: pED.Fc was restricted with XbaI and gel purified in linear form. The 950 bp fragment from pMT21:PL85

5 described above comprised the second fragment. The third fragment consisted of annealed synthetic oligonucleotide DNAs having the following sequence:

5' - CTGCGGCCGCGAGT (SEQ ID NO:18)

5' - CTAGACTGCGGCCGCGAG (SEQ ID NO:19)

The ligation products were grown as plasmid DNAs and individual clones having the correct  
10 configuration were identified by DNA sequencing. The plasmid was designated pED.PSL.Fc. The DNA coding region of the resulting soluble P-selectin ligand /Fc fusion protein is shown in SEQ ID NO:6.

### EXAMPLE 6

#### 15 CHARACTERIZATION OF EXPRESSED P-SELECTIN LIGANDS

##### A. Binding Characterization of Full-Length P-selectin Ligand Protein Expressed on COS Cells

Co-transfection of COS cells with the pEA.3/4FT plasmid of Example 2 and the  
20 pMT21:PL85 plasmid of Example 1 yields COS cells which specifically bind to CHO:P-selectin cells. This binding is observed only upon co-transfection of pEA.3/4FT and pMT21:PL85; use of either plasmid alone generates COS cells which do not bind to CHO:P-selectin cells. No binding is observed between the parental CHO(DUKX) cell line which does not express P-selectin and COS cells co-transfected with pEA.3/4FT and pMT21:PL85. The binding between the co-  
25 transfected COS cells and CHO:P-selectin cells is sensitive to chelators of divalent ions such as EDTA and EGTA, consistent with the Ca<sup>++</sup> dependency of P-selectin mediated cellular adhesion. A neutralizing anti-P-selectin monoclonal antibody blocked the binding between the CHO:P-selectin cells and the COS cells which had been co-transfected with pEA.3/4FT and pMT21:PL85, while a non-neutralizing anti-P-selectin monoclonal antibody had no effect on the  
30 binding. The antibody results indicate that the functional domain(s) of P-selectin are required for binding to P-selectin ligand protein expressed on the surface of COS cells.

##### B. Electrophoretic Characterization of Full-Length P-selectin Ligand Expressed in COS Cells

35 Detergent extracts of co-transfected COS cells were prepared as follows: 45 hours post co-transfection, approximately  $1.5 \times 10^7$  cells were suspended in 5 ml of lysis buffer (10mM Piperazine-N,N'-bis[2-ethanesulfonic acid] (PIPES) pH 7.5, 100 mM KCl, 3 mM MgCl<sub>2</sub>, 1 mM benzamidine, 0.5 µg/ml leupeptin, 0.75 µg/ml pepstatin, 1 mM ethylmaleimide, and 1 µg/ml aprotinin) and lysed by sonication. Cellular debris was removed by low speed centrifugation (500  
40 x g, 10 minutes), and a membrane fraction collected by ultracentrifugation (100,000 x g, 60 min).

5 The high speed membrane pellet was resuspended in an extraction buffer (10 mM 3-[N-Morpholino]propanesulfonic acid] (MOPS) pH 7.5, 0.1 M NaCl, 0.02% NaN<sub>3</sub>, 1% Thesit® (Sigma), 1 mM benzamidine, 0.5 µg/ml leupeptin, 0.75 µg/ml pepstatin, 1 mM ethylmaleimide, and 1 µg/ml aprotinin). Samples were then subjected to SDS polyacrylamide gel electrophoresis and transfer to nitrocellulose blots as follows: an aliquot of the detergent extract was suspended  
10 in 1% SDS loading buffer and heated for 5 minutes at 100°C before loading onto an 8-16% polyacrylamide gel (reduced) or a 6% gel (non-reduced) and electrophoresed in the Laemmli buffer system. Blots were prepared using Immobilon-P® transfer membranes. The blots were immersed in 10 mM MOPS pH 7.5, 0.1 M NaCl, 0.02% NaN<sub>3</sub>, 1 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>, and 10% non-fat milk overnight at 4°C. Blots were rinsed once in the above buffer, minus the milk,  
15 and incubated in blotting buffer (10 mM MOPS pH 7.5, 0.1M NaCl, 1% bovine serum albumin, 0.05% Thesit, 1 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>) for 30 minutes at room temperature.

The blots were then probed for the P-selectin ligand as follows: 50 ng of a P-selectin/Fc chimera was pre-incubated with 3 µCi of <sup>125</sup>I-Protein A in blotting buffer for 30 minutes at room temperature. Additional excipients (e.g., EDTA, EGTA, monoclonal antibodies) could be added  
20 to the pre-incubation mixture at this point to evaluate their effects on binding of the chimera to the P-selectin ligand. The pre-incubated mixture was then incubated with the blots (prepared as above) for 60 minutes at room temperature, and the blots were subsequently washed four times with the same blotting buffer (without bovine serum albumin), air dried, and autoradiographed at -70°C.

25 Under non-reducing conditions, two bands were observed with this technique for membrane extracts prepared from co-transfected COS cells. The major band migrated with an estimated molecular weight of approximately 220 kD, whereas the minor band migrated with a molecular weight of approximately 110 kD. Under reducing conditions, only a single band was observed with a molecular weight of approximately 110 kD, indicating that under non-reducing  
30 conditions, the P-selectin ligand exists as a homodimer. The approximate molecular weight of the reduced monomer is greater than that predicted from the deduced amino acid sequence of the cDNA clone (45 kD), indicating that the expressed protein undergoes extensive post-translational modification (see Example 6(C)). The specificity of the P-selectin/Fc chimera was confirmed by the observation that a nonspecific IgG<sub>1</sub> probe yielded no bands on the blots. Additionally, the  
35 binding of the P-selectin/Fc chimera to the blots was abolished by EDTA, EGTA, and a neutralizing anti-P-selectin monoclonal antibody. Specific bands on the blots were observed only from membrane extracts of COS cells co-transfected with the pEA.3/4FT and pMT21:PL85

5 plasmids. Membrane extracts from control transfections (pEA.3/4FT or pMT21:PL85 alone) failed to yield observable bands on blots.

C. Glycosylation of P-selectin Ligand Protein

10 The presence of covalently attached carbohydrate on recombinant P-selectin ligand and its role in binding to P-selectin was determined as follows: COS cells were co-transfected with pED.sPSL.T7 of Example 5(C) and the pEA.3/4FT plasmid of Example 2. After 48 hours, the cells were pulsed with <sup>35</sup>S-methionine. 200 µl of <sup>35</sup>S methionine-labeled sPSL.T7 conditioned medium was incubated with 5 µg LEC-γ1 in the presence of 2 mM CaCl<sub>2</sub> and 1 mg/ml bovine serum albumin (BSA). After rotating for 2 hours at 4°C, Protein A-Sepharose beads (Pharmacia) were added for 1 hour at 4°C, pelleted by centrifugation and washed twice in Tris buffered saline (20 mM Tris-HCl, 150 mM NaCl pH 7.5, hereinafter TBS) containing 2 mM CaCl<sub>2</sub> and 1 mg/ml BSA. The pellets were then resuspended and treated with neuraminidase (*Streptococcus pneumoniae*), O-glycanase, and N-glycanase (all from Genzyme) as follows. All glycosidase digestions were done at 37°C overnight. For neuraminidase digestion, the pellet was resuspended in 50 µl 2-(N-morpholino)-ethanesulfonic acid (MES) buffer, pH 6.5 (Calbiochem) and 0.1% SDS, heated at 95°C for 5 minutes, then pelleted. The supernatant was modified to contain 1.4% n-Octyl B-D-glucopyranoside (OGP), 10mM calcium acetate, 20 mM sodium cacodylate and 2.5 mM PMSF, final pH 7.0 Eight µl neuraminidase was added for a final concentration of 1 unit/ml. For neuraminidase/O-glycanase digestion, the sample was prepared as above and along with the neuraminidase, the O-glycanase was also added to a final concentration of 0.1 unit/ml. For N-glycanase digestion, the pellet was resuspended in 54 ul MES buffer and 1% SDS, heated at 95°C for 5 minutes, then pelleted. The supernatant was modified to contain 0.2 M sodium phosphate, 3.5% OGP, and 2.5 mM PMSF, final pH 8.5. N-glycanase was added for a final concentration of 12 units/ml and incubated as above.

30 The effect of glycosidase treatment on sPSL.T7 was assessed in two ways. For this, each digested protein sample was divided into two equal fractions. One fraction was precipitated with the P-selectin polyclonal antibody of Example 7(A), to show the effect of digestion on the electrophoretic mobility. The other fraction was precipitated with the LEC-γ1 chimera of Example 4(A), to assess the remaining P-selectin ligand binding activity after digestion. The immunoprecipitated samples were analyzed by SDS-polyacrylamide gel electrophoresis under reducing conditions and autoradiography.

35 In the absence of glycosidase treatment, autoradiography revealed comparable bands (with molecular weights of 110 kD) for each precipitation. When the P-selectin ligand protein



5 was treated with neuraminidase, anti-P-selectin ligand polyclonal antibody precipitation revealed a slight decrease in mobility, consistent with removal of sialic acid residues. The amount of P-selectin ligand protein precipitated by LEC- $\gamma$ 1 was significantly reduced after neuraminidase treatment, consistent with the role of sialic acid residues in the P-selectin/P-selectin ligand interaction. When the P-selectin ligand protein was treated with both neuraminidase and O-glycanase, a substantial increase in electrophoretic mobility was observed after precipitation with the anti-P-selectin ligand polyclonal antibody, indicating that a number of O-linked oligosaccharide chains had been removed. However, removal of O-linked oligosaccharides from the P-selectin ligand protein may not have been complete, since the electrophoretic mobility did not correspond to a protein with a molecular weight of 38 kD, as would be predicted from the amino acid sequence set forth in SEQ ID NO:1. The neuraminidase/O-glycanase digested P-selectin ligand protein bound to LEC- $\gamma$ 1 very poorly, further indicating the role of oligosaccharides in the P-selectin/P-selectin ligand interaction. Treatment of the purified P-selectin ligand with N-glycanase resulted in a slight increase in electrophoretic mobility, demonstrating that some of the consensus sites for N-linked glycosylation are occupied. The amount of P-selectin ligand protein precipitated by LEC- $\gamma$ 1 was slightly reduced, indicating that N-linked glycosylation also contributes to the P-selectin/P-selectin ligand interaction, though not as dramatically as sialylation and O-linked glycosylation.

#### EXAMPLE 7

##### 25 POLYCLONAL ANTIBODIES SPECIFIC FOR P-SELECTIN LIGANDS

###### A. Polyclonal Rabbit anti-P-selectin Ligand Protein/Maltose Binding Protein Fusion Protein

The anti-P-selectin ligand polyclonal antibody was generated by immunizing rabbits with a fusion protein generated in *E. coli*. The fusion protein consisted of the amino terminal one-third of the P-selectin ligand (amino acids 1 to 110 of SEQ ID NO:1) fused in frame to the maltose binding protein (Maina, C. V. *et al.*, *Gene* **74**, 365-373 (1988); Riggs, P., in Current Protocols in Molecular Biology, F. M. Ausubel *et al.*, Eds., Greene Associates/Wiley Interscience (New York, 1990) chapter 16.6). Under conditions employed herein, the fusion protein antibody recognizes the P-selectin ligand protein.

###### B. Polyclonal Rabbit Anti-sPSL.T7 Protein

A soluble form of the invention (sPSL.T7; see example 5(C)) was purified to apparent homogeneity according to the following scheme: COS cells were transfected with three plasmids,

5 one encoding each of the following: sPSL.T7 (Example 5(C)), 3/4FT (Example 2), and a soluble form of PACE (as set forth in SEQ ID NO:5). After 72 hours, the conditioned medium was collected and recombinant sPSL.T7 was purified as follows.

Conditioned medium was diluted two fold with 50 mM MOPS, 150 mM NaCl, 0.5 mM CaCl<sub>2</sub> and 0.5 mM MnCl<sub>2</sub>, pH 7.2, and applied to a column of lentil lectin-Sepharose 4B  
10 equilibrated in the same buffer. After loading, the column was washed with the same buffer until the optical absorbance at 280 nm dropped to a stable baseline. The column was then eluted with the same buffer which had been adjusted to 0.5 M  $\alpha$ -methyl-mannoside and 0.3 M NaCl. Recombinant sPSL.T7 was collected over 5-15 column volumes of this elution buffer. The lentil lectin eluate was then subjected to a 0-70% ammonium sulfate precipitation by adding 472g of  
15 ammonium sulfate per liter of column eluate at 4°C. After stirring for 30 minutes, the precipitate was resuspended in a minimal volume of TBS (20 mM Tris-HCl, 150 mM NaCl, pH 7.5) and applied to a TSK G4000SW<sub>XL</sub> gel filtration column equilibrated in TBS. The flow rate on the column was 0.5 ml/min and a guard column was employed. In aliquots of < 250  $\mu$ l, the resuspended ammonium sulfate pellet was injected on the column and fractions analyzed by SDS-  
20 PAGE with Western analysis. Fractions containing sPLS.T7 were pooled and then used for immunizing rabbits.

Antibodies to sPSL.T7 were generated in the standard fashion by antigen priming and subsequent boosting over a 3 month period. Specifically, primary immunization was performed by mixing 50  $\mu$ g of sPSL.T7 (denatured by mixing in 0.1% SDS and heating for 10 minutes at  
25 100°C) with complete Freund's adjuvant and injected at five sites subcutaneously. The second (and all subsequent) boosts were performed by mixing 25  $\mu$ g of sPSL.T7 (denatured by mixing in 0.1% SDS and heating for 10 minutes at 100°C) [12.5  $\mu$ g for the third and subsequent boosts] with incomplete Freund's adjuvant and injecting at two sites subcutaneously (or later, intramuscularly) every two weeks. Test bleeds were performed every two weeks to monitor  
30 antibody titer. When the antibody titer reached a suitable level, a larger scale bleed was performed and a total serum fraction prepared. This polyclonal antibody preparation was used to inhibit the specific binding of HL60 cells to CHO:P-selectin cells in a manner similar to that described in Example 4.

This assay employed fluorescently-labeled HL60 cells (labelled with BCECFAM; 2',7'-  
35 bis-(2-carboxymethyl)-5-(and-6)-carboxyfluorescein, acetoxymethyl ester) binding to CHO cells plated on the bottom of microtiter plates. The labelled HL60 cells were pre-incubated with either sera containing polyclonal antibody or with pre-immune sera for 30 minutes at 4°C. The cells were then washed and incubated with the CHO:P-selectin cells for 10 minutes. The plates were



5           1.     Binding of mPSGL-1 transfectants to immobilized soluble P-selectin or P-selectin/IgG chimera.

48- well plates were coated with 1 ug/ml anti human Fc antibody in 50 mM Tris pH 9.5 at 4° C for five to six hours. After washing twice with HBSS buffer, P-selectin/IgG chimera (0.1-1 ug/ml conc., Example 5) was plated in HBSS buffer overnight at 4° C. The plates were blocked with BSA (3 mg/ml) at 4° C for three to four hours. In the case of soluble P-selectin ligand protein, the protein was coated directly onto plates in the same buffer. The <sup>3</sup>H labelled CHO cells were lifted with 2 mM EGTA, washed three times with PBS, and resuspended to a final density of 10<sup>6</sup> cells/ml. A 300 ul aliquot of this suspension was added to each well (300,000 cell/well). After incubating for 12 minutes at room temperature, wells were washed four times with serum free DMEM to remove unbound cells. Bound cells were lifted with 5 mM EGTA and counted in scintillation counter. U937 cells, used as a positive control for native P-selectin ligand protein binding, were pretreated with gamma globulin (5 mg/ml) to block endogenous Fc receptor before binding to P-selectin IgG chimera. Comparative binding data are shown in Fig. 1.

15           2.     Immunoprecipitation of PSGL-1 with P-selectin/IgG Chimera.

20     Recombinant full-length or soluble P-selectin ligand protein prepared from transformants, with and without additional core2, was labelled with <sup>35</sup>S-methionine and subsequently immunoprecipitated with either the anti P-selectin ligand protein polyclonal antibody or P-selectin ligand/IgG chimera as described previously in Examples 7 and 5; Sako, D., Cell 175, 1179-1186 (1993). Data are depicted in Fig. 2.

25           3.     Flow Cytometry.

Stable murine P-selectin ligand protein transfectants (with and without core2) were analyzed by standard FACS techniques using either P-selectin/IgG chimera (LecY1) (Example 5) or anti P-selectin ligand protein monoclonal antibody (MAb 275, raised against a peptide having the sequence from amino acid 42 to amino acid 56 of SEQ ID NO:2). Both reagents were pre-conjugated to FITC labelled Protein A. Cells were analyzed by FACS after incubating with this conjugate for 30 minutes at 4° C in the presence of 2 mM CaCl<sub>2</sub>. Data are depicted in Fig. 3.

### EXAMPLE 9

#### E-SELECTIN BINDING OF P-SELECTIN BINDING PROTEIN

35     E-Selectin/IgG chimera was made as described in Example 5 for the P-selectin IgG chimera using an E-selectin encoding DNA including amino acids -21 to 536 of the sequence reported in Bevilacqua *et al.*, Science, 243:1160 (1989).

5 U937 cells (approximately  $6.5 \times 10^7$ ) were recovered from tissue culture plates and divided equally into two 50 mL cultures (final concentration of  $1.3 \times 10^6$  cells/mL) containing fresh complete RPMI medium and 50  $\mu$ Ci/ml of  $^3$ H-glucosamine hydrochloride (labels the protein-linked carbohydrate of glycoproteins [Varki, FASEB 5:226-235 (1991)]. After 48 hours incubation, the cells from both cultures were recovered by centrifugation and washed three times  
10 with PBS. The pelleted cells were suspended in 2.5 mL each of a lysis buffer containing 1% Triton X-100 and disrupted by probe sonication for two minutes. The detergent lysates were placed on ice for three hours and then resonicated for an additional two minutes. The lysates were centrifuged at 16,000 rpm for five minutes, the supernatants were recovered and each adjusted to 12 mL with lysis buffer containing no detergent. To one of the two diluted cell lysates was  
15 added 100  $\mu$ L of protein A sepharose precoupled with P-selectin/IgG chimera (see Example 5) and to the other was added 100  $\mu$ L of protein A sepharose precoupled with E-selectin/IgG chimera. Both chimeric proteins were present at a density of approximately 2 mg protein/mL of resin. Binding reactions were allowed to proceed overnight at 4 degrees C with end-over-end mixing. On occasion, purified membranes from U937 cells served as the starting material for the  
20 detergent extraction of labeled proteins. In these cases, the detergent extraction and affinity precipitation steps were essentially identical to the above.

Following incubation, the two parallel reaction mixtures were each centrifuged at 2,000 rpm and supernatants were discarded. The resin pellets were washed four times with buffer (10 mM MOPS, 100 mM NaCl, 1 mM  $\text{CaCl}_2$ , 1 mM  $\text{MgCl}_2$ , 0.02%  $\text{NaN}_3$ , pH 7.5 with  
25 Triton X-100 [0.25% for the first and second washes, 0.1% for the third wash and 0.01% for the fourth wash]). A final 1 mL pre-elution wash of each resin pellet using buffer containing 0.01% Triton X-100 was conducted and these were retained for quantitation of radioactive counts by liquid scintillation counting (LSC). The resins were then eluted overnight at 4 degrees C with end-over-end mixing in 1 mL each of buffer containing 0.01% Triton X-100 and 10 mM EDTA.  
30 The supernatants were recovered by centrifugation and then quantitated by LSC.

Autoradiography of the materials released from the resins by EDTA was performed by electrophoresis of samples (approximately 10,000 cpm samples concentrated by Centricon-10 units where needed) on 10% cross-linked SDS-PAGE gels, subsequent treatment of the gels with ENHANCE (Dupont) as per the manufacturer's instructions followed by drying  
35 for two hours on a commercially available gel dryer (Bio-Rad). Exposure of the dried gels to X-ray film was conducted for a minimum of three days at -80 degrees C.

Elution of immobilized E- or P-selectin, previously exposed to detergent extracts of U937 cells and exhaustively washed, with EDTA yielded liberated,  $^3$ H-glucosamine labeled



5 5'-TATTATCTGTGCGGCCGCGGAGGCTCCGTTTCTGGCAG-3' (SEQ ID NO:23)

The template DNA for PCR reaction was pED.PSL.Fc. The PCR conditions were 94°C, 1 min.; 42°C, 1 min.; 72°C, 3 min.; 25 cycles, using a Perkin-Elmer Thermocycler. After completion of the last cycle, the reaction was treated with Klenow enzyme at 25°C for 30 min., extracted with phenol chloroform, sodium acetate added to 0.3M, and the PCR product DNA was precipitated  
10 with 2.5 volumes of ethanol. The DNA pellet was rinsed with 70% ethanol and residual ethanol was evaporated. The resuspended DNA was digested with PstI and NotI, gel purified and ligated with the pEDFc6kb fragment described above. Correct constructs were identified by restriction analysis and confirmed by DNA sequencing.

Plasmid pED.ΔY148.Fc, pED.H24.Q70.148.Fc were created by site directed mutagenesis  
15 (Maniatis et al., 1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratories) using pED.148Fc as template and the following mutagenesis oligonucleotides:

for ΔY148: 5'-CGGAGACAGGCCACCGAATTCCTGCCAGAAACG-3' (SEQ ID NO:24)

for H24: 5'-CCTCCAGAAATGCTGAGGCACAGCACTGACACCACTCCTC-3' (SEQ ID NO:25)

for Q70: 5'-GAGCTGGCCAACATGGGGCAACTGTCCACGGATTCAGCAG-3' (SEQ ID NO:26)

20 Positive clones were identified by colony hybridization (Maniatis et al, supra).

pED.FFFE.148.Fc was constructed by restricting pED.ΔY148.Fc with EcoRI and ligating the following duplexed oligonucleotides:

5'-AATTCGAGTTCCTAGATTTTG-3' (SEQ ID NO:27) and

5'-AATTCAAATCTAGGAACTCG-3' (SEQ ID NO:28).

25 Constructs of the series pED.FYYD.19.Fc, pED.FFYD.19.Fc and pED.FFFD.19.Fc, were made by restricting pED.ΔY148.Fc with EcoRI and NotI and ligating the following duplexed oligonucleotides:

for pED.FYYD.19.Fc:

5'-AATTCGAGTACCTAGATTATGATTTCTGCCAGAACTGAGCCTCCGC-3' (SEQ ID NO:29)

30 and

5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCATAATCTAGGTACTCG-3' (SEQ ID NO:30);

for pED.FFYD.19.Fc:

5'-AATTCGAGTTCCTAGATTATGATTTCTGCCAGAACTGAGCCTCCGC-3' (SEQ ID NO:31)

and

35 5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCATAATCTAGGAACTCG-3' (SEQ ID NO:32);

for pED.FFFD.19.Fc:

5'-AATTCGAGTTCCTAGATTTCTGATTTCTGCCAGAACTGAGCCTCCGC-3' (SEQ ID NO:33)

and

5 5'-GGCCGCGGAGGCTCAGTTTCTGGCAGGAAATCGAAATCTAGGAACTCG-3' (SEQ ID NO:34).

**B. Plate Binding Assay for Analysis of Deleted or Altered Forms of Soluble P-Selectin Ligand Protein**

The individual plasmid DNAs encoding the various mutated forms of soluble PSGL-1/Fc chimeras were co-transfected with pEA.3/4FT and PACE cDNA in COS cells as described in Example 3(c). 50mls of serum free medium, collected 40-64 hours post transfection from approximately 10<sup>7</sup> COS cells, was purified on a column of 0.25ml of protein A sepharose (Pharmacia) equilibrated with TBS supplemented with 2mM CaCl<sub>2</sub>. After washing with 20mls of TBS/CaCl<sub>2</sub>, the bound material was eluted with 0.5mls of 0.1M acetic acid, 0.15M NaCl, 2mM CaCl<sub>2</sub>. The eluted material was neutralized with 1/20th volume 3M Tris pH 9.0. The material was quantitated by measuring absorbance at 280nm and by comassie blue staining of PAGE/SDS/Laemmli gels.

In order to produce non-sulfated forms of soluble PSGL-1, COS cell transfections of the relevant Fc chimeras were performed as described above except that following transfection the cells were cultured in the presence of 50mM Chlorate (Sigma).

Quantitative adhesion of CHO:P-selectin, CHO:E-selectin and CHO:L-selectin expressing cells was performed as described in Example 4(c), with the following modifications: COS cell and antibodies were omitted. Instead, 48-well microtiter plates (Costar) were coated for 16 hours at 4°C with varying quantities of protein A-purified soluble PSGL-1/Fc chimeras. The unbound material was removed and the coated wells were treated with Hank's buffered saline (HBS) with 1mg/ml BSA and 2mM CaCl<sub>2</sub> for 1 hour at 4°C. Tritium labeled CHO selectin expressing cells were added and binding quantitated as described in Example 4(c).

**C. Effects of Alteration of N-Linked Glycosylation Sites**

Constructs expressing three P-selectin ligand-IgG chimeras were constructed to examine the effects of N-linked glycosylation sites on selectin binding. These constructs had the following characteristics:

148.Fc	amino acids 42-189 of SEQ ID NO:2
Q70.148.Fc	amino acids 42-189 of SEQ ID NO:2, with the asparagine residue at position 111 of SEQ ID NO:2 replaced with a glutamine residue
H24.Q70.148.Fc	amino acids 42-189 of SEQ ID NO:2, with the asparagine residue at position 65 of SEQ ID NO:2 with a histidine residue and the asparagine residue at position 111 of SEQ ID NO:2 replaced with glutamine residue

These constructs are schematically represented in Fig. 6.





5 substantially less sulfated. Thus, the tyrosine residues of the anionic region of P-selectin ligand proteins are the major site of sulfation.

Removal of sulfate from P-selectin ligand protein substantially reduces its binding to P-selectin. The binding of 148.Fc treated with chlorate to P-selectin was examined. As shown in Fig. 11, inhibition of sulfation by chlorate treatment substantially reduced the amount of P-selectin ligand protein binding to P-selectin.

10 **E. Effects of C-terminal Deletions**

Several additional C-terminal deleted constructs were made as follows:

254.Fc amino acids 42-295 of SEQ ID NO:2

47.Fc amino acids 42-88 of SEQ ID NO:2

15 19.Fc amino acids 42-60 of SEQ ID NO:2

These constructs are schematically represented in Fig. 12.

The binding of 254.Fc, 148.Fc, 47.Fc and 19.Fc to P-selectin, E-selectin and L-selectin was tested. Figs. 23 and 24 compare the binding of these deletion chimeras to selectins and controls. Results are also summarized in Fig. 12.

20 **F. Binding to P-selectin and E-selectin Expressing Cells**

Binding of various constructs described above to cells expressing P-selectin and E-selectin was compared using a quantitative plate binding assay of Example 4(c) (which is schematically described in Fig. 13).

Fig. 14 compares the binding of 148.Fc,  $\Delta$ Y.148.Fc, FFFE.148.Fc and human IgG1 to P-selectin expressing CHO cells. Deletion of all of the tyrosine residues in the anionic region in  $\Delta$ Y.148.Fc eliminated binding. Changing the tyrosine residues to phenylalanine residues in FFFE.148.Fc substantially reduced binding as compared to 148.Fc. Thus, it was demonstrated that the presence of the full length anionic region is essential to P-selectin binding and that P-selectin binding is enhanced by sulfation in this region. Fig. 14 also reports control experiments demonstrating that 148.Fc,  $\Delta$ Y.148.Fc and FFFE.148.Fc do not bind to CHO cells which do not express selectin.

Fig. 15 compares the binding of 148.Fc,  $\Delta$ Y.148.Fc, FFFE.148.Fc and human IgG1 to E-selectin expressing CHO cells. E-selectin binding was unaffected by the deletions or alterations of the native sequence. Thus, it was demonstrated that the anionic region is not required for E-selectin binding.

Fig. 16 summarizes the results of Figs. 14 and 15.

5 Fig. 17 compares the binding of 47.Fc to P- and E-selectin expressing CHO cells. 47.Fc demonstrated substantial binding to both selectins despite deletion of the N-linked glycosylation sites at positions 111 and 292 of SEQ ID NO:2.

Fig. 19 compares the binding of FYYD.19.Fc, FFFD.19.Fc, H24.Q70.148.Fc, 148.Fc, and human IgG1 to P-selectin expressing CHO cells. Replacement of all of the tyrosine residues  
10 in the anionic region in FFFD.19.Fc eliminated binding. Changing the tyrosine residue at position 46 to a phenylalanine residue in FYYD.19.Fc substantially reduced binding as compared to 148.Fc. Alteration of the N-linked glycosylation sites in H24.Q70.148.Fc did not affect binding. Thus, it was demonstrated that P-selectin binding is enhanced by sulfation in the anionic region and that N-linked glycosylation is not required for P-selectin binding. Fig. 19 also reports control  
15 experiments demonstrating that FYYD.19.Fc, FFFD.19.Fc, H24.Q70.148.Fc and 148.Fc do not bind to CHO cells which do not express selectin more than human IgG1 alone.

Fig. 20 compares the binding of FYYD.19.Fc, FFFD.19.Fc, H24.Q70.148.Fc, 148.Fc, and human IgG1 to E-selectin expressing CHO cells. Truncation of the ligand protein to the degree of FYYD.19.Fc and FFFD.19.Fc substantially reduced E-selectin binding. Alteration of  
20 the N-linked glycosylation sites in H24.Q70.148.Fc did not significantly affect E-selectin binding. Thus, it was demonstrated that P-selectin ligand proteins comprising amino acids 42 to 60 of SEQ ID NO:2 can selectively bind P-selectin, and, to a substantially less, extent E-selectin.

Fig. 21 summarizes the results of Figs. 19 and 20.

**G. Conclusions Regarding P- and E-Selectin Binding**

25 Although applicants do not wish to be bound by any theory, these data allow several conclusion regarding the relationship between P-selectin binding and E-selectin binding by P-selectin ligand proteins. N-linked carbohydrates are not required for binding of a P-selectin ligand protein to either P- or E-selectin. P-selectin ligand proteins as small comprising as little as amino acids 42-60 of SEQ ID NO:2 are capable of binding to P-selectin, and, to a substantially less,  
30 extent E-selectin.

Fig. 22 depicts a proposed schematic model for binding of P-selectin ligand proteins to P- and E-selectin. O-linked sLe<sup>x</sup> carbohydrate has been demonstrated to be required for both P- and E-selectin binding. Data presented herein demonstrate that sulfated tyrosine residues are implicated in P-selectin binding, but not E-selectin binding. Applicants' data also suggests that  
35 no N-linked glycosylation binding site is required.

5

**Example 11**  
**Examination of Aggregation Phenomena and Dimer Formation**  
**in Forms of PSGL-1**

A panel of PSGL-1 mutants were constructed by site-directed mutagenesis and/or PCR amplification with primers that introduced a stopcodon. The template for all mutagenesis experiments was pPL85.R16 (ATCC 75577, deposited by applicants).

The first group of mutants (C310S and C327S) encode full-length PSGL-1.R16 with only one amino acid change compared to wild-type PSGL-1.R16 (Cys to Ser at position 310 or 327, respectively). COS cells, co-transfected with pEA.3/4FT and the mutants C310S or C327S, were labeled with <sup>35</sup>S-methionine. Cell lysates were prepared and the mutant proteins were immunoprecipitated with the P-selectin ligand polyclonal antibody of Example 7(A) and analyzed by SDS-polyacrylamide gel electrophoresis under non-reducing and reducing conditions.

The mutant C327S as well as wild-type PSGL-1.R16 migrated as a homodimer under non-reducing conditions and as a monomer under reducing conditions. In contrast, the mutant C310S migrated as a monomer both under non-reducing and reducing conditions, indicating that the cysteine at position 310 is required for dimer formation of PSGL-1.

Both mutants were also analyzed for their ability to bind to P-selectin. Detergent extracts of co-transfected COS cells were precipitated with the LEC-γ1 chimera of Example 4(A). The precipitates were analyzed by SDS-PAGE under non-reducing and reducing conditions and by autoradiography. Both PSGL-1.R16 and C327S were efficiently precipitated by LEC-g1, whereas C310S binding to LEC-γ1 was greatly reduced, indicating that the dimeric form of PSGL-1 binds P-selectin more tightly than the monomeric form.

The second set of mutants encode soluble forms of PSGL-1.R16 and are listed in Table 1. The mutant ΔTM was generated by site-directed mutagenesis and has a deletion of the transmembrane domain (amino acids 313-333) followed by RLSRKA. The mutants L311, L312, A313, I314, L315, A318 and T322 were generated by site-directed mutagenesis or PCR amplification with PCR primers that introduced a stop codon in the desired position. The name of the mutant refers to the C-terminal amino acid of each truncated soluble form of PSGL-1.R16. The mutants were analyzed according to the following criteria:

1. Expression and secretion from transfected COS cells
2. Dimer versus monomer formation
3. Lack of aggregate formation
4. P-selectin binding (LEC-γ1 chimera)

5 The mutants  $\Delta$ TM and I316 fulfilled all four criteria. The shorter soluble forms of PSGL-1, such as sPSL.QC of Example 5(A), L311, L312, A313, I314 and L315 did not form dimers as well and the longer soluble forms of PSGL-1, such as sPSL.T7 of Example 5(C), A318 and T322 formed high molecular weight aggregates which were less desirable.

10 CHO cells, already expressing 3/4 fucosyltransferase and Core2 transferase, were transfected with psPSL.T7,  $\Delta$ TM, I316 or psPSL.QC and amplified using methotrexate. Stable clones were isolated and labeled with  $^{35}$ S-methionine. Conditioned media was either analyzed directly or first precipitated with LEC- $\gamma$ 1 and then analyzed by SDS-PAGE under non-reducing and reducing conditions (Figure 25). The results indicated that  $\Delta$ TM and I316 were most efficient in dimer formation and P-selectin binding.

15

Table I

20	Mutant	Dimer Formation	High MW Aggregates	P-selectin binding
	PSL.QC	+	-	+
	L311	+	-	+
25	L312	-	-	-
	A313	-	-	-
30	I314	-	-	-
	L315	+	-	+
	I316	++	-	++
35	A318	++	+	++
	T322	++	+	++
40	$\Delta$ TM	++	-	++

### Example 12

#### Specificity of PSGL-1 Binding to P- and E-Selectins

45 **Materials.** A chimeric protein comprising the extracellular domain of human E-selectin and the Fc portion of human IgG<sub>1</sub> was constructed analogously to the P-selectin chimera, LEC- $\gamma$ 1, described earlier. The soluble E-selectin chimera was expressed in baculovirus-infected

5 *Trichoplusia ni* high five cells (Invitrogen) and purified to homogeneity by Protein A Sepharose chromatography. Plasmid vectors pEA.3/4FT, pPL85, pFCD43, and pEA.sPACE, for COS expression of  $\alpha(1,3/1,4)$ -fucosyltransferase (Fuc-TIII), PSGL-1, CD43 (leukosialin), and soluble paired basic amino acid converting enzyme (PACE), respectively, have been described herein and in the literature (Sako et al. (1993) *Cell* 75, 1179-1186; Rehemtulla, A. & Kaufman, R. J. (1992) *Curr. Opin. Biotechnol.* 3, 560-565; Wasley et al. (1993) *J. Biol. Chem.* 268, 8458-8465).  
10 Fuc-TVII cDNA (plasmid pMT.FT7) was cloned from an HL60 cDNA expression library using oligonucleotide probes derived from the published sequence (Natsuka et al. (1994) *Journal of Biological Chemistry* 269, 16789-16794; Sasaki et al. (1994) *J. Biol. Chem.* 269, 14730-14737). A polyclonal neutralizing rabbit antibody, Rb3443, was raised against a peptide comprising the  
15 first 15 amino acids of the mature (PACE-cleaved) N-terminus of PSGL-1. Monoclonal anti-CD43 antibodies from either Becton Dickinson or Biodesign International and isotype control antibodies were coupled to a solid support consisting of Sepharose TM-4B with a covalently attached goat affinity-purified antibody to mouse IgG (Cappel, Organon Teknika Corporation). Affinity coupling of selectin chimeras and murine antibodies to Protein A Sepharose 4 Fast Flow  
20 (Pharmacia) and to the anti-mouse IgG resin, respectively, was carried out at a ratio of 2 mg protein/ml of resin. Antiserum Rb3443 was coupled to Protein A Sepharose at 1 ml/ml resin. Coupling efficiencies, indicated by micro-BCA assay (Pierce) of the post-reacted supernatants, were at least 95%. Aprotinin and pepstatin were from Boehringer Mannheim and benzamidine, leupeptin, and phenylmethylsulfonyl fluoride (PMSF) were from Sigma.

25 Labeling and Membrane Extraction of Myeloid Cells. U937 or HL60 cells grown in suspension to a density of  $\sim 1.3 \times 10^6$  cells/ml were labeled in 50 ml of RPMI1640 medium supplemented with 10% fetal bovine serum and 2.5 mCi of  $^3\text{H}$ -glucosamineHCl (Dupont/NEN) for 48 hr. Activities of greater than 1 cpm/cell were routinely obtained by this technique. The labeled cells were washed with PBS, resuspended in cell lysis buffer (10 mM MOPS, 150 mM  
30 NaCl, 4 mM  $\text{CaCl}_2$  and 4 mM  $\text{MgCl}_2$ , pH 7.5 containing protease inhibitors 20 mg/ml aprotinin, 10 mM benzamidine, 20 mg/ml leupeptin, 8 mg/ml pepstatin, and 10 mM PMSF) and subjected to several cycles of probe sonication on ice. Nuclei and cell debris were removed by low speed centrifugation and the cell membranes recovered from the supernatant by centrifugation at 100,000g RCF for 1 hr, washed by resuspension and high speed centrifugation in cell lysis buffer  
35 containing 1 M NaCl, and finally resuspended in 3 ml membrane solubilization buffer (cell lysis buffer containing 1% Triton X-100). Several cycles of sonication and incubation on ice were employed to solubilize the membrane fraction. Finally, a low speed centrifugation step was employed to remove insoluble membrane residue.

5            Labeling and Membrane Extraction of Transfected COS Cells. COS M6 cells were transfected using DEAE-dextran and chloroquine (25) employing 8 µg of plasmids pPL85 or pFCD43 and 4 µg of pEA.sPACE, as well as 4 µg of pEA.3/4FT or pMT.FT7. After 40-45 hr recovery the transfected cells were starved in serum-and methionine-free DME medium for 30 min and then fed [<sup>35</sup>S]-methionine in serum-free DME for 5 hr. The labeled cells were washed,  
10            incubated with EGTA to loosen them from the dish surface, scraped from the dish, pelleted, and suspended in cold 10mM PIPES buffer, pH 7.5, containing 100 mM KCl, 3 mM NaCl, 3.5 mM MgCl<sub>2</sub>, and protease inhibitors (see above). Membrane extraction then was carried out by sonication, low speed centrifugation, high speed centrifugation, and solubilization in membrane lysis buffer as above, for labeled myeloid cells.

15            Affinity Precipitations. Membrane extracts were diluted 1:4 or 1:5 with cell lysis buffer or with TBSC buffer (20 mM TrisHCl, 150 mM NaCl, 2 mM CaCl<sub>2</sub> pH 7.5) supplemented with 5 mg/ml bovine serum albumin (approximately 99%, Sigma). Extracts thus diluted to 0.2-0.25% Triton X-100 were incubated with human IgG<sub>1</sub>-Protein A Sepharose with end-over-end mixing at 4°C overnight. The precleared supernatants then were reacted for 6-12 hrs at 4°C with Protein  
20            A Sepharose precoupled with E-or P-selectin chimeras, control human IgG<sub>1</sub>, Rb3443 or with rabbit pre-immune serum or with anti-CD43 antibody or isotype control precoupled to goat anti-mouse IgG Sepharose. The resins were washed 5 or more times in buffer containing 0.1-0.5% Triton X-100 until the radioactivity of the wash supernatants was reduced to background level. Elution of proteins bound specifically to P-or E-selectin resins was accomplished with 10 mM  
25            EDTA or 5 mM EDTA/5 mM EGTA at room temperature or by boiling in SDS-PAGE sample buffer (Laemmli, U. K. (1970) *Nature* 227, 680-685), whereas elution of proteins bound to antibody resins was achieved exclusively by the latter means. For resolution under reducing conditions, dithiothreitol was added to the sample buffer to a final concentration of 100 mM. Samples thus prepared were resolved by SDS-PAGE on 7.5% gels, treated with En<sup>3</sup>Hance  
30            (Dupont), dried, and exposed to autoradiography film.

For sequential affinity capture experiments, membrane extracts were precleared, affinity precipitated with P-or E-selectin or human IgG<sub>1</sub>, and washed as above. Samples then were eluted twice from the resins with 5 mM EDTA in 10 mM MOPS, 150 mM NaCl, pH 7.5 for 1 hr at 4°C with tumbling. The first and second eluates were combined and then immunoprecipitated with  
35            immobilized Rb3443 according to the protocols outlined above.

### RESULTS:

Soluble E-and P-selectin chimeras were used, in parallel with control human IgG<sub>1</sub> to probe detergent-solubilized membrane extracts of <sup>3</sup>H-glucosamine-labeled U937 cells as described

5 under "Methods". Examination of eluates from the immobilized selectins by SDS-PAGE/autoradiography (Fig. 26) revealed the presence in both P-and E-selectin eluates of a major protein species with identical electrophoretic properties: Mr 200-kDa non-reduced with conversion to a species of Mr 120-kDa following reduction (Fig. 26, lanes 2 and 3, respectively). Occasionally, additional bands were observed in both E-and P-selectin eluates presumably  
10 corresponding to this major band and reflecting the presence of naturally reduced material (the 120-kDa species in non-reduced samples) and incomplete reduction (the 200-kDa species in reduced samples). Additionally, a trace band of Mr 150-kDa in the E-selectin eluate which was unaffected by reduction with DTT was occasionally observed. No bands were observed in control experiments using immobilized human IgG<sub>1</sub> (Fig. 26, lane 1) or where elution of selectin resins  
15 was performed in the absence of EDTA or SDS (data not shown). Essentially identical results were obtained using HL-60 cells (data not shown). Hence, the nature of these recognition events is interpreted to be specific metal-dependant interactions of these proteins with the respective selectins, presumably via the lectin domains (Lasky, L. A. (1992) *Science* 258, 964-969; Drickamer, K. (1988) *J. Biol. Chem.* 263, 9557).

20 The metal-dependant recognition and electrophoretic behavior of the major band precipitated with E-selectin was consistent with the properties of the previously identified P-selectin counterreceptor, P-selectin glycoprotein ligand or PSGL-1 (Moore et al. (1994) *J. Biol. Chem.* 269, 23318-23327; Moore et al. (1992) *J. Cell Biol.* 118, 445-456; Sako et al.).

To assess whether this species was indeed PSGL-1, EDTA eluates of the both E-and  
25 P-selectin precipitates were subsequently reacted with the PSGL-1 specific polyclonal antiserum Rb3443. As shown in Fig. 27, the major band isolated by affinity capture with either selectin was immunoprecipitated using this antiserum (lanes 3 and 4, respectively). No species were detected after immunoprecipitation of the control IgG<sub>1</sub> EDTA eluate (Fig. 27, lane 5). Direct immunoprecipitations using fresh <sup>3</sup>H-labeled U937 membrane extracts confirmed the specificity  
30 of Rb3443: precipitation with Rb3443 results in the recovery of a single band with the electrophoretic properties of PSGL-1 (Mr 200-kDa non-reduced, Mr 120-kDa reduced; Fig. 27, lane 2) whereas precipitation with pre-immune antiserum fails to capture any material (Fig. 27, lane 1). These results indicate that the major protein species specifically captured from myeloid cells by both E-and P-selectins is PSGL-1.

35 To further assess the specificity of E-selectin for PSGL-1, U937 membrane lysates were probed directly for the presence of CD43 (or leukosialin), an abundant cell surface sialoglycoprotein known to bear the major portion of myeloid cell SLe<sup>x</sup> residues (Maemura, K. & Fukuda, M. (1992) *J. Biol. Chem.* 267, 24379-24386). Thus, membrane extracts of



5 <sup>3</sup>H-glucosamine-labeled U937 cells were probed with an anti-CD43 antibody in parallel with the  
PSGL-1 specific antiserum Rb3443 and control antibodies as described under "Methods". From  
identical quantities of membrane lysate, the CD43 antibody precipitated in excess of 30-fold  
greater radioactive counts than did the PSGL-1 antiserum. Evaluation of the immunoprecipitates  
by SDS-PAGE/autoradiography (Fig. 28) revealed a single specific band for each antibody.  
10 Rb3443 captured a single species with the electrophoretic characteristics of PSGL-1 (Fig. 28,  
Lane 2). In contrast, the CD43 antibody precipitated a species with an Mr 120-kDa which was  
insensitive to reduction (Fig. 28, Lane 4), consistent with the absence of cysteine in CD43.  
Immunoprecipitations with control antibodies (Fig. 28, lanes 1 and 3) proved negative as  
expected. There appears to be considerably greater quantities of CD43 than PSGL-1 in U937  
15 cells, consistent with the quantitation of these proteins in HL-60 cells (Ushiyama et al. (1993) *J.*  
*Biol. Chem.* 268, 15229-15237). Thus, the inability of E-selectin to precipitate CD43 from  
myeloid cells does not appear to be due to its absence in these cell lines. While we cannot exclude  
the possibility that E-selectin captures trace quantities of CD43 (ie., the low-intensity Mr  
120-kDa band in Fig. 26, non-reduced lane 3 which is also consistent with monomeric PSGL-1),  
20 PSGL-1 appears to be the major protein precipitated from myeloid membrane extracts.

Recombinant PSGL-1 expressed in COS cells is best achieved with cotransfection of the  
PSGL-1 cDNA with a cDNA encoding an  $\alpha(1,3/1,4)$ fucosyltransferase (Fuc-TIII) for P-selectin  
binding (Sako et al.). Interestingly, initial efforts to demonstrate E-selectin recognition of  
recombinant PSGL-1 failed: E-selectin was unable to capture the counterreceptor from  
25 cotransfected COS cell membrane lysates under conditions where P-selectin capture was  
successful. One interpretation of this result is that Fuc-TIII was able to modify recombinant  
PSGL-1 for recognition by P-selectin but was unable to replicate the appropriate modification(s)  
found in myeloid PSGL-1 necessary for E-selectin recognition. The recent cloning of a myeloid  
fucosyltransferase, Fuc-TVII (Natsuka et al. (1994) *Journal of Biological Chemistry* 269, 16789-  
30 16794; Sasaki et al. (1994) *J. Biol. Chem.* 269, 14730-14737), that is also capable of generating  
SLe<sup>x</sup> carbohydrate structures, allowed evaluation of this interpretation.

COS cells were cotransfected with cDNAs encoding either PSGL-1 or CD43 and either  
Fuc-TIII or Fuc-TVII. Membrane lysates were prepared from the transfected COS cells and these  
were precipitated with either immobilized E- or P-selectin chimeras or with antibodies to either  
35 PSGL-1 or to CD43. The precipitated products were evaluated by SDS-PAGE/autoradiography  
following their release by EDTA/EGTA (for selectin mediated binding) or by boiling in SDS (for  
immunoprecipitations). The results are shown in Fig. 29.

5 As observed in Fig. 29A, E-selectin capture of COS-expressed PSGL-1 was dependant upon the nature of the fucosyltransferase used in the transfection. In three separate experiments, Fuc-TVII, but not Fuc-TIII, supported PSGL-1 precipitation by E-selectin. The inability of Fuc-TIII to confer E-selectin reactivity to PSGL-1 cannot be attributed to a lack of PSGL-1 expression as the specific antiserum Rb3443 immunoprecipitated significant and comparable quantities of PSGL-1 from both Fuc-TIII and Fuc-TVII transfections (Fig 29C). Furthermore, P-selectin was capable of precipitating equivalent quantities of PSGL-1 with either fucosyltransferase (Fig. 29B), demonstrating that Fuc-TIII and Fuc-TVII are expressed and active in these cotransfections.

15 Within the COS recombinant expression system as in myeloid cells, high-affinity E-selectin recognition was also dependent upon the presence of an appropriate polypeptide. Although the polypeptide length, apparent molecular weight, and high frequency and specific types of posttranslational modifications are similar in CD43 and PSGL-1 (Maemura, K. & Fukuda, M. (1992) *J. Biol. Chem.* 267, 24379-24386), neither fucosyltransferase was able to confer high-affinity E-selectin (or P-selectin) recognition to recombinant leukosialin in cotransfected COS cells (Fig 29A). Immunoprecipitations with the anti-CD43 antibody indicate that comparable quantities of leukosialin were expressed in both Fuc-TIII and Fuc-TVII cotransfections (Fig. 29C). The failure of E-selectin to capture CD43 was not due to lack of fucosyltransferase activity within the cotransfected COS cells. FACS analysis of COS cells transfected with either PSGL-1 or CD43 and either Fuc-TIII or Fuc-TVII all show high levels of reactivity with the SLeX specific antibody CSLEX-1. Therefore, these results suggest that high-affinity E-selectin recognition requires the presence of a specific polypeptide(s) that is appropriately modified by a specific fucosyltransferase.

### Example 13

#### Inhibition of P-Selectin/PSGL-1 Binding

##### by PSGL-1 Derived Peptides

30 A number of peptides derived from the sequence of PSGL-1 (SEQ ID NO:2) were tested for their ability to inhibit P-selectin/PSGL-1 binding. The tested peptides are listed in Fig. 30.

35 Inhibition was tested according to the following protocol. The wells of a 96 well plate were coated overnight at 4°C with PSGL-1 in 50 µl of 10 mM MOPS, 150 mM NaCl, 1 mM CaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub> at pH 7.5. After removal of the liquid from the wells, 150 µl of 10 mM MOPS, 150 mM NaCl, 1 mM CaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub>, 0.05% tween-20, 0.05% gelatin at pH 7.5 was added per well to block the unoccupied sites. After 1/2 hr. to 2 hr. the block buffer was removed

5 from the wells and 100  $\mu$ l of a complex of Lec- $\gamma$ 1 (P-selectin-human IgG Fc chimera)(2  $\mu$ g/ml),  
biotinylated goat anti-human antibody, and streptavidin-conjugated alkaline phosphatase (which  
had been allowed to tumble at room temperature for 30 minutes to 1 hour) plus any potential  
inhibitors were added per well. Each plate was shaken and rapped to remove the block from plate.  
The incubation proceeded for 1 hr at room temperature rotating in the dark. The unbound  
10 complex was washed off the plate with two 150  $\mu$ l portions of 10 mM MOPS, 150 mM NaCl,  
1 mM CaCl<sub>2</sub>, 1 mM MgCl<sub>2</sub>, 0.05% tween-20 followed by 150  $\mu$ l of 1M diethanolamine, 0.5 mM  
MgCl<sub>2</sub>. The chromogenic substrate for alkaline phosphatase, PNPP, in 10 mM DEA/0.5 mM  
MgCl<sub>2</sub> was added and the plate is then read at 405 nm.

The results of these assays are reported in Fig. 30. The peptides comprising amino acids  
15 48-51 (in which the tyrosine residues have been phosphorylated) and amino acids 42-56 of SEQ  
ID NO:2 provided particularly desirable results.

#### Example 14

##### Purification of a Soluble Form of PSGL-1

20 Substantial purification of a soluble form of P-selectin ligand protein has been achieved  
according to the protocol described below.

A soluble P-selectin ligand protein, I316 (amino acid 42 to amino acid 316 of SEQ ID  
NO:2) was expressed in CHO cells as described herein. CHO cell conditioned media was  
concentrated with a Pellicon ultrafiltration membrane unit (Millipore) with either 10,000  
25 molecular weight cutoff (MWCO) or 30,000 MWCO to about 10 times the original concentration.  
The buffer was then exchanged into 25 mM Tris, 1mM CaCl<sub>2</sub>, pH 7.4.

The buffer-exchanged concentrate was loaded onto a Toyopearl QAE 550C (TosoHaas)  
column. Alternatively, the buffer exchange step can be omitted and the concentrate can be diluted  
one part concentrate to three parts 25 mM Tris, 1mM CaCl<sub>2</sub>, pH 7.4, and then loaded onto the  
30 column. The column was washed with 5-10 column volumes (CV) of 25 mM Tris, 1mM CaCl<sub>2</sub>,  
pH 7.4 at 4°C.

The P-selectin ligand protein eluted from the column with a linear NaCl gradient (0 M  
NaCl to 1.0 M NaCl) in the 25 mM Tris, 1mM CaCl<sub>2</sub>, pH 7.4 buffer in approximately five column  
volumes. Two peaks were eluted from the column. The second peak contained the P-selectin  
35 ligand protein and was collected in bulk.

The peak from the QAE column was concentrated with a tangential flow ultrafiltration  
membrane (Millipore) with a 30,000 MWCO and was then buffer exchanged into 25 mM Tris,  
150 mM NaCl, 1mM CaCl<sub>2</sub>, pH 7.4 at 4°C.

5 The buffer exchanged concentrate was loaded onto a Jacalin Agarose column overnight at 4°C. The column was washed with the diafiltration buffer and the P-selectin ligand protein was eluted with a gradient of methyl  $\alpha$ -D-galactopyranoside (0-100 mM or )-50 mM methyl  $\alpha$ -D-galactopyranoside) at 20°C. Fractions from the Jacalin column were analyzed by SDS-PAGE and the purest fractions were pooled.

10

### Example 15

#### P-Selectin Ligand Protein Fusions

Four fusions of a P-selectin ligand protein with a different amino acid sequence were constructed: 47.Fc, 47.AGP, 47.BMP and 47.IL11.

15 47.Fc: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to a mutated Fc region of human IgG1 at His224 of the native Fc sequence. The sequence of the cDNA construct is reported as SEQ ID NO:35. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:36. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:36. The mutations in the Fc portion were a change of Leu 234 and Gly237 of the native Fc sequence to Ala.

20 47.AGP: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to the first leucine residue of mature human AGP. The sequence of the cDNA construct is reported as SEQ ID NO:37. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:38. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:38.

25 47.BMP: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to the sequence of mature human BMP-2 (with its first 8 amino acids deleted). The sequence of the cDNA construct is reported as SEQ ID NO:39. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as SEQ ID NO:40. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:40.

30 47.IL11: A cDNA was constructed encoding the signal peptide, PACE cleavage site and first 47 amino acids of the mature P-selectin ligand sequence fused to mature human IL-11. The sequence of the cDNA construct is reported as SEQ ID NO:41. The fusion point is a novel NotI site at nucleotide 261. The amino acid sequence encoded by the cDNA construct is reported as

- 5 SEQ ID NO:42. The mature amino acid sequence of the encoded fusion protein begins at amino acid 42 of SEQ ID NO:42.

Patent and literature references cited herein are incorporated as if fully set forth.

## SEQUENCE LISTING

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    - (A) MEDIUM TYPE: Floppy disk
    - (B) COMPUTER: IBM PC compatible
    - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
    - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
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    - (C) CLASSIFICATION:
  - (vii) PRIOR APPLICATION DATA:
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    - (B) FILING DATE: 23-OCT-1992
  - (vii) PRIOR APPLICATION DATA:
    - (A) APPLICATION NUMBER: US 08/112,608
    - (B) FILING DATE: 26-AUG-1993
  - (vii) PRIOR APPLICATION DATA:
    - (A) APPLICATION NUMBER: PCT/US93/10168
    - (B) FILING DATE: 22-OCT-1993
  - (vii) PRIOR APPLICATION DATA:
    - (A) APPLICATION NUMBER: US 08/235,398
    - (B) FILING DATE: 28-APR-1994
  - (vii) PRIOR APPLICATION DATA:
    - (A) APPLICATION NUMBER: US 08/316,305
    - (B) FILING DATE: 30-SEP-1994

(vii) PRIOR APPLICATION DATA:

- (A) APPLICATION NUMBER: US 08/428,734
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(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1649 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens
- (G) CELL TYPE: Promyelocyte
- (H) CELL LINE: HL60

(vii) IMMEDIATE SOURCE:

- (B) CLONE: PMT21:PL85

(ix) FEATURE:

- (A) NAME/KEY: 5'UTR
- (B) LOCATION: 1..59

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 60..1268

(ix) FEATURE:

- (A) NAME/KEY: 3'UTR
- (B) LOCATION: 1269..1649

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

```

GCCACTTCTT CTGGGCCAC GAGGCAGCTG TCCCATGCTC TGCTGAGCAC GGTGGTGCC      59
ATG CCT CTG CAA CTC CTC CTG TTG CTG ATC CTA CTG GGC CCT GGC AAC      107
Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
  1           5           10           15
AGC TTG CAG CTG TGG GAC ACC TGG GCA GAT GAA GCC GAG AAA GCC TTG      155
Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu
      20           25           30
    
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GGT CCC CTG CTT GCC CGG GAC CGG AGA CAG GCC ACC GAA TAT GAG TAC Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr 35 40 45	203
CTA GAT TAT GAT TTC CTG CCA GAA ACG GAG CCT CCA GAA ATG CTG AGG Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg 50 55 60	251
AAC AGC ACT GAC ACC ACT CCT CTG ACT GGG CCT GGA ACC CCT GAG TCT Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser 65 70 75 80	299
ACC ACT GTG GAG CCT GCT GCA AGG CGT TCT ACT GGC CTG GAT GCA GGA Thr Thr Val Glu Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly 85 90 95	347
GGG GCA GTC ACA GAG CTG ACC ACG GAG CTG GCC AAC ATG GGG AAC CTG Gly Ala Val Thr Glu Leu Thr Thr Glu Leu Ala Asn Met Gly Asn Leu 100 105 110	395
TCC ACG GAT TCA GCA GCT ATG GAG ATA CAG ACC ACT CAA CCA GCA GCC Ser Thr Asp Ser Ala Ala Met Glu Ile Gln Thr Thr Gln Pro Ala Ala 115 120 125	443
ACG GAG GCA CAG ACC ACT CCA CTG GCA GCC ACA GAG GCA CAG ACA ACT Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr 130 135 140	491
CGA CTG ACG GCC ACG GAG GCA CAG ACC ACT CCA CTG GCA GCC ACA GAG Arg Leu Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu 145 150 155 160	539
GCA CAG ACC ACT CCA CCA GCA GCC ACG GAA GCA CAG ACC ACT CAA CCC Ala Gln Thr Thr Pro Pro Ala Ala Thr Glu Ala Gln Thr Thr Gln Pro 165 170 175	587
ACA GGC CTG GAG GCA CAG ACC ACT GCA CCA GCA GCC ATG GAG GCA CAG Thr Gly Leu Glu Ala Gln Thr Thr Ala Pro Ala Ala Met Glu Ala Gln 180 185 190	635
ACC ACT GCA CCA GCA GCC ATG GAA GCA CAG ACC ACT CCA CCA GCA GCC Thr Thr Ala Pro Ala Ala Met Glu Ala Gln Thr Thr Pro Pro Ala Ala 195 200 205	683
ATG GAG GCA CAG ACC ACT CAA ACC ACA GCC ATG GAG GCA CAG ACC ACT Met Glu Ala Gln Thr Thr Gln Thr Thr Ala Met Glu Ala Gln Thr Thr 210 215 220	731
GCA CCA GAA GCC ACG GAG GCA CAG ACC ACT CAA CCC ACA GCC ACG GAG Ala Pro Glu Ala Thr Glu Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu 225 230 235 240	779
GCA CAG ACC ACT CCA CTG GCA GCC ATG GAG GCC CTG TCC ACA GAA CCC Ala Gln Thr Thr Pro Leu Ala Ala Met Glu Ala Leu Ser Thr Glu Pro 245 250 255	827



AGT GCC ACA GAG GCC CTG TCC ATG GAA CCT ACT ACC AAA AGA GGT CTG Ser Ala Thr Glu Ala Leu Ser Met Glu Pro Thr Thr Lys Arg Gly Leu 260 265 270	875
TTC ATA CCC TTT TCT GTG TCC TCT GTT ACT CAC AAG GGC ATT CCC ATG Phe Ile Pro Phe Ser Val Ser Ser Val Thr His Lys Gly Ile Pro Met 275 280 285	923
GCA GCC AGC AAT TTG TCC GTC AAC TAC CCA GTG GGG GCC CCA GAC CAC Ala Ala Ser Asn Leu Ser Val Asn Tyr Pro Val Gly Ala Pro Asp His 290 295 300	971
ATC TCT GTG AAG CAG TGC CTG CTG GCC ATC CTA ATC TTG GCG CTG GTG Ile Ser Val Lys Gln Cys Leu Leu Ala Ile Leu Ile Leu Ala Leu Val 305 310 315 320	1019
GCC ACT ATC TTC TTC GTG TGC ACT GTG GTG CTG GCG GTC CGC CTC TCC Ala Thr Ile Phe Phe Val Cys Thr Val Val Leu Ala Val Arg Leu Ser 325 330 335	1067
CGC AAG GGC CAC ATG TAC CCC GTG CGT AAT TAC TCC CCC ACC GAG ATG Arg Lys Gly His Met Tyr Pro Val Arg Asn Tyr Ser Pro Thr Glu Met 340 345 350	1115
GTC TGC ATC TCA TCC CTG TTG CCT GAT GGG GGT GAG GGG CCC TCT GCC Val Cys Ile Ser Ser Leu Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala 355 360 365	1163
ACA GCC AAT GGG GGC CTG TCC AAG GCC AAG AGC CCG GGC CTG ACG CCA Thr Ala Asn Gly Gly Leu Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro 370 375 380	1211
GAG CCC AGG GAG GAC CGT GAG GGG GAT GAC CTC ACC CTG CAC AGC TTC Glu Pro Arg Glu Asp Arg Glu Gly Asp Asp Leu Thr Leu His Ser Phe 385 390 395 400	1259
CTC CCT TAGCTCACTC TGCCATCTGT TTTGGCAAGA CCCCACCTCC ACGGGCTCTC Leu Pro	1315
CTGGGCCACC CCTGAGTGCC CAGACCCCAA TCCACAGCTC TGGGCTTCCT CGGAGACCCC	1375
TGGGGATGGG GATCTTCAGG GAAGGAACTC TGGCCACCCA AACAGGACAA GAGCAGCCTG	1435
GGGCCAAGCA GACGGGCAAG TGGAGCCACC TCTTTCCTCC CTCCGCGGAT GAAGCCCAGC	1495
CACATTTTCAG CCGAGGTCCA AGGCAGGAGG CCATTTACTT GAGACAGATT CTCTCCTTTT	1555
TCCTGTCCCC CATCTTCTCT GGGTCCCTCT AACATCTCCC ATGGCTCTCC CCGCTTCTCC	1615
TGGTCACTGG AGTCTCCTCC CCATGTACCC AAGG	1649

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 402 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Pro Leu Gln Leu Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn  
 1 5 10 15  
 Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu  
 20 25 30  
 Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr  
 35 40 45  
 Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg  
 50 55 60  
 Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser  
 65 70 75 80  
 Thr Thr Val Glu Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly  
 85 90 95  
 Gly Ala Val Thr Glu Leu Thr Thr Glu Leu Ala Asn Met Gly Asn Leu  
 100 105 110  
 Ser Thr Asp Ser Ala Ala Met Glu Ile Gln Thr Thr Gln Pro Ala Ala  
 115 120 125  
 Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr  
 130 135 140  
 Arg Leu Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu  
 145 150 155 160  
 Ala Gln Thr Thr Pro Pro Ala Ala Thr Glu Ala Gln Thr Thr Gln Pro  
 165 170 175  
 Thr Gly Leu Glu Ala Gln Thr Thr Ala Pro Ala Ala Met Glu Ala Gln  
 180 185 190  
 Thr Thr Ala Pro Ala Ala Met Glu Ala Gln Thr Thr Pro Pro Ala Ala  
 195 200 205  
 Met Glu Ala Gln Thr Thr Gln Thr Thr Ala Met Glu Ala Gln Thr Thr  
 210 215 220  
 Ala Pro Glu Ala Thr Glu Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu  
 225 230 235 240  
 Ala Gln Thr Thr Pro Leu Ala Ala Met Glu Ala Leu Ser Thr Glu Pro

	245		250		255
Ser	Ala Thr	Glu Ala Leu Ser Met	Glu Pro Thr Thr	Lys Arg Gly	Leu
	260		265		270
Phe	Ile Pro	Phe Ser Val Ser Ser	Val Thr His Lys	Gly Ile Pro	Met
	275		280		285
Ala	Ala Ser	Asn Leu Ser Val Asn Tyr	Pro Val Gly	Ala Pro Asp	His
	290		295		300
Ile	Ser Val	Lys Gln Cys Leu Leu	Ala Ile Leu	Ile Leu Ala	Leu Val
305		310		315	320
Ala	Thr Ile	Phe Phe Val Cys Thr	Val Val Leu	Ala Val Arg	Leu Ser
		325		330	335
Arg	Lys Gly	His Met Tyr Pro Val	Arg Asn Tyr Ser	Pro Thr Glu	Met
		340		345	350
Val	Cys Ile	Ser Ser Leu Leu Pro	Asp Gly Gly	Glu Gly Pro	Ser Ala
	355		360		365
Thr	Ala Asn	Gly Gly Leu Ser Lys	Ala Lys Ser	Pro Gly Leu	Thr Pro
	370		375		380
Glu	Pro Arg	Glu Asp Arg Glu Gly	Asp Asp Leu Thr	Leu His Ser	Phe
385		390		395	400
Leu	Pro				

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1239 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA (synthetic)

- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Homo sapiens
  - (G) CELL TYPE: placenta

- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 1..1239

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATG CCT CTG CAA CTC CTC CTG TTG CTG ATC CTA CTG GGC CCT GGC AAC



CAA ACC ACA GCC ATG GAG GCA CAG ACC ACT GCA CCA GAA GCC ACG GAG Gln Thr Thr Ala Met Glu Ala Gln Thr Thr Ala Pro Glu Ala Thr Glu 225 230 235 240	720
GCA CAG ACC ACT CAA CCC ACA GCC ACG GAG GCA CAG ACC ACT CCA CTG Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu 245 250 255	768
GCA GCC ATG GAG GCC CTG TCC ACA GAA CCC AGT GCC ACA GAG GCC CTG Ala Ala Met Glu Ala Leu Ser Thr Glu Pro Ser Ala Thr Glu Ala Leu 260 265 270	816
TCC ATG GAA CCT ACT ACC AAA AGA GGT CTG TTC ATA CCC TTT TCT GTG Ser Met Glu Pro Thr Thr Lys Arg Gly Leu Phe Ile Pro Phe Ser Val 275 280 285	864
TCC TCT GTT ACT CAC AAG GGC ATT CCC ATG GCA GCC AGC AAT TTG TCC Ser Ser Val Thr His Lys Gly Ile Pro Met Ala Ala Ser Asn Leu Ser 290 295 300	912
GTC AAC TAC CCA GTG GGG GCC CCA GAC CAC ATC TCT GTG AAG CAG TGC Val Asn Tyr Pro Val Gly Ala Pro Asp His Ile Ser Val Lys Gln Cys 305 310 315 320	960
CTG CTG GCC ATC CTA ATC TTG GCG CTG GTG GCC ACT ATC TTC TTC GTG Leu Leu Ala Ile Leu Ile Leu Ala Leu Val Ala Thr Ile Phe Val 325 330 335	1008
TGC ACT GTG GTG CTG GCG GTC CGC CTC TCC CGC AAG GGC CAC ATG TAC Cys Thr Val Val Leu Ala Val Arg Leu Ser Arg Lys Gly His Met Tyr 340 345 350	1056
CCC GTG CGT AAT TAC TCC CCC ACC GAG ATG GTC TGC ATC TCA TCC CTG Pro Val Arg Asn Tyr Ser Pro Thr Glu Met Val Cys Ile Ser Ser Leu 355 360 365	1104
TTG CCT GAT GGG GGT GAG GGG CCC TCT GCC ACA GCC AAT GGG GGC CTG Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala Thr Ala Asn Gly Gly Leu 370 375 380	1152
TCC AAG GCC AAG AGC CCG GGC CTG ACG CCA GAG CCC AGG GAG GAC CGT Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro Glu Pro Arg Glu Asp Arg 385 390 395 400	1200
GAG GGG GAT GAC CTC ACC CTG CAC AGC TTC CTC CCT TAG Glu Gly Asp Asp Leu Thr Leu His Ser Phe Leu Pro 405 410	1239

## (2) INFORMATION FOR SEQ ID NO:4:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 412 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Pro Leu Gln Leu Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn  
 1 5 10 15  
 Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu  
 20 25 30  
 Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr  
 35 40 45  
 Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg  
 50 55 60  
 Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser  
 65 70 75 80  
 Thr Thr Val Glu Pro Ala Ala Arg Arg Ser Thr Gly Leu Asp Ala Gly  
 85 90 95  
 Gly Ala Val Thr Glu Leu Thr Thr Glu Leu Ala Asn Met Gly Asn Leu  
 100 105 110  
 Ser Thr Asp Ser Ala Ala Met Glu Ile Gln Thr Thr Gln Pro Ala Ala  
 115 120 125  
 Thr Glu Ala Gln Thr Thr Gln Pro Val Pro Thr Glu Ala Gln Thr Thr  
 130 135 140  
 Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr Arg Leu Thr Ala Thr Glu  
 145 150 155 160  
 Ala Gln Thr Thr Pro Leu Ala Ala Thr Glu Ala Gln Thr Thr Pro Pro  
 165 170 175  
 Ala Ala Thr Glu Ala Gln Thr Thr Gln Pro Thr Gly Leu Glu Ala Gln  
 180 185 190  
 Thr Thr Ala Pro Ala Ala Met Glu Ala Gln Thr Thr Ala Pro Ala Ala  
 195 200 205  
 Met Glu Ala Gln Thr Thr Pro Pro Ala Ala Met Glu Ala Gln Thr Thr  
 210 215 220  
 Gln Thr Thr Ala Met Glu Ala Gln Thr Thr Ala Pro Glu Ala Thr Glu  
 225 230 235 240  
 Ala Gln Thr Thr Gln Pro Thr Ala Thr Glu Ala Gln Thr Thr Pro Leu  
 245 250 255  
 Ala Ala Met Glu Ala Leu Ser Thr Glu Pro Ser Ala Thr Glu Ala Leu  
 260 265 270

Ser Met Glu Pro Thr Thr Lys Arg Gly Leu Phe Ile Pro Phe Ser Val  
 275 280 285

Ser Ser Val Thr His Lys Gly Ile Pro Met Ala Ala Ser Asn Leu Ser  
 290 295 300

Val Asn Tyr Pro Val Gly Ala Pro Asp His Ile Ser Val Lys Gln Cys  
 305 310 315 320

Leu Leu Ala Ile Leu Ile Leu Ala Leu Val Ala Thr Ile Phe Phe Val  
 325 330 335

Cys Thr Val Val Leu Ala Val Arg Leu Ser Arg Lys Gly His Met Tyr  
 340 345 350

Pro Val Arg Asn Tyr Ser Pro Thr Glu Met Val Cys Ile Ser Ser Leu  
 355 360 365

Leu Pro Asp Gly Gly Glu Gly Pro Ser Ala Thr Ala Asn Gly Gly Leu  
 370 375 380

Ser Lys Ala Lys Ser Pro Gly Leu Thr Pro Glu Pro Arg Glu Asp Arg  
 385 390 395 400

Glu Gly Asp Asp Leu Thr Leu His Ser Phe Leu Pro  
 405 410

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2151 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cdNA

(vii) IMMEDIATE SOURCE:

- (B) CLONE: pacesol

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATGGAGCTGA GGCCCTGGTT GCTATGGGTG GTAGCAGCAA CAGGAACCTT GGTCTGCTA 60

GCAGCTGATG CTCAGGGCCA GAAGGTCTTC ACCAACACGT GGGCTGTGCG CATCCCTGGA 120

GGCCAGCGG TGGCCAACAG TGTGGCACGG AAGCATGGGT TCCTCAACCT GGGCCAGATC 180

TTCGGGGACT ATTACCACTT CTGGCATCGA GGAGTGACGA AGCGGTCCCT GTCGCCTCAC 240

CGCCCGCGGC ACAGCCGGCT GCAGAGGGAG CCTCAAGTAC AGTGGCTGGA ACAGCAGGTG 300

GCAAAGCGAC GGAATAACG GGACGTGTAC CAGGAGCCCA CAGACCCCAA GTTTCCTCAG 360  
 CAGTGGTACC TGTCTGGTGT CACTCAGCGG GACCTGAATG TGAAGGCGGC CTGGGCGCAG 420  
 GGCTACACAG GGCACGGCAT TGTGGTCTCC ATTCTGGACG ATGGCATCGA GAAGAACCAC 480  
 CCGGACTTGG CAGGCAATTA TGATCCTGGG GCCAGTTTTG ATGTCAATGA CCAGGACCCT 540  
 GACCCCCAGC CTCGGTACAC ACAGATGAAT GACAACAGGC ACGGCACACG GTGTGCGGGG 600  
 GAAGTGGCTG CCGTGGCCAA CAACGGTGTC TGTGGTGTAG GTGTGGCCTA CAACCCCCG 660  
 ATTGGAGGGG TGCGCATGCT GGATGGCGAG GTGACAGATG CAGTGGAGGC ACGCTCGCTG 720  
 GGCCTGAACC CCAACCACAT CCACATCTAC AGTGCCAGCT GGGGCCCCGA GGATGACGGC 780  
 AAGACAGTGG ATGGGCCAGC CCGCCTCGCC GAGGAGGCCT TCTTCCGTGG GGTAGCCAG 840  
 GGCCGAGGGG GGCTGGGCTC CATCTTTGTC TGGGCCTCGG GGAACGGGGG CCGGGAACAT 900  
 GACAGCTGCA ACTGCGACGG CTACACCAAC AGTATCTACA CGCTGTCCAT CAGCAGCGCC 960  
 ACGCAGTTTG GCAACGTGCC GTGGTACAGC GAGGCCTGCT CGTCCACACT GGCCACGACC 1020  
 TACAGCAGTG GCAACCAGAA TGAGAAGCAG ATCGTGACGA CTGACTTGCG GCAGAAGTGC 1080  
 ACGGAGTCTC ACACGGGCAC CTCAGCCTCT GCCCCCTTAG CAGCCGGCAT CATTGCTCTC 1140  
 ACCCTGGAGG CCAATAAGAA CCTCACATGG CGGGACATGC AACACCTGGT GGTACAGACC 1200  
 TCGAAGCCAG CCCACCTCAA TGCCAACGAC TGGGCCACCA ATGGTGTGGG CCGGAAAGTG 1260  
 AGCCACTCAT ATGGCTACGG GCTTTTGGAC GCAGGCGCCA TGGTGGCCCT GGCCAGAAT 1320  
 TGGACCACAG TGGCCCCCA GCGGAAGTGC ATCATCGACA TCCTCACCGA GCCCAAAGAC 1380  
 ATCGGGAAAC GGCTCGAGGT GCGGAAGACC GTGACCGCGT GCCTGGGCGA GCCCAACCAC 1440  
 ATCACTCGGC TGGAGCACGC TCAGGCGCGG CTCACCCTGT CCTATAATCG CCGTGGCGAC 1500  
 CTGGCCATCC ACCTGGTCAG CCCCATGGGC ACCCGCTCCA CCCTGCTGGC AGCCAGGCCA 1560  
 CATGACTACT CCGCAGATGG GTTTAATGAC TGGGCCTTCA TGACAACTCA TTCCTGGGAT 1620  
 GAGGATCCCT CTGGCGAGTG GGTCCCTAGAG ATTGAAAACA CCAGCGAAGC CAACAACATAT 1680  
 GGGACGCTGA CCAAGTTCAC CCTCGTACTC TATGGCACCG CCCCTGAGGG GCTGCCCGTA 1740  
 CCTCCAGAAA GCAGTGGCTG CAAGACCCTC ACGTCCAGTC AGGCCTGTGT GGTGTGCGAG 1800  
 GAAGGCTTCT CCCTGCACCA GAAGAGCTGT GTCCAGCACT GCCCTCCAGG CTTCGCCCCC 1860  
 CAAGTCCTCG ATACGCACTA TAGCACCGAG AATGACGTGG AGACCATCCG GGCCAGCGTC 1920  
 TGCGCCCCCT GCCACGCCTC ATGTGCCACA TGCCAGGGGC CGGCCCTGAC AGACTGCCTC 1980



AGCTGCCCCA GCCACGCCTC CTTGGACCCT GTGGAGCAGA CTTGCTCCCG GCAAAGCCAG 2040  
 AGCAGCCGAG AGTCCCCGCC ACAGCAGCAG CCACCTCGGC TGCCCCCGGA GGTGGAGGCG 2100  
 GGGCAACGGC TCGGGGCAGG GCTGCTGCCC TCACACCTGC CTGAGTGATG A 2151

## (2) INFORMATION FOR SEQ ID NO:6:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1587 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (ii) MOLECULE TYPE: cDNA

## (vi) ORIGINAL SOURCE:

- (A) ORGANISM: Homo sapiens

## (vii) IMMEDIATE SOURCE:

- (B) CLONE: sPSL.Fc

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATGCCTCTGC AACTCCTCCT GTTGCTGATC CTA CTACTGGGCC CTGGCAACAG CTTGCAGCTG 60  
 TGGGACACCT GGGCAGATGA AGCCGAGAAA GCCTTGGGTC CCCTGCTTGC CCGGGACCGG 120  
 AGACAGGCCA CCGAATATGA GTACCTAGAT TATGATTTC T GCCAGAAAC GGAGCCTCCA 180  
 GAAATGCTGA GGAACAGCAC TGACACCACT CCTCTGACTG GGCCTGGAAC CCCTGAGTCT 240  
 ACCACTGTGG AGCCTGCTGC AAGGCGTTCT ACTGGCCTGG ATGCAGGAGG GGCAGTCACA 300  
 GAGCTGACCA CGGAGCTGGC CAACATGGGG AACCTGTCCA CGGATTCAGC AGCTATGGAG 360  
 ATACAGACCA CTCAACCAGC AGCCACGGAG GCACAGACCA CTCCACTGGC AGCCACAGAG 420  
 GCACAGACAA CTCGACTGAC GGCCACGGAG GCACAGACCA CTCCACTGGC AGCCACAGAG 480  
 GCACAGACCA CTCCACCAGC AGCCACGGAA GCACAGACCA CTCAACCCAC AGGCCTGGAG 540  
 GCACAGACCA CTGCACCAGC AGCCATGGAG GCACAGACCA CTGCACCAGC AGCCATGGAA 600  
 GCACAGACCA CTCCACCAGC AGCCATGGAG GCACAGACCA CTCAAACCAC AGCCATGGAG 660  
 GCACAGACCA CTGCACCAGA AGCCACGGAG GCACAGACCA CTCAACCCAC AGCCACGGAG 720  
 GCACAGACCA CTCCACTGGC AGCCATGGAG GCCCTGTCCA CAGAACCAG TGCCACAGAG 780  
 GCCCTGTCCA TGGAACCTAC TACCAAAAGA GGTCTGTTCA TACCCTTTTC TGTGTCTCT 840  
 GTTACTCACA AGGGCATTCC CATGGCAGCC AGCAATTTGT CCGTCCTGCG GCCGCAGTCT 900

AGAGACAAAA CTCACACATG CCCACCGTGC CCAGCACCTG AACTCCTGGG GGGACCGTCA 960  
 GTCTTCCTCT TCCCCCAAA ACCCAAGGAC ACCCTCATGA TCTCCCGGAC CCCTGAGGTC 1020  
 ACATGCGTGG TGGTGGACGT GAGCCACGAA GACCCTGAGG TCAAGTTCAA CTGGTACGTG 1080  
 GACGGCGTGG AGGTGCATAA TGCCAAGACA AAGCCGCGGG AGGAGCAGTA CAACAGCACG 1140  
 TACCGTGTGG TCAGCGTCTT CACCGTCTTG CACCAGGACT GGCTGAATGG CAAGGAGTAC 1200  
 AAGTGCAAGG TCTCCAACAA AGCCCTCCCA GTCCCCATCG AGAAAACCAT CTCCAAGCC 1260  
 AAAGGGCAGC CCCGAGAACC ACAGGTGTAC ACCCTGCCCC CATCCCGGGA GGAGATGACC 1320  
 AAGAACCAGG TCAGCCTGAC CTGCCTGGTC AAAGGCTTCT ATCCAGCGA CATCGCCGTG 1380  
 GAGTGGGAGA GCAATGGGCA GCCGGAGAAC AACTACAAGA CCACGCCTCC CGTGCTGGAC 1440  
 TCCGACGGCT CCTTCTTCTT CTATAGCAAG CTCACCGTGG ACAAGAGCAG GTGGCAGCAG 1500  
 GGGAACGTCT TCTCATGCTC CGTGATGCAT GAGGCTCTGC ACAACCACTA CACGCAGAAG 1560  
 AGCCTCTCCC TGTCCCCGGG TAAATAG 1587

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 19 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

AATTCGTCG ACTCTAGAG

19

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 15 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

CTCTAGAGTC GACGG

15

## (2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 43 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

TAGCATACGC TCTAGAGCAT GGATCCCCTG GGTGCAGCCA AGC

43

## (2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 27 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CCGGAATTCT CAGGTGAACC AAGCCGC

27

## (2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 25 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAGTATCTGT CCAGGGCTTC CAGGT

25

## (2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 48 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

AACTACCCAG TGGGAGCACC AGACCACATC TCTGTGAAGC AGTGCTAG  
48

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 52 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

AATTCTAGCA CTGCTTCACA GAGATGTGGT CTGGTGCTCC CACTGGGTTAG TT  
52

(2) INFORMATION FOR SEQ ID NO:14:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 45 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

AACTACCCAG TGGGAGCACC AGACCACATC TCTGTGAAGC AGTAG  
45

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 49 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

AATTCTACTG CTTACACAGAG ATGTGGTCTG GTGCTCCCAC TGGGTAGTT

49

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 52 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CTAGACCCGG GATGGCATCC ATGACAGGAG GACAACAAAT GGTAGGCCGT AG

52

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 53 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

AATTCTACGG CCTACCCATT TGTTGTCCTC CTGTTCATGGA TGCCATCCCG GGT

53

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 13 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

CTGCGGCCGC AGT

13

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 17 base pairs
  - (B) TYPE: nucleic acid

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CTAGACTGCG GCCGCAG

17

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 50 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

CCAGGTCCAA CTGCAGGTCG ACTCTAGAGG GCACTTCTTC TGGGCCACG

50

(2) INFORMATION FOR SEQ ID NO:21:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 50 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

TATTATCTGT GCGGCCGCC TCCAGAACCC ATGGCTGCTG GTTGCAGTGG

50

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 40 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

TATTATCTGT GCGGCCGCGC AGCAGGCTCC ACAGTGGTAG

40

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 38 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

TATTATCTGT GCGGCCGCGG AGGCTCCGTT TCTGGCAG

38

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 33 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CGGAGACAGG CCACCGAATT CCTGCCAGAA ACG

33

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 40 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

CCTCCAGAAA TGCTGAGGCA CAGCACTGAC ACCACTCCTC

40

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 40 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

GAGCTGGCCA ACATGGGGCA ACTGTCCACG GATTCAGCAG

40

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 21 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

AATTCGAGTT CCTAGATTTT G

21

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 21 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

AATTCAAAT CTAGGAATC G

21

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 48 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA



(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

AATTCGAGTA CCTAGATTAT GATTCCTGC CAGAACTGA GCCTCCGC

48

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 48 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

GGCCGCGGAG GCTCAGTTTC TGGCAGGAAA TCATAATCTA GGTACTCG

48

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 48 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

AATTCGAGTT CCTAGATTAT GATTCCTGC CAGAACTGA GCCTCCGC

48

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 48 base pairs  
(B) TYPE: nucleic acid  
(C) STRANDEDNESS: single  
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

GGCCGCGGAG GCTCAGTTTC TGGCAGGAAA TCATAATCTA GGAACTCG

48

## (2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 48 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

AATTCGAGTT CCTAGATTTTC GATTTCTCTGC CAGAACTGA GCCTCCGC

48

## (2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 48 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

GGCCGCGGAG GCTCAGTTTC TGGCAGGAAA TCGAAATCTA GGAACCTCG

48

## (2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 942 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..939

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

ATGCCTCTGC AACTCCTCCT GTTGCTGATC CTACTGGGCC CTGGCAACAG CTGTCAGCTG 60

TGGGACACCT GGGCAGATGA AGCCGAGAAA GCCTTGGGTC CCCTGCTTGC CCGGGACCGG 120

AGACAGGCCA CCGAATATGA GTACCTAGAT TATGATTTCC TGCCAGAAAC GGAGCCTCCA 180  
 GAAATGCTGA GGAACAGCAC TGACACCACT CCTCTGACTG GGCCTGGAAC CCCTGAGTCT 240  
 ACCACTGTGG AGCCTGCTGC GCGGCCGCAC ACATGCCCCAC CGTGCCCAGC ACCTGAAGCC 300  
 CTGGGGGCAC CGTCAGTCTT CCTCTTCCCC CCAAACCCA AGGACACCCT CATGATCTCC 360  
 CGGACCCCTG AGGTCACATG CGTGGTGGTG GACGTGAGCC ACGAAGACCC TGAGGTCAAG 420  
 TTEAACTGGT ACGTGGACGG CGTGGAGGTG CATAATGCCA AGACAAAGCC GCGGGAGGAG 480  
 CAGTACAACA GCACGTACCG TGTGGTCAGC GTCCTCACCG TCCTGCACCA GGAAGTGGCTG 540  
 AATGGCAAGG AGTACAAGTG CAAGGTCTCC AACAAAGCCC TCCCAGTCCC CATCGAGAAA 600  
 ACCATCTCCA AAGCCAAAGG GCAGCCCCGA GAACCACAGG TGTACACCCT GCCCCCATCC 660  
 CGGGAGGAGA TGACCAAGAA CCAGGTCAGC CTGACCTGCC TGGTCAAAGG CTTCTATCCC 720  
 AGCGACATCG CCGTGGAGTG GGAGAGCAAT GGGCAGCCGG AGAACAACTA CAAGACCACG 780  
 CCTCCCGTGC TGGACTCCGA CGGCTCCTTC TTCCTCTATA GCAAGCTCAC CGTGGACAAG 840  
 AGCAGGTGGC AGCAGGGGAA CGTCTTCTCA TGCTCCGTGA TGCATGAGGC TCTGCACAAC 900  
 CACTACACGC AGAAGAGCCT CTCCTGTCC CCGGGTAAAT GA 942

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 313 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn  
 1 5 10 15  
 Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu  
 20 25 30  
 Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr  
 35 40 45  
 Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg  
 50 55 60

Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser  
 65 70 75 80  
 Thr Thr Val Glu Pro Ala Ala Arg Pro His Thr Cys Pro Pro Cys Pro  
 85 90 95  
 Ala Pro Glu Ala Leu Gly Ala Pro Ser Val Phe Leu Phe Pro Pro Lys  
 100 105 110  
 Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val  
 115 120 125  
 Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr  
 130 135 140  
 Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu  
 145 150 155 160  
 Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His  
 165 170 175  
 Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys  
 180 185 190  
 Ala Leu Pro Val Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln  
 195 200 205  
 Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu Met  
 210 215 220  
 Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro  
 225 230 235 240  
 Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn  
 245 250 255  
 Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu  
 260 265 270  
 Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val  
 275 280 285  
 Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln  
 290 295 300  
 Lys Ser Leu Ser Leu Ser Pro Gly Lys  
 305 310

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 810 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..807

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

```

ATGCCTCTGC AACTCCTCCT GTTGCTGATC CTA CTACTGGGCC CTGGCAACAG CTTGCAGCTG      60
TGGGACACCT GGGCAGATGA AGCCGAGAAA GCCTTGGGTC CCCTGCTTGC CCGGGACCGG      120
AGACAGGCCA CCGAATATGA GTACCTAGAT TATGATTTCC TGCCAGAAAC GGAGCCTCCA      180
GAAATGCTGA GGAACAGCAC TGACACCACT CCTCTGACTG GGCCTGGAAC CCCTGAGTCT      240
ACCACTGTGG AGCCTGCTGC GCGGCCGCTG TGTGCCAACC TAGTACCGGT GCCCATCACC      300
AACGCCACCC TGGACCAGAT CACTGGCAAG TGGTTTTATA TCGCATCGGC CTTTCGAAAC      360
GAGGAGTACA ATAAGTCGGT TCAGGAGATC CAAGCAACCT TCTTTTACTT CACCCCCAAC      420
AAGACAGAGG ACACGATCTT TCTCAGAGAG TACCAGACCC GACAGGACCA GTGCATCTAT      480
AACACCACCT ACCTGAATGT CCAGCGGGAA AATGGGACCA TCTCCAGATA CGTGGGAGGC      540
CAAGAGCATT TCGCTCACTT GCTGATCCTC AGGGACACCA AGACCTACAT GCTTGCTTTT      600
GACGTGAACG ATGAGAAGAA CTGGGGGCTG TCTGTCTATG CTGACAAGCC AGAGACGACC      660
AAGGAGCAAC TGGGAGAGTT CTACGAAGCT CTCGACTGCT TGCGCATTCC CAAGTCAGAT      720
GTCGTGTACA CCGATTGGAA AAAGGATAAG TGTGAGCCAC TGGAGAAGCA GCACGAGAAG      780
GAGAGGAAAC AGGAGGAGGG GGAATCCTAG      810
    
```

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 269 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

```

Met Pro Leu Gln Leu Leu Leu Leu Ile Leu Leu Gly Pro Gly Asn
1           5           10           15
    
```

Ser Leu Gln Leu Trp Asp Thr Trp Ala Asp Glu Ala Glu Lys Ala Leu  
 20 25 30

Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr  
 35 40 45

Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg  
 50 55 60

Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser  
 65 70 75 80

Thr Thr Val Glu Pro Ala Ala Arg Pro Leu Cys Ala Asn Leu Val Pro  
 85 90 95

Val Pro Ile Thr Asn Ala Thr Leu Asp Gln Ile Thr Gly Lys Trp Phe  
 100 105 110

Tyr Ile Ala Ser Ala Phe Arg Asn Glu Glu Tyr Asn Lys Ser Val Gln  
 115 120 125

Glu Ile Gln Ala Thr Phe Phe Tyr Phe Thr Pro Asn Lys Thr Glu Asp  
 130 135 140

Thr Ile Phe Leu Arg Glu Tyr Gln Thr Arg Gln Asp Gln Cys Ile Tyr  
 145 150 155 160

Asn Thr Thr Tyr Leu Asn Val Gln Arg Glu Asn Gly Thr Ile Ser Arg  
 165 170 175

Tyr Val Gly Gly Gln Glu His Phe Ala His Leu Leu Ile Leu Arg Asp  
 180 185 190

Thr Lys Thr Tyr Met Leu Ala Phe Asp Val Asn Asp Glu Lys Asn Trp  
 195 200 205

Gly Leu Ser Val Tyr Ala Asp Lys Pro Glu Thr Thr Lys Glu Gln Leu  
 210 215 220

Gly Glu Phe Tyr Glu Ala Leu Asp Cys Leu Arg Ile Pro Lys Ser Asp  
 225 230 235 240

Val Val Tyr Thr Asp Trp Lys Lys Asp Lys Cys Glu Pro Leu Glu Lys  
 245 250 255

Gln His Glu Lys Glu Arg Lys Gln Glu Glu Gly Glu Ser  
 260 265

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1314 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..1311

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

ATGGTGGCCG GGACCCGCTG TCTTCTAGCG TTGCTGCTTC CCCAGGTCCT CCTGGGCGGC	60
GCGGCTGGCC TCGTTCGGGA GCTGGGCCG AGGAAGTTCG CGGCGGCGTC GTCGGGCCGC	120
CCCTCATCCC AGCCCTCTGA CGAGGTCCTG AGCGAGTTCG AGTTGCGGCT GCTCAGCATG	180
TTCGGCCTGA AACAGAGACC CACCCCCAGC AGGGACGCCG TGGTGCCCCC CTACATGCTA	240
GACCTGTATC GCAGGCACTC AGGTCAGCCG GGCTEACCCG CCCCAGACCA CCGGTTGGAG	300
AGGGCAGCCA GCCGAGCCAA CACTGTGCGC AGCTTCCACC ATGAAGAATC TTTGGAAGAA	360
CTACCAGAAA CGAGTGGGAA AACAACCCGG AGATTCTTCT TTAATTTAAG TTCTATCCCC	420
ACGGAGGAGT TTATCACCTC AGCAGAGCTT CAGGTTTTCC GAGAACAGAT GCAAGATGCT	480
TTAGGAAAACA ATAGCAGTTT CCATCACCGA ATTAATATTT ATGAAATCAT AAAACCTGCA	540
ACAGCCAACT CGAAATTCCC CGTGACCAGA CTTTTGGACA CCAGGTTGGT GAATCAGAAT	600
GCAAGCAGGT GGGAAAGTTT TGATGTCACC CCCGCTGTGA TGCGGTGGAC TGCACAGGGA	660
CACGCCAACC ATGGATTTCGT GGTGGAAGTG GCCCACTTGG AGGAGAAACA AGGTGTCTCC	720
AAGAGACATG TTAGGATAAG CAGGTCTTTG CACCAAGATG AACACAGCTG GTCACAGATA	780
AGGCCATTGC TAGTAACTTT TGGCCATGAT GGAAAAGGGC ATCCTCTCCA CAAAAGAGAA	840
AAACGTCAGG CCACCGAATA TGAGTACCTA GATTATGATT TCCTGCCAGA AACGGAGCCT	900
CCAGAAATGC TGAGGAACAG CACTGACACC ACTCCTCTGA CTGGGCCTGG AACCCCTGAG	960
TCTACCACTG TGGAGCCTGC TGCAAGGCGG AAACGCCTTA AGTCCAGCTG TAAGAGACAC	1020
CCTTTGTACG TGGACTTCAG TGACGTGGGG TGGAACTACT GGATTGTGGC TCCCCGGGG	1080
TATCACGCCT TTTACTGCCA CGGAGAATGC CCTTTTCCTC TGGCTGATCA TCTGAACTCC	1140
ACTAATCATG CCATTGTTCA GACGTTGGTC AACTCTGTTA ACTCTAAGAT TCCTAAGGCA	1200
TGCTGTGTCC CGACAGAACT CAGTGCTATC TCGATGCTGT ACCTTGACGA GAATGAAAAG	1260
GTTGTATTAA AGAACTATCA GGACATGGTT GTGGAGGGTT GTGGGTGTCTG CTAG	1314

(2) INFORMATION FOR SEQ ID NO:40:





Gly Phe Val Val Glu Val Ala His Leu Glu Glu Lys Gln Gly Val Ser  
 225 230 235 240

Lys Arg His Val Arg Ile Ser Arg Ser Leu His Gln Asp Glu His Ser  
 245 250 255

Trp Ser Gln Ile Arg Pro Leu Leu Val Thr Phe Gly His Asp Gly Lys  
 260 265 270

Gly His Pro Leu His Lys Arg Glu Lys Arg Gln Ala Thr Glu Tyr Glu  
 275 280 285

Tyr Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu  
 290 295 300

Arg Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu  
 305 310 315 320

Ser Thr Thr Val Glu Pro Ala Ala Arg Arg Lys Arg Leu Lys Ser Ser  
 325 330 335

Cys Lys Arg His Pro Leu Tyr Val Asp Phe Ser Asp Val Gly Trp Asn  
 340 345 350

Asp Trp Ile Val Ala Pro Pro Gly Tyr His Ala Phe Tyr Cys His Gly  
 355 360 365

Glu Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr Asn His Ala  
 370 375 380

Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Lys Ile Pro Lys Ala  
 385 390 395 400

Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp  
 405 410 415

Glu Asn Glu Lys Val Val Leu Lys Asn Tyr Gln Asp Met Val Val Glu  
 420 425 430

Gly Cys Gly Cys Arg  
 435

## (2) INFORMATION FOR SEQ ID NO:41:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 795 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: CDS



Gly Pro Leu Leu Ala Arg Asp Arg Arg Gln Ala Thr Glu Tyr Glu Tyr  
 35 40 45

Leu Asp Tyr Asp Phe Leu Pro Glu Thr Glu Pro Pro Glu Met Leu Arg  
 50 55 60

Asn Ser Thr Asp Thr Thr Pro Leu Thr Gly Pro Gly Thr Pro Glu Ser  
 65 70 75 80

Thr Thr Val Glu Pro Ala Ala Arg Pro Pro Pro Gly Pro Pro Arg Val  
 85 90 95

Ser Pro Asp Pro Arg Ala Glu Leu Asp Ser Thr Val Leu Leu Thr Arg  
 100 105 110

Ser Leu Leu Ala Asp Thr Arg Gln Leu Ala Ala Gln Leu Arg Asp Lys  
 115 120 125

Phe Pro Ala Asp Gly Asp His Asn Leu Asp Ser Leu Pro Thr Leu Ala  
 130 135 140

Met Ser Ala Gly Ala Leu Gly Ala Leu Gln Leu Pro Gly Val Leu Thr  
 145 150 155 160

Arg Leu Arg Ala Asp Leu Leu Ser Tyr Leu Arg His Val Gln Trp Leu  
 165 170 175

Arg Arg Ala Gly Gly Ser Ser Leu Lys Thr Leu Glu Pro Glu Leu Gly  
 180 185 190

Thr Leu Gln Ala Arg Leu Asp Arg Leu Leu Arg Arg Leu Gln Leu Leu  
 195 200 205

Met Ser Arg Leu Ala Leu Pro Gln Pro Pro Pro Asp Pro Pro Ala Pro  
 210 215 220

Pro Leu Ala Pro Pro Ser Ser Ala Trp Gly Gly Ile Arg Ala Ala His  
 225 230 235 240

Ala Ile Leu Gly Gly Leu His Leu Thr Leu Asp Trp Ala Val Arg Gly  
 245 250 255

Leu Leu Leu Leu Lys Thr Arg Leu  
 260

CLAIMS

1. An isolated DNA encoding a fusion protein comprising (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second amino acid sequence derived from the sequence of a protein other than P-selectin ligand.
2. The DNA of claim 1 which further comprises an expression control sequence operably linked to said nucleotide sequence.
3. A host cell transformed with the DNA of claim 2.
4. A process for producing a fusion protein, which comprises:
  - (a) culturing the host cell of claim 3 under condition suitable for expression of the fusion protein; and
  - (b) purifying the fusion protein from the culture medium.
5. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 402 of SEQ ID NO:2.
6. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 310 of SEQ ID NO:2.
7. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 88 of SEQ ID NO:2.
8. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 118 of SEQ ID NO:2.
9. The DNA of claim 1 wherein said first amino acid sequence comprises amino acid 42 to amino acid 189 of SEQ ID NO:2.

10. The DNA of claim 1 wherein said second amino acid sequence is linked to the C-terminus of said first amino acid sequence.
11. The DNA of claim 10 wherein said sequences are linked by a linking sequence.
12. The DNA of claim 1 wherein said second amino acid sequence is joined to the N-terminus of said first amino acid sequence.
13. The DNA of claim 12 wherein said sequences are linked by a linking sequence.
14. The DNA of claim 1 wherein said second amino acid sequence is derived from a protein selected from the group consisting of an antibody, a cytokine, a growth factor, a differentiation factor, a hormone, an enzyme, a receptor or fragment thereof and a ligand.
15. The DNA of claim 14 wherein said second amino acid sequence is derived from the sequence of an antibody.
16. The DNA of claim 15 wherein said second amino acid sequence is derived from the Fc portion of an antibody.
17. The DNA of claim 15 wherein said second amino acid sequence is a mutation of a sequence derived from an antibody.
18. The DNA of claim 14 wherein said second amino acid sequence is derived from the sequence of a cytokine.
19. The DNA of claim 14 wherein said second amino acid sequence is derived from the sequence of a growth factor.
20. The DNA of claim 19 wherein said growth factor is a BMP.
21. The DNA of claim 7 comprising the nucleotide sequence of SEQ ID NO:35 from nucleotide 123 to nucleotide 939.

22. The DNA of claim 21 comprising the nucleotide sequence of SEQ ID NO:35.
23. The DNA of claim 7 comprising the nucleotide sequence of SEQ ID NO:37 from nucleotide 123 to nucleotide 807.
24. The DNA of claim 23 comprising the nucleotide sequence of SEQ ID NO:37.
25. The DNA of claim 7 comprising the nucleotide sequence of SEQ ID NO:39 from nucleotide 123 to nucleotide 1311.
26. The DNA of claim 25 comprising the nucleotide sequence of SEQ ID NO:39.
27. The DNA of claim 7 comprising the nucleotide sequence of SEQ ID NO:41 from nucleotide 123 to nucleotide 792.
28. The DNA of claim 27 comprising the nucleotide sequence of SEQ ID NO:41.
29. A fusion protein comprising (a) a first amino acid sequence comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second amino acid sequence derived from the sequence of a protein other than P-selectin ligand.
30. The fusion protein of claim 29 wherein said first amino acid sequence comprises amino acid 42 to amino acid 402 of SEQ ID NO:2.
31. The fusion protein of claim 29 wherein said first amino acid sequence comprises amino acid 42 to amino acid 310 of SEQ ID NO:2.
32. The fusion protein of claim 29 wherein said first amino acid sequence comprises amino acid 42 to amino acid 88 of SEQ ID NO:2.
33. The fusion protein of claim 29 wherein said first amino acid sequence comprises amino acid 42 to amino acid 118 of SEQ ID NO:2.

34. The fusion protein of claim 29 wherein said first amino acid sequence comprises amino acid 42 to amino acid 189 of SEQ ID NO:2.

35. The fusion protein of claim 29 wherein said second amino acid sequence is linked to the C-terminus of said first amino acid sequence.

36. The fusion protein of claim 35 wherein said sequences are linked by a linking sequence.

37. The fusion protein of claim 29 wherein said second amino acid sequence is joined to the N-terminus of said first amino acid sequence.

38. The fusion protein of claim 37 wherein said sequences are linked by a linking sequence.

39. The fusion protein of claim 29 wherein said second amino acid sequence is derived from a protein selected from the group consisting of an antibody, a cytokine, a growth factor, a differentiation factor, a hormone, an enzyme, a receptor or fragment thereof and a ligand.

40. The fusion protein of claim 39 wherein said second amino acid sequence is derived from the sequence of an antibody.

41. The fusion protein of claim 40 wherein said second amino acid sequence is derived from the Fc portion of an antibody.

42. The fusion protein of claim 40 wherein said second amino acid sequence is a mutation of a sequence derived from an antibody.

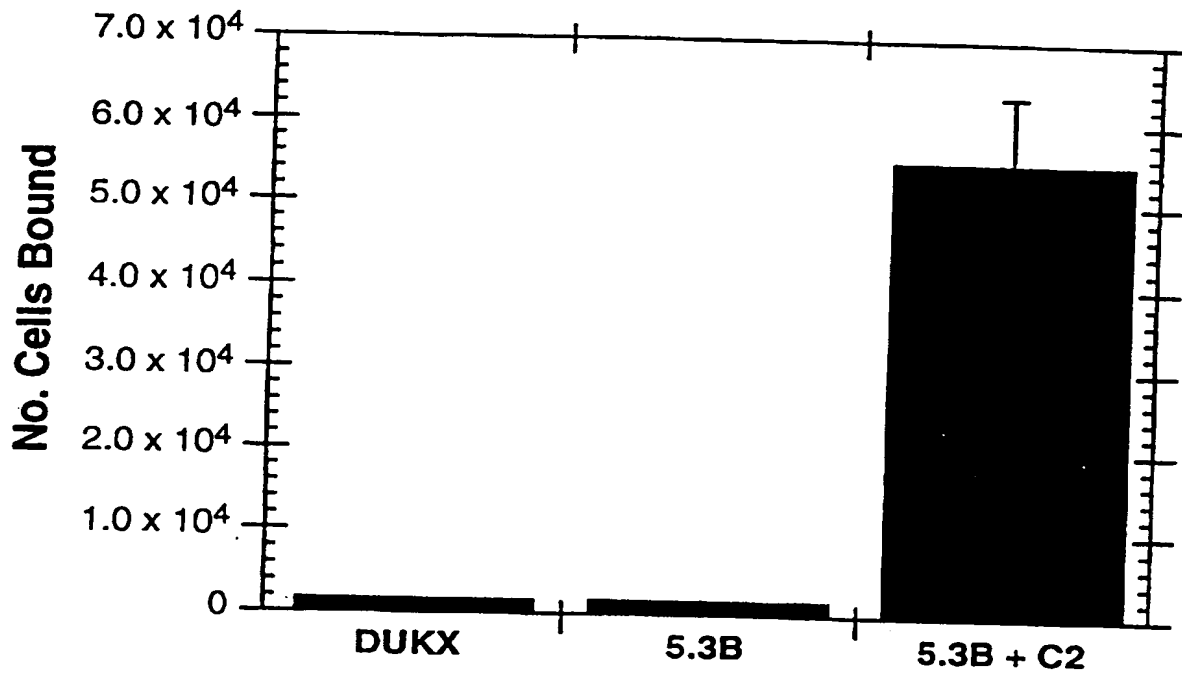
43. The fusion protein of claim 39 wherein said second amino acid sequence is derived from the sequence of a cytokine.

44. The fusion protein of claim 39 wherein said second amino acid sequence is derived from the sequence of a growth factor.
45. The fusion protein of claim 44 wherein said growth factor is a BMP.
46. The fusion protein of claim 32 comprising the amino acid sequence of SEQ ID NO:36 from amino acid 42 to amino acid 313.
47. The fusion protein of claim 46 comprising the amino acid sequence of SEQ ID NO:36.
48. The fusion protein of claim 32 comprising the amino acid sequence of SEQ ID NO:38 from amino acid 42 to amino acid 269.
49. The fusion protein of claim 48 comprising the amino acid sequence of SEQ ID NO:38.
50. The fusion protein of claim 32 comprising the amino acid sequence of SEQ ID NO:40 from amino acid 42 to amino acid 437.
51. The fusion protein of claim 50 comprising the amino acid sequence of SEQ ID NO:40.
52. The fusion protein of claim 32 comprising the amino acid sequence of SEQ ID NO:42 from amino acid 42 to amino acid 264.
53. The fusion protein of claim 52 comprising the amino acid sequence of SEQ ID NO:42.
54. A fusion protein made according to the process of claim 4.

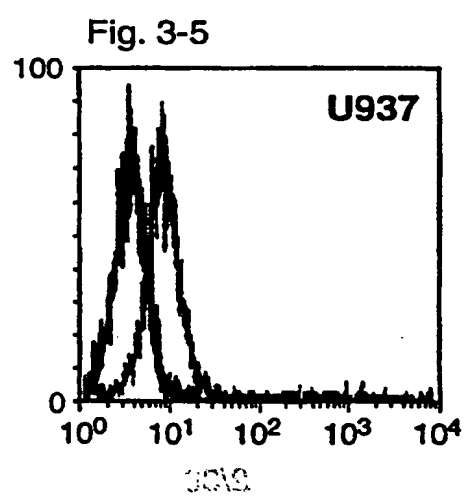
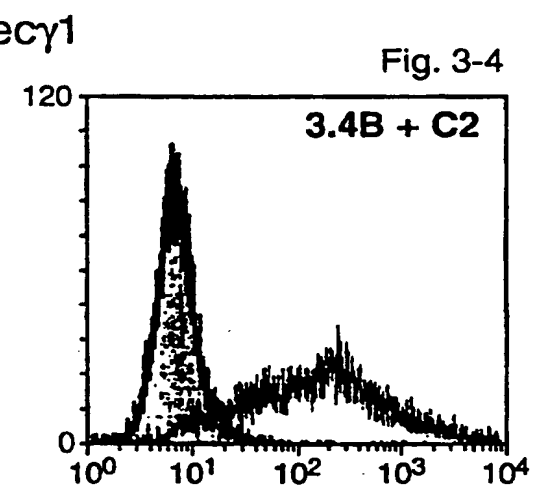
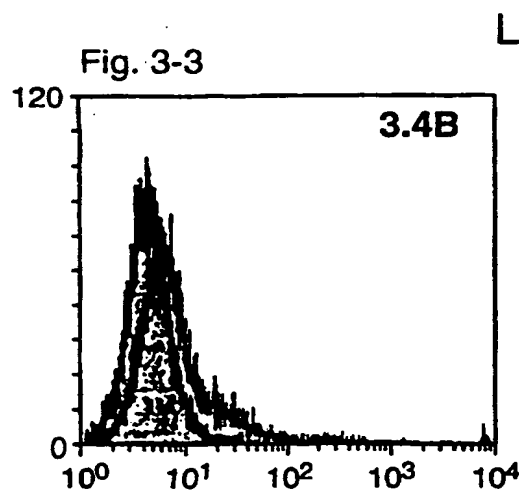
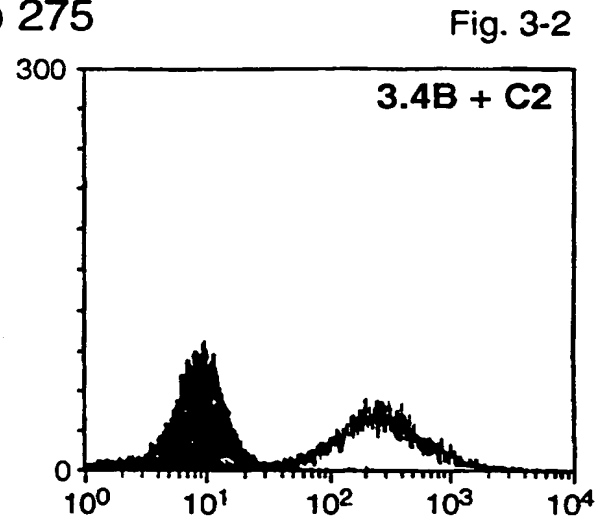
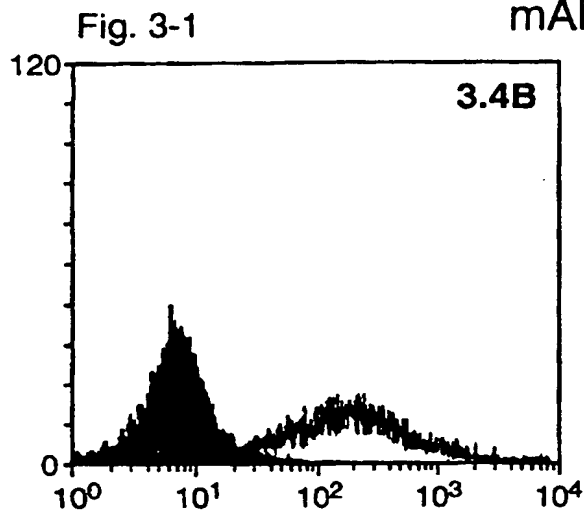


55. A composition comprising (a) a first peptide comprising amino acid 42 to amino acid 60 of SEQ ID NO:2, and (b) a second peptide derived from the sequence of a protein other than P-selectin ligand, wherein said first peptide and said second peptide are chemically linked by a moiety other than a peptide bond.

Fig. 1







3019

Fig. 4

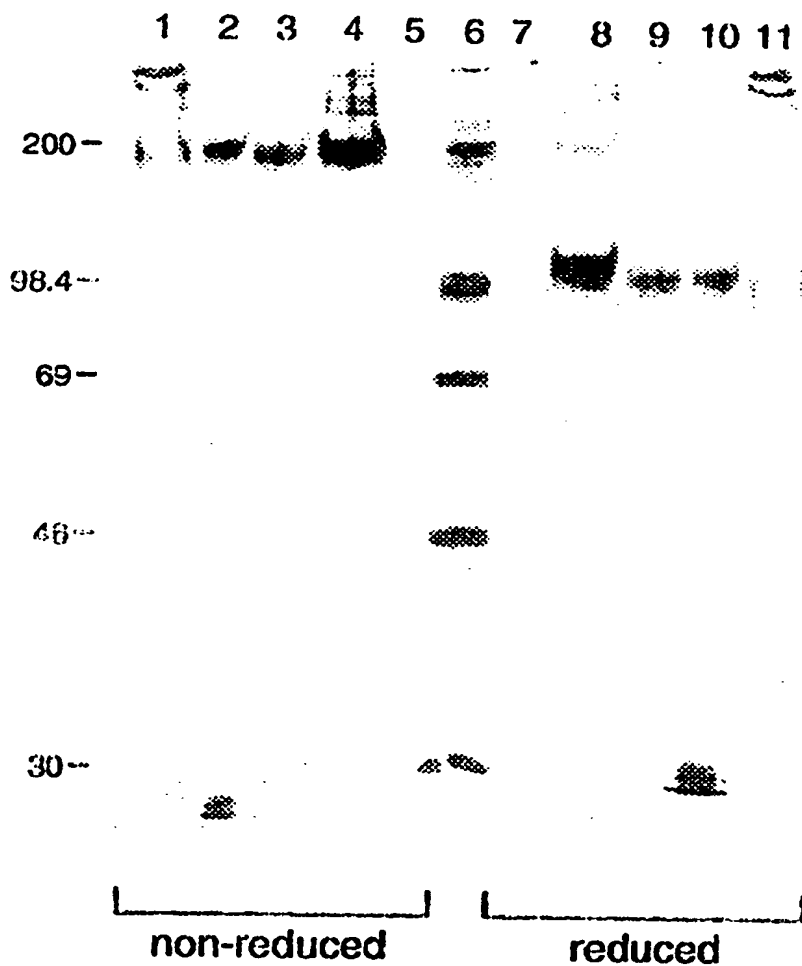
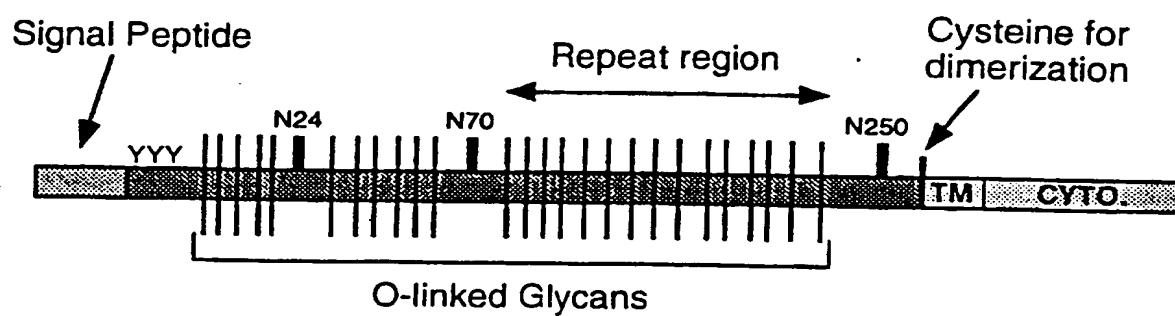


Fig. 5



**Soluble "T7" Truncated Form:**

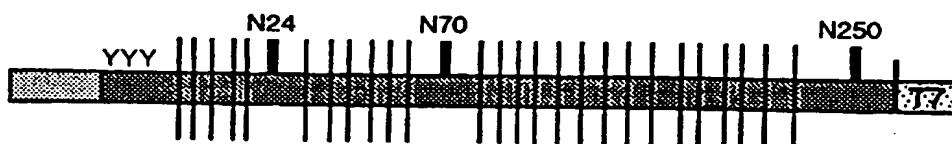


Fig. 6

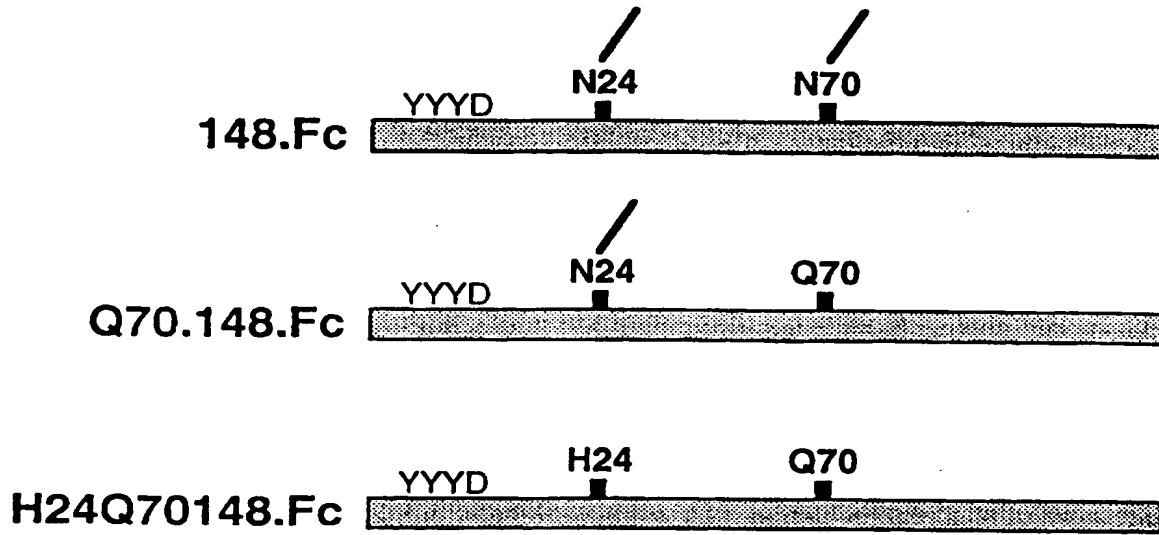


Fig. 7

1 2 3 4 5 6



**<sup>35</sup>S-Met Labeled**

*Captured with Protein A*

- 1. 148.Fc
- 2. 148.Q70.Fc
- 3. 148.H24.Q70.Fc

*Captured with P-Sel.Fc*

- 4. 148.Fc
- 5. 18.Q70.Fc
- 6. 148.H24.Q70.Fc

7/30

SUBSTITUTE SHEET (RULE 26)



Fig. 8

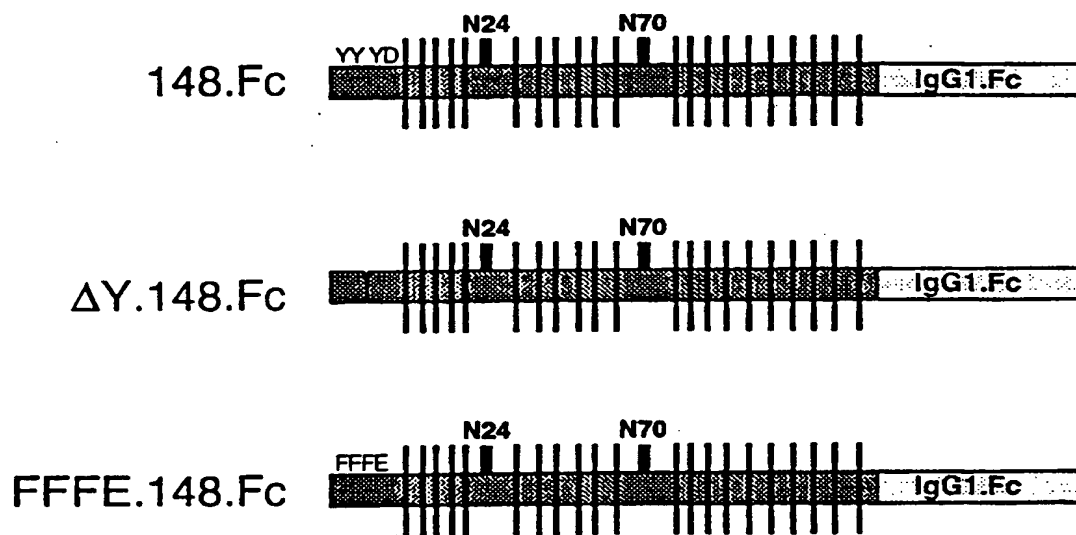
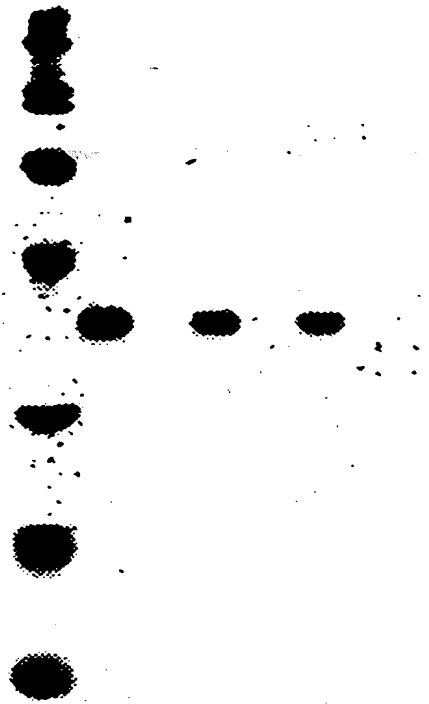


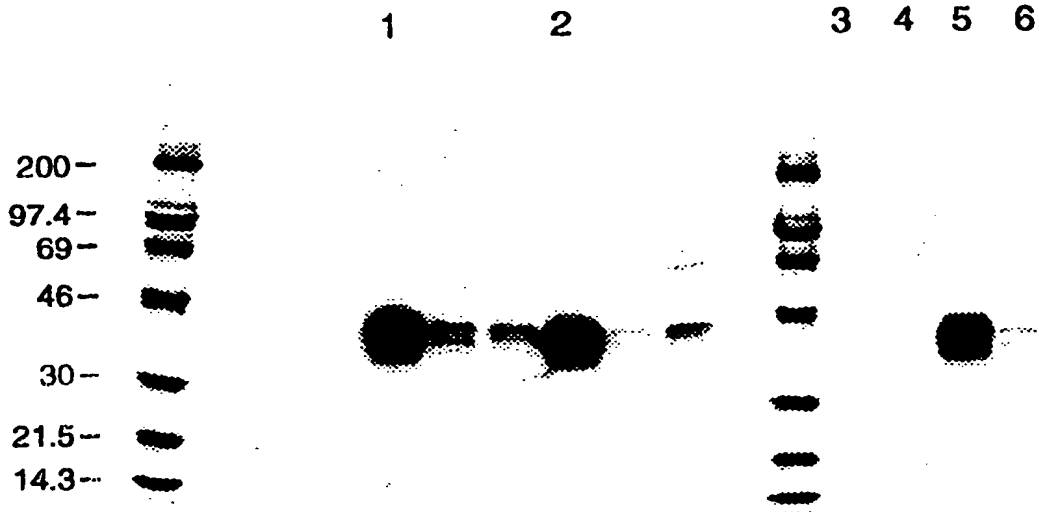
Fig. 9

M 1 2 3 4 5 6 7 8



<u>Sample</u>	<u>Clonotype</u>
M. mw marker	
1. YYY.19Fc	-
2. YYY.19Fc	+
3. FYY.19Fc	-
4. FYY.19Fc	+
5. FFY.19Fc	-
6. FFY.19Fc	+
7. FFF.19Fc	-
8. FFF.19Fc	+

Fig. 10



**<sup>35</sup>S-Met Labeled**

- 1. FYY.19.Fc
- 2. FFF.19.Fc

**<sup>35</sup>SO<sub>4</sub> Labeled**

- 3. Mock
- 4. P-Sel.LE.Fc
- 5. FYY.19.Fc
- 6. FFF.19.Fc

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Fig. 11

	<sup>35</sup> S-Met Labeled				<sup>35</sup> S-SO <sub>4</sub> Labeled			
	┌───────────┐				┌───────────┐			
Chlorate	-	-	+	+	-	-	+	+
P-Sel.Fc	-	+	-	+	-	+	-	+
Protein A	+	-	+	-	+	-	+	-



Fig. 12

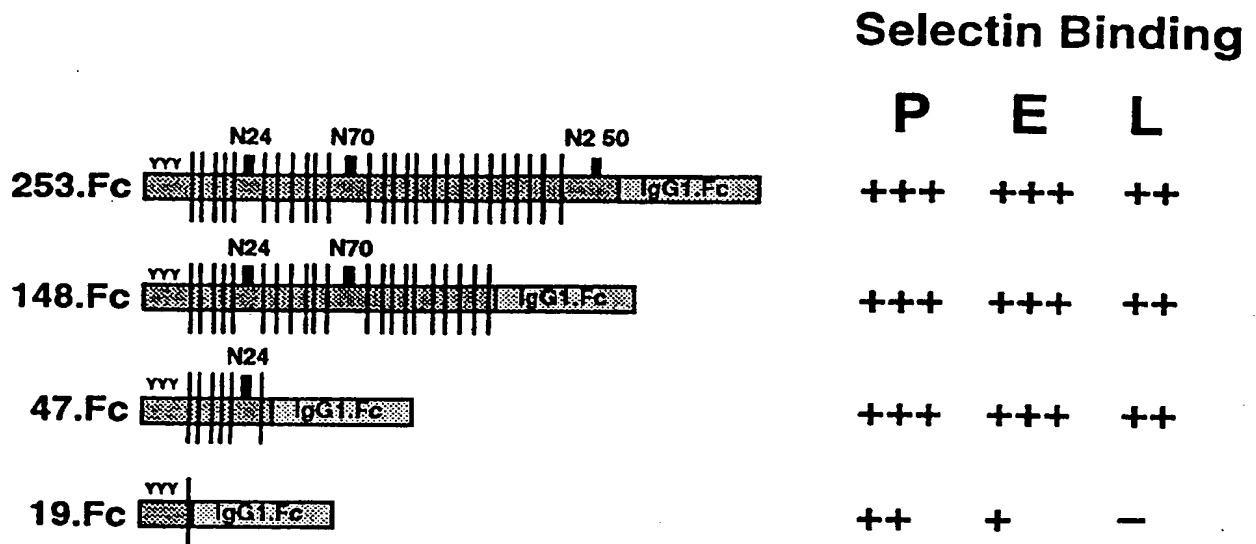


Fig. 13

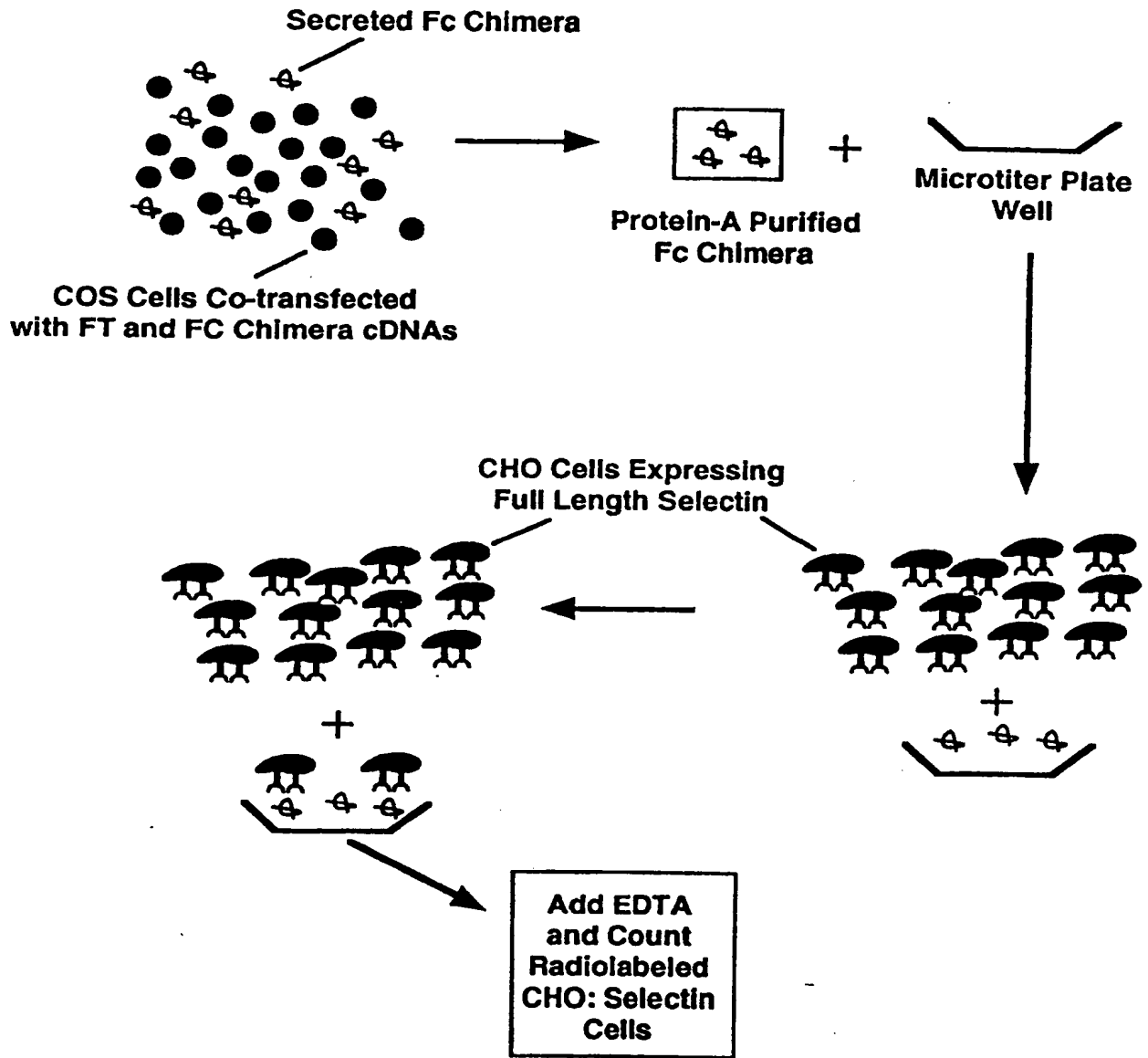


Fig. 14-1

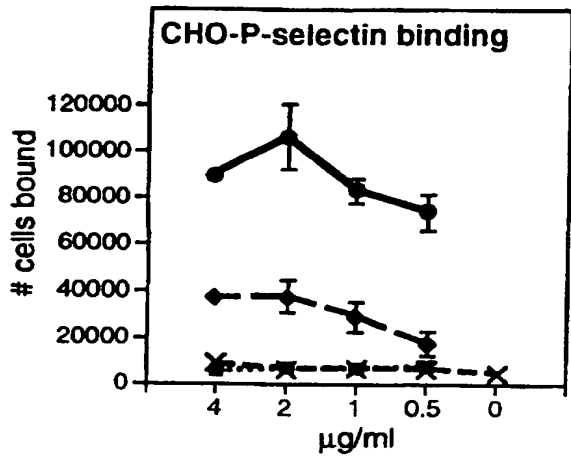


Fig. 14-2

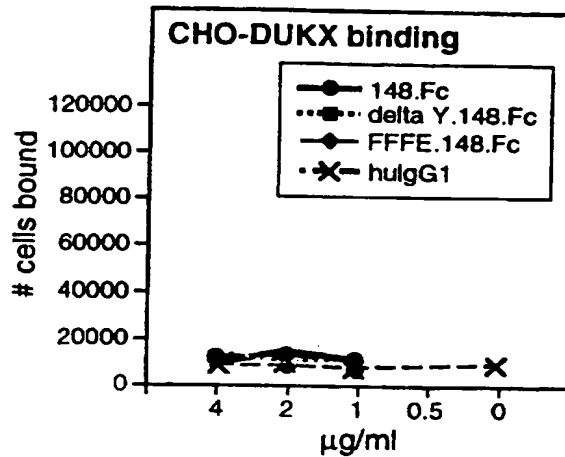






Fig. 16

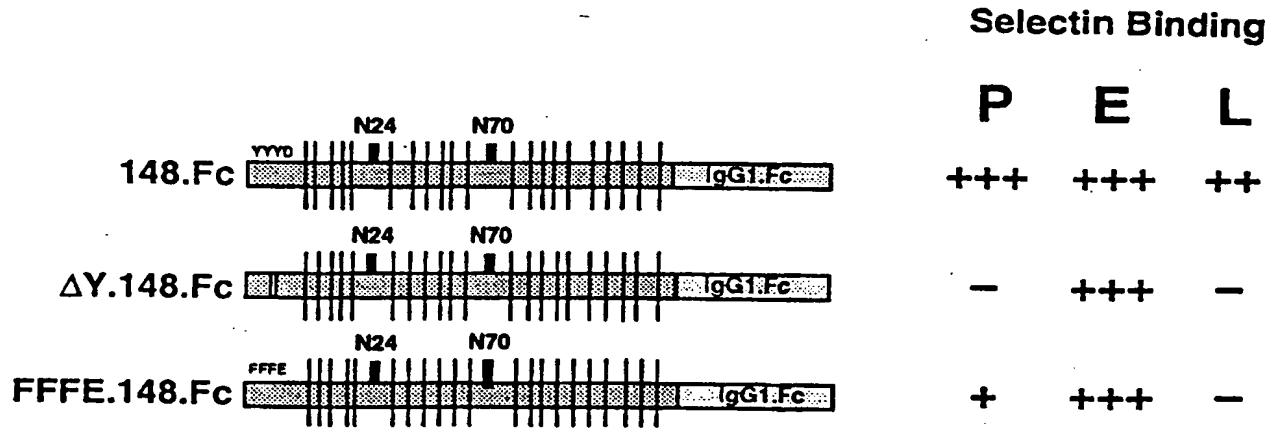


Fig. 17

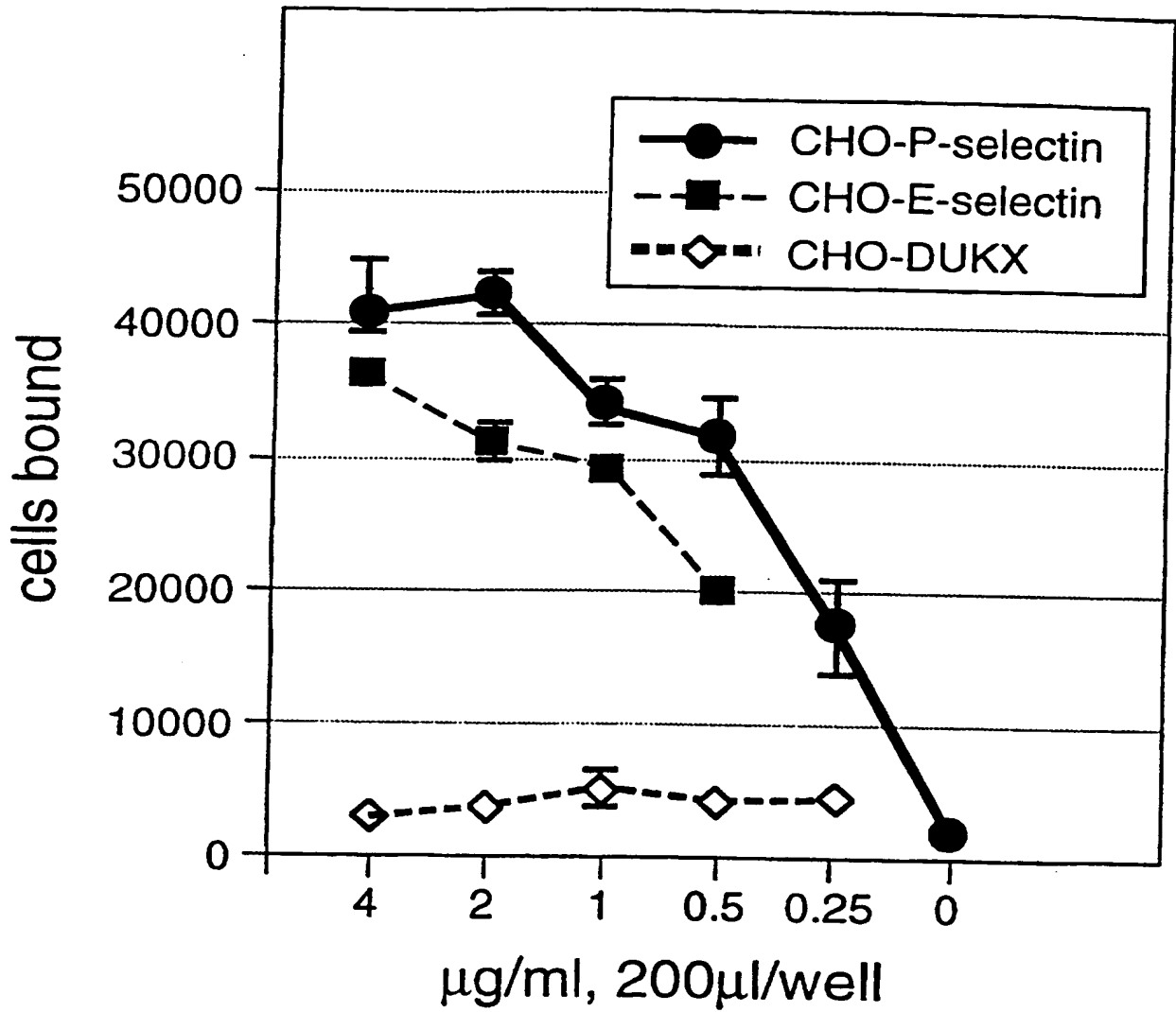




Fig. 19-1

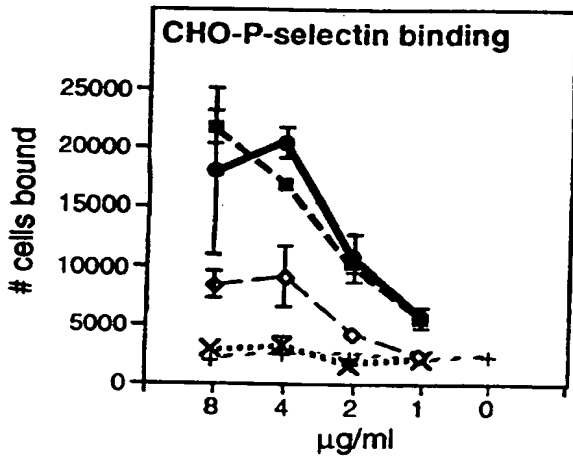


Fig. 19-2

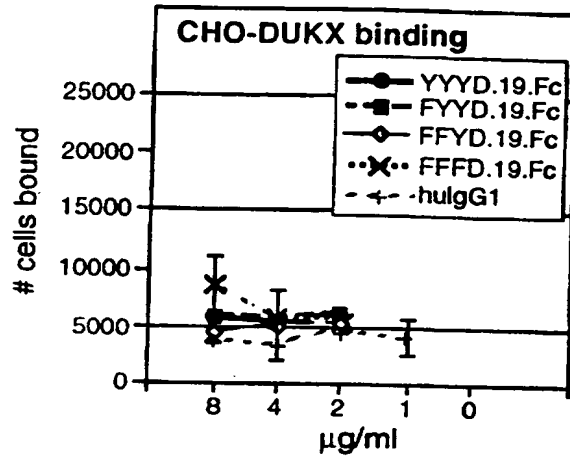


Fig. 20-1

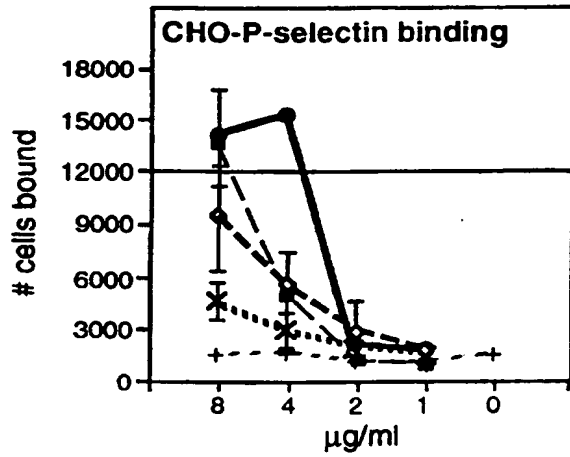


Fig. 20-2

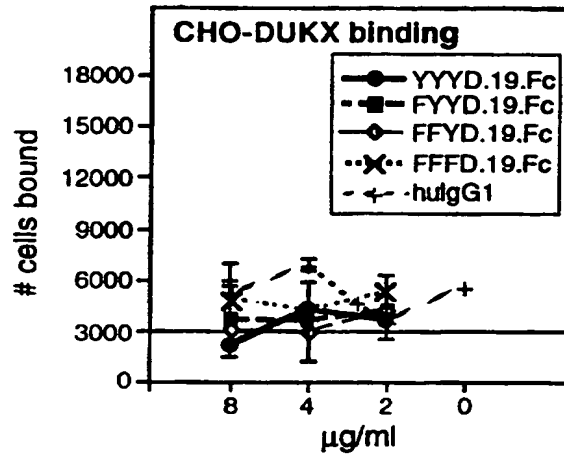


Fig. 21

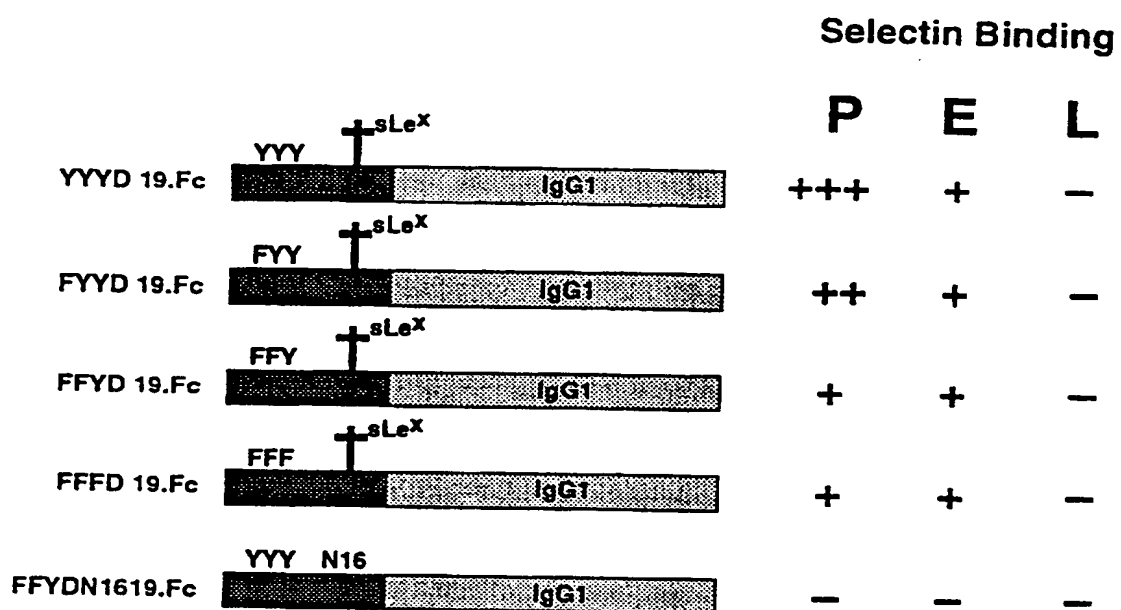


Fig. 22

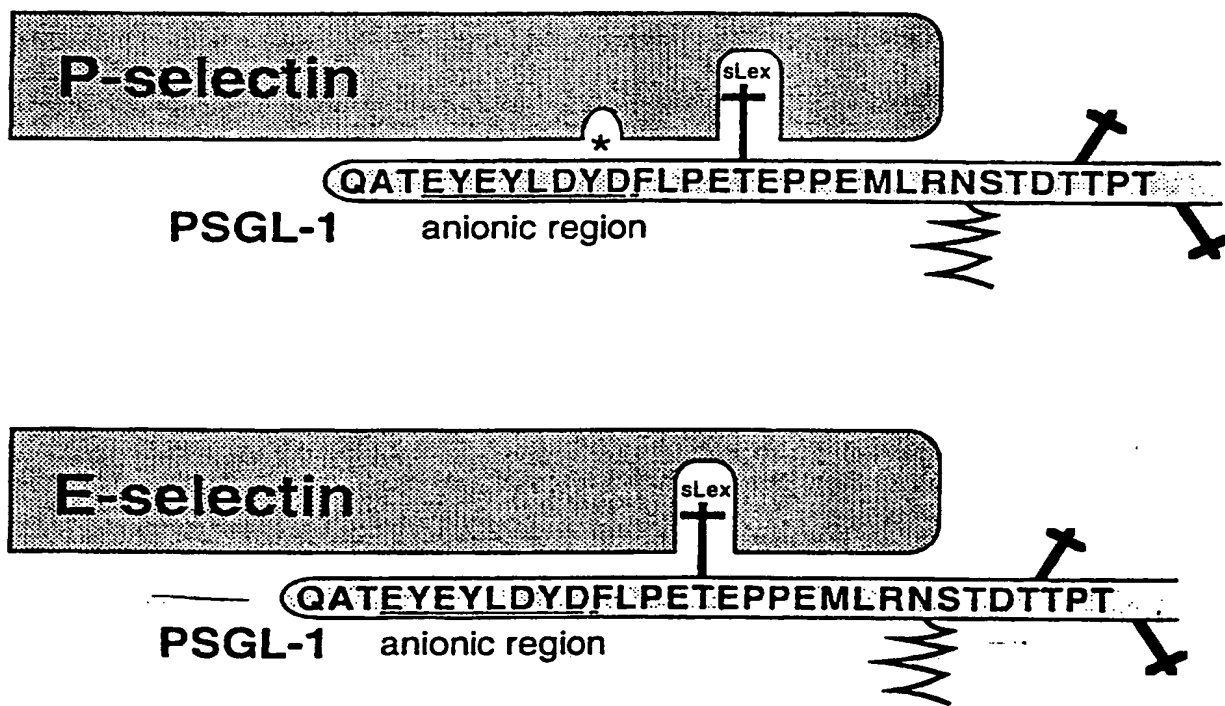


Fig. 23-1

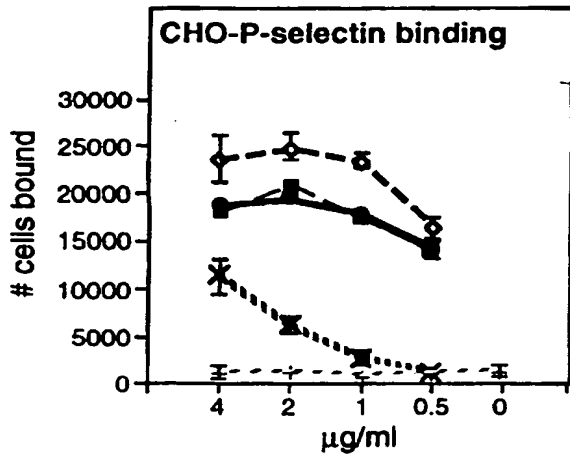


Fig. 23-2

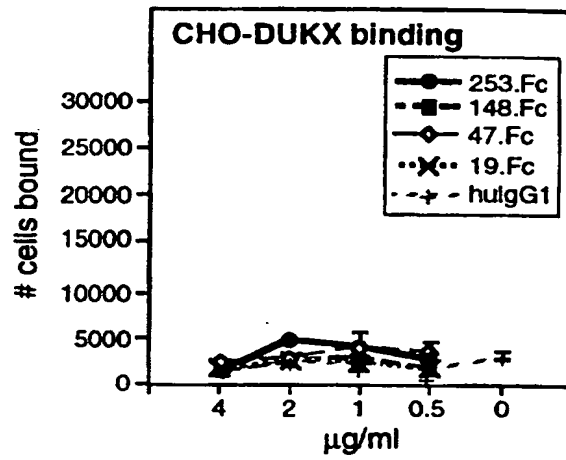




Fig. 24-1

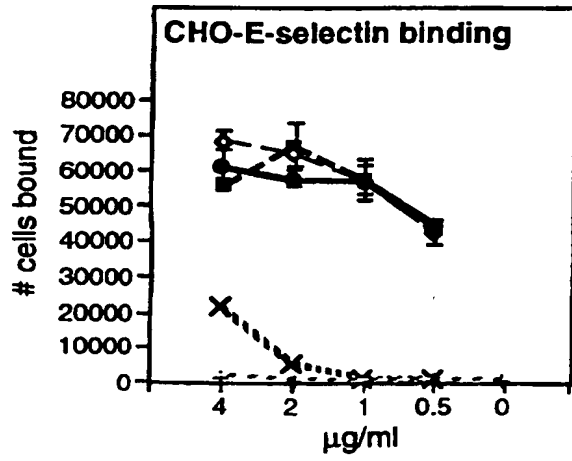


Fig. 24-2

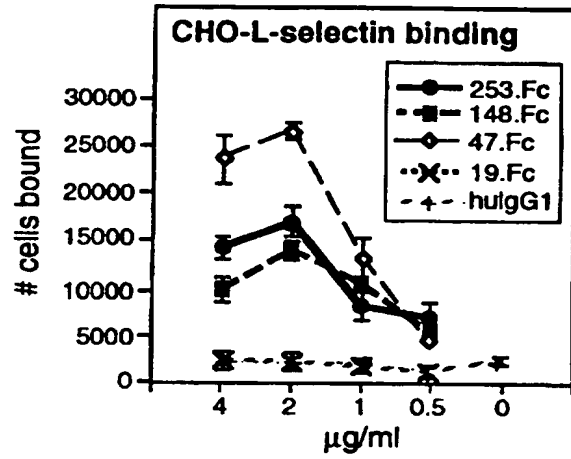
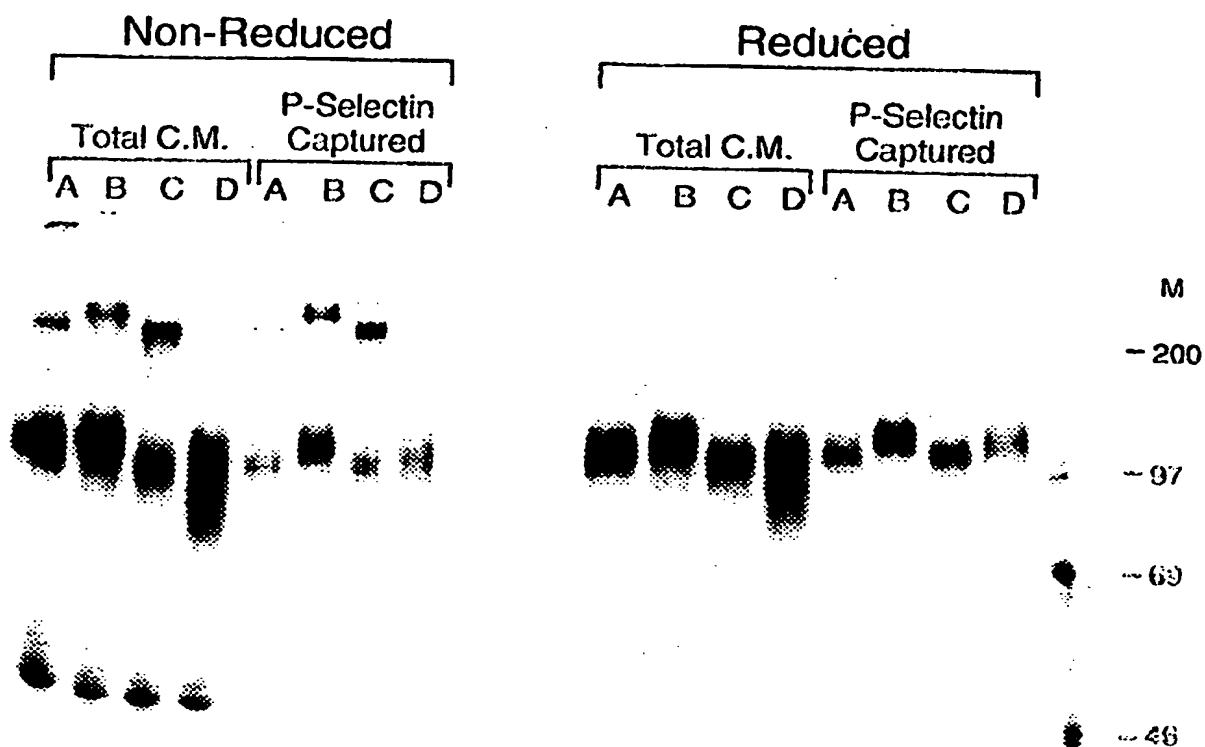


Fig. 25



*Key:*

- A - "17" sPSGL-1
- B - "ΔTM" sPSGL-1
- C - "I316" sPSGL-1
- D - "Qc" sPSGL-1

Fig. 26



Fig. 27

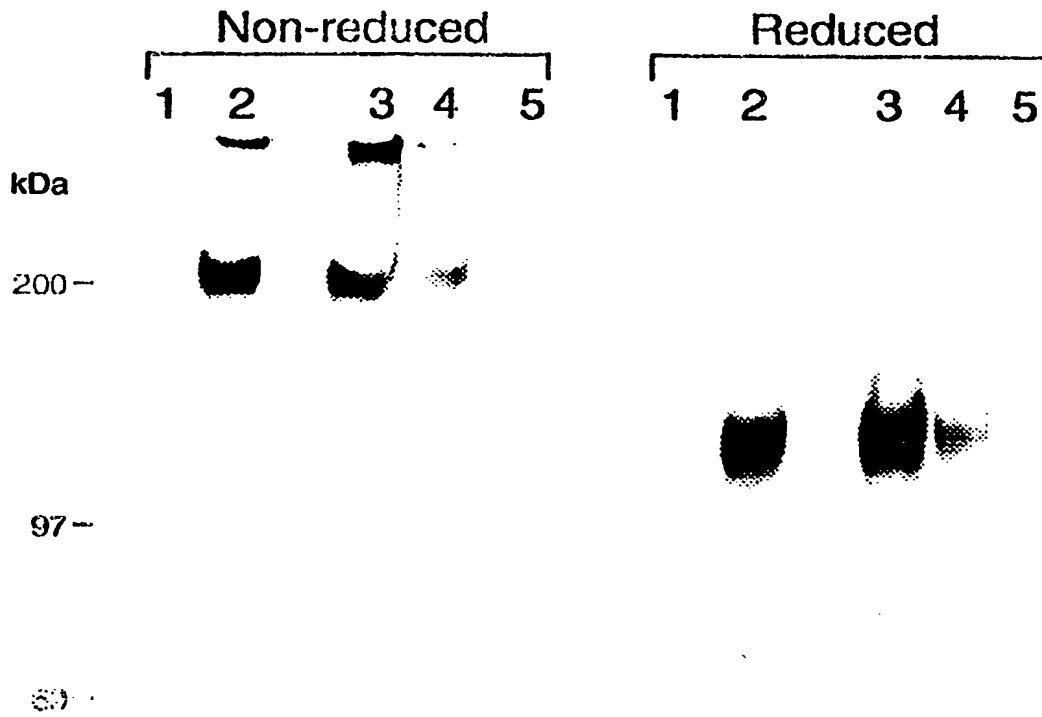


Fig. 28



Fig. 29-1

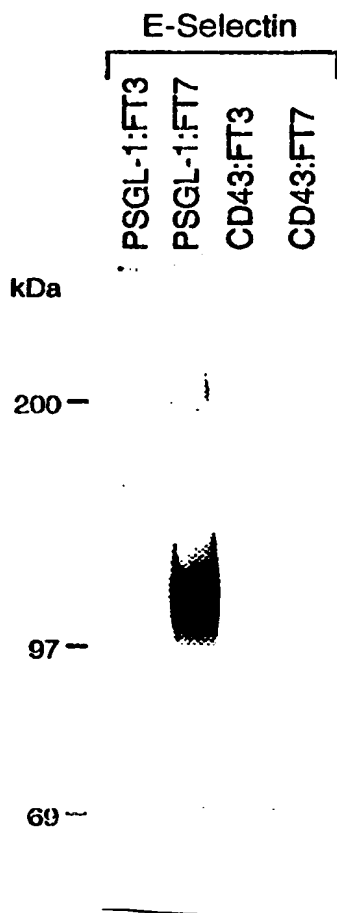


Fig. 29-2

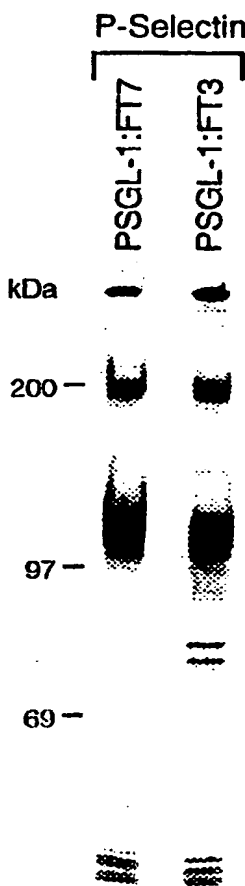


Fig. 29-3

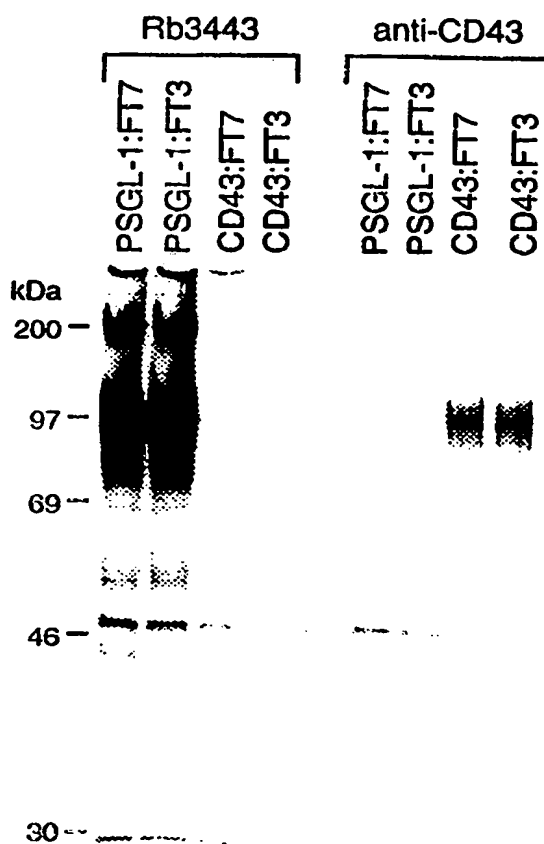


Fig. 30

	<u>E-selectin Inhibition</u>	<u>P-selectin Inhibition</u>
QATEYEYLDYDFLPEC	-	++ (~60μM)
TEYEYLDYDF		++
$\begin{array}{c} \text{PO}_3 \quad \text{PO}_3 \\   \quad   \\ \text{SYLDYS} \end{array}$	+	+++ (~10μM)
$\begin{array}{c} \text{PO}_3 \\   \\ \text{SYLDYS} \end{array}$		+
SYLDYS		-
$\begin{array}{c} \text{PO}_3 \\   \\ \text{SFLDYS} \end{array}$		-
$\begin{array}{c} \text{PO}_3 \quad \text{PO}_3 \\   \quad   \\ \text{AcYLDYNH}_2 \end{array}$		+
$\begin{array}{c} \text{PO}_3 \\   \\ \text{AcLDYNH}_2 \end{array}$		-
$\begin{array}{c} \text{SO}_3 \\   \\ \text{SYLDYS} \end{array}$		-

# INTERNATIONAL SEARCH REPORT

Int. Application No  
PCT/US 97/14159

**A. CLASSIFICATION OF SUBJECT MATTER**

IPC 6 C12N15/12 C12N15/62 C07K14/705 C07K14/51 C07K14/52  
C07K14/54 C07K16/46 C07K19/00

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)  
IPC 6 C12N C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>WO 95 30001 A (GENETICS INST) 9 November 1995 cited in the application</p> <p>see page 15, line 21 - line 21 see page 16, line 22 - line 34; claims 53-55</p>	<p>1-11, 14-16, 29-36, 39-41, 54</p>
X	<p>WO 94 10309 A (GENETICS INST) 11 May 1994 cited in the application</p> <p>see page 8, line 10 - line 11 see page 9, line 35 - page 10, line 9 see page 52, line 28 - page 53, line 3</p>	<p>1-11, 14-16, 29-36, 39-41, 54</p>

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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Date of the actual completion of the international search

Date of mailing of the international search report

17 December 1997

02/01/1998

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Hornig, H



# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 97/14159

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WO 9530001 A	09-11-95	AU 2361395 A	29-11-95
WO 9410309 A	11-05-94	AU 677261 B	17-04-97
		AU 5538894 A	24-05-94
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		JP 8502886 T	02-04-96

